

Physiology and Pharmacology 29 (2025) 284-292

Experimental Research Article



Pancreatic HB9 protein expression is affected by long-term high-fat diet in female rat dams





Roxana Karbaschi¹, Mina Salimi², Homeira Zardooz^{3,4*}

- 1. Faculty of Nursing and Midwifery, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- 2. Traditional Medicine and Hydrotherapy Research Center, Ardabil University of Medical Sciences, Ardabil, Iran
- 3. Department of Physiology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- 4. Neurophysiology Research Center, Institute of Neuroscience and Cognition, Shahid Beheshti University of Medical Sciences, Tehran, Iran

ABSTRACT

Introduction: Maternal high-fat feeding has been identified as a risk factor for metabolic disorders involving abnormal glucose homeostasis and reduced whole-body insulin sensitivity. Recognizing factors like HB9, which play a role in the development of pancreatic β -cells and the release of insulin, is crucial for preventing disruptions in glucose metabolism.

Methods: Twenty female Wistar rats were randomly divided into normal (N) and high-fat (HF) groups, each receiving their specific diets during the pre-pregnancy, pregnancy, and lactation periods (10 weeks). At the end of lactation, the animals' body weight, food consumption, and calorie intake were measured. Additionally, fasting plasma levels of glucose, insulin, and corticosterone were assessed, and the homeostatic model assessment of β -cell function (HOMA- β) was calculated. Pancreatic tissue was collected to evaluate HB9 protein levels, while the adrenal glands were separated and weighed.

Results: The HF group showed significant increases in adrenal gland weight, plasma corticosterone levels, and pancreatic HB9 protein expression compared to the N group. Despite this, there were no significant variations in plasma glucose and insulin concentrations, $HOMA-\beta$ index, or body weight among the study groups. Whereas, the HF group consumed less food

Conclusion: Chronic intake of high-fat foods can act as a psychophysical stressor, triggering the hypothalamic-pituitary-adrenal (HPA) axis and resulting in elevated plasma corticosterone levels in female rat dams. This rise in corticosterone may lead to an upsurge in HB9 protein expression, potentially preventing disturbances in glucose regulation.

Keywords:

Chronic high-fat diet HB9 protein expression Pancreas

Introduction

A long-term consumption of a high-fat diet, especially during pregnancy and lactation, could be a metabolic health-threatening factor for both mother and her chil-

dren (Gawlińska et al., 2021; Li et al., 2017; Saengnipanthkul et al., 2021). Therefore, considering the globally prevalence of metabolic disorders (Barnett et al., 2022; Fenichel 2017; Gosadi 2016; Ji et al., 2014), the high-

* Corresponding author:Homeira Zardooz, homeira_zardooz@sbmu.ac.ir Received 11 May 2024; Revised from 7 October 2024; Accepted 28 October 2024

Citation: Karbaschi R, Salimi M, Zardooz H. Pancreatic HB9 protein expression is affected by long-term high-fat diet in female rat dams. Physiology and Pharmacology 2025; 29: 284-292. http://dx.doi.org/10.61882/phypha.29.3.284

fat content in modern food sources (Khalil et al., 2023; Wali et al., 2020; Wali et al., 2023), and given that the mother is also highly inclined to eat this type of food during her pregnancy and breastfeeding, it is essential to explore the factors and mechanisms contributing to the prevalence of metabolic disorders linked to a high-fat diet during this crucial time.

Experimental models have demonstrated that maternal high-fat feeding not only affects the mother's metabolic health but also leads to altered glucose metabolism and insulin sensitivity in the offspring, which can persist into adulthood (Li et al., 2017; Mendes-da-Silva et al., 2014). Accordingly, the long-term consumption of a high-fat diet in pregnant (Elsakr et al., 2021; Satokar et al., 2022) and non-pregnant conditions (Mosser et al., 2015) can result in reduced sensitivity to insulin, glucose intolerance, as well as β -cells dysfunction. Evidence suggests that sustained exposure to free fatty acids derived from a high-fat diet can lead to lipotoxic damage in the pancreatic islets, which in turn suppresses the synthesis and release of insulin (Oh et al., 2018; Sharma and Alonso 2014; Ye et al., 2019; Acosta-Montaño and García-González 2018).

It has also been shown that a high-fat diet can affect the activity of the hypothalamic-pituitary-adrenal (HPA) axis and the plasma levels of glucocorticoids. This may lead to changes in the function of beta cells and insulin secretion, as well as alterations in the plasma levels of insulin and glucose (Acosta-Montaño and García-González 2018). In this regard, after a glucose load, a decreased insulin secretion and a lower plasma insulin level may occur, which ultimately causes insufficient maintenance of normal plasma glucose levels.

It is revealed that the high-fat diet could change the expression of HB9, which is a transcription factor encoded by the homeobox gene HLXB9, that predominantly expresses in the pancreas of humans and rodents (Leotta et al., 2014). This gene is essential for the formation, differentiation, and function of pancreatic beta cells (O'Dowd and Stocker 2013; Sadeghimahalli et al., 2021). HB9 regulates insulin secretion by inhibiting two target genes that negatively control insulin expression and secretion, leading to an increase in insulin release and biosynthesis (Sahoo et al., 2015; Shi et al., 2013). In this respect, this is worth noting that the expression of proteins, like HB9, that regulate the function of beta cells and insulin secretion, can be changed by a high-fat

diet for various reasons, including changes in corticosterone (Busceti et al., 2019; Janssen 2022) and free fatty acids (Armoni et al., 2005; Lam et al., 2003) levels.

Considering the earlier discussions regarding the link between maternal high-fat food consumption and the incidence of metabolic disorders, and the curial role of HB9 protein in preserving beta cell function and glucose homeostasis, it is likely that the expression of this protein varies when a high-fat diet is consumed during pre-pregnancy, pregnancy, and lactation periods, potentially impacting glucose homeostasis. Therefore, this study was conducted to investigate this hypothesis and clarify the potential role of HB9 protein in glucose homeostasis under these dietary conditions.

Material and methods

Animals and study design

In a 10-week study, 16 female Wistar rats were randomly split into two groups: Group N, which was fed a normal diet, and Group HF, which was given a high-fat diet. The animals of each group used their respective diets during the pre-pregnancy (4 weeks), pregnancy, and lactation periods. The high-fat food contained 65% by weight of standard pellets mixed with 35% by weight of animal butter (58.2% of energy from fat). In normal food (4.75% of energy from fat), saturated fatty acids make up 18.53% while unsaturated fatty acids constitute 81.37%. Throughout the study, all animals had unrestricted access to food and water and were housed in a controlled environment with a temperature of 22±2°C and a 12-hour light/dark cycle.

At the end of lactation (10-week), the animals' weight, food consumption, and calorie intake were measured. Subsequently, fasting blood samples were taken to determine the concentrations of plasma corticosterone and insulin. The amount of the homeostasis model assessment of β -cell function (HOMA- β) was calculated, using HOMA- β formula [HOMA- β = 20×fasting insulin (μ U/mL)/ fasting glucose (mM) - 3.5] (Izadi et al., 2022), to evaluate the secretory function of beta cells. Following the dissection of the animals, the pancreatic tissue was extracted to measure the level of HB9 protein; moreover, the adrenal glands were isolated and weighed.

The animals' body weight and food consumption were measured using a digital scale (FWE Company, Japan, with a sensitivity of 1 gram) at the end of the 10-week research. Also, the level of kilocalories consumed by

each animal was calculated by multiplying the weight of food consumed by the energy available in each gram of food. The authors ensured ethical considerations in animal studies, and the study was approved by the Ethics Committee of Shahid Beheshti University of Medical Sciences (ethical code IR.SBMU.MSP.REC.1398.173).

Blood sampling

Blood sampling was conducted at the end of the 10-week research period, between 8:00 to 9:00 am, under overnight fasting conditions (16 h). The rats were anesthetized with intraperitoneal injection of sodium pentobarbital (60 mg/kg) (Sigma, USA), then decapitated to collect their trunk blood in a microtube containing heparin (5000 IU/ml, Caspian Tamin, Tehran, Iran) (10 microliters per 1 milliliter of blood). The collected blood was centrifuged at a speed of 3000 rpm for 10 minutes at 4°C, then the plasma was separated and frozen at -80°C.

Assays

Plasma concentrations of insulin and corticosterone were evaluated with the rat insulin ELISA kit (detection limit: 0.07 mg/l) from Mercodia, Sweden, and the corticosterone ELISA kit (detection limit: 1.631 nmo-l/l) from DRG Instruments GmbH, Marburg, Germany, respectively. The concentration of plasma glucose was measured utilizing the glucose oxidase method (detection limit: 5 mg/dl) from Pars Azmoon, Tehran, Iran.

Western blot

Western blotting was employed to assess the levels of pancreatic HB9 protein. After completing the experimental process, the anaesthetized rats in all groups (n=3/group) were decapitated, and their pancreatic tissue was extracted. After being frozen in an Azot tank, the samples were preserved at -80 °C to assess the HB9 protein level. The pancreatic tissues were homogenized in lysis buffer and then centrifuged to eliminate cell debris. The supernatant was then gathered and analyzed for total protein concentration using the Bradford method (Kruger 2009).

The proteins were applied to polyacrylamide gels with 12% sodium dodecyl sulfate (SDS) and subjected to electrophoresis before being transferred to a polyvinylidene fluoride membrane.

Overnight treatment of the membranes at 4°C was conducted using a rabbit polyclonal anti-HB9/HLXB9

antibody (Abcam, Cambridge, MA, Cat. No. ab92606). The following day, the membranes were treated with a secondary antibody, horseradish peroxidase-conjugated goat anti-rabbit IgG from Santa Cruz Biotechnology Inc. (Dallas, TX) for 90 minutes at room temperature. Visualization was achieved using the ECL advance kit from Amersham Bioscience (Piscataway, NJ). Results were quantified by conducting a densitometry scan of the films, and the data analysis was carried out using Image J software (Sadeghimahalli et al., 2021).

Statistical analysis

The data were presented as mean \pm SEM (standard error of mean), and the results were analyzed using GraphPad Prism 6 statistical software. The Kolmogorov-Smirnov test was conducted to determine the normality of data distribution. To compare different diets in the groups, the unpaired t-test was used. The limit of significance of differences was set at P \leq 0.05 in all cases.

Results

The study findings revealed that mothers who ingested high-fat food in HF group, plasma corticosterone levels (P<0.0001) (t_{14} =8.712) (Figure 1a), exhibited higher adrenal gland weight per body weight (P<0.0001) $(t_{14}=5.501)$ (Figure 1b), and pancreatic HB9 protein expression (P<0.05) (t_4 =4.438) (Figure 2) than those in the N group. The HF group showed no variance in body weight (P=0.2970) (t_{14} =1.083) (Figure 3a). Whereas, this group consumed less food (P<0.05) ($t_{14}=2.840$) (Figure 3b), compared to the N group. There was also no variation in calorie intake (P=0.5197) (t₁₄=0.660) (Figure 3c) between groups. Additionally, HF animals did not display significant differences in fasting plasma glucose (P=0.5922) (t₁₄=0.5483) (Figure 4a), insulin levels (P=0.5721) (t_{14} =0.5785) (Figure 4b), or HOMA- β (P=0.7430) (t₁₄=0.3344) (Figure 4c) between the HF animals and the N group.

Discussion

Our results indicated that mothers who consumed high-fat food had higher pancreatic HB9 protein level, plasma corticosterone concentration, along percentage of adrenal gland weight/body weight compared to the N group. According to available reports, the expression of HB9 protein in the islets of Langerhans is critical for the development of the pancreas in humans and rodents,

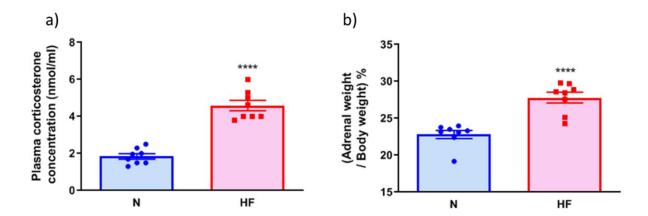


FIGURE 1. The effect of high-fat diet on adrenal gland weight/body weight (%) (a) and plasma corticosterone concentration (b) in dams at the end of lactation. Each column denotes mean \pm SEM (n = 8); N: Normal group, HF: High-fat group; * (P < 0.05) **** (P < 0.0001) showed a significant difference compared to the N group.

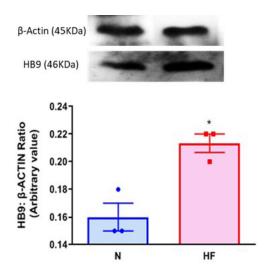


FIGURE 2. The effect of a high-fat diet on pancreatic HB9 protein amount in dams at the end of lactation. Each column denotes mean \pm SEM (n = 3); N: Normal group, HF: High-fat group; *(P<0.05) showed a significant difference compared to the control group.

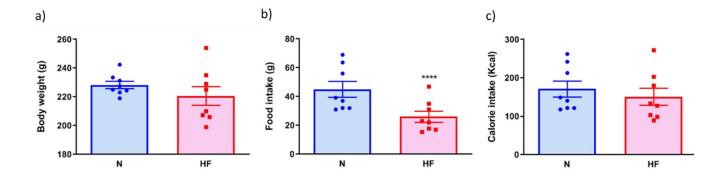


FIGURE 3. The effect of a high-fat diet on body weight (a), food intake (b), and calorie intake (c) in dams at the end of lactation. Each column denotes mean \pm SEM (n = 8); N: Normal group, HF: High-fat group. * (P < 0.05) showed a significant difference compared to the control group.

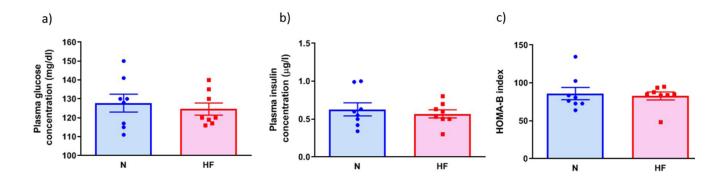


FIGURE 4. The effect of a high-fat diet on plasma glucose (a) and insulin (b) concentrations, also on HOMA-β index (c) in dams at the end of lactation. Each column denotes mean \pm SEM (n = 8); N: Normal group, HF: High-fat group.

resulting in normal beta-cell function. Any change in its expression can disrupt insulin secretion from the islets (Harrison et al., 1999a; Kaneto and Matsuoka 2015). In this regard, investigations on Hlxb9-deficient mice have highlighted the restricted expression of HB9 to beta cells, which was accompanied by small islets of Langerhans with reduced numbers of insulin-producing β-cells and low levels of the glucose transporter Glut2 (Harrison et al., 1999b). Studies on diet-induced obese C57BL/6J mice have shown alterations in pancreatic islet structure, metabolism, and gene expression when exposed to a HF diet. These alterations include changes in islet morphology, insulin secretion, and gene expression profiles in high-fat-fed mice compared to those on a normal diet (Roat et al., 2014). Reports on the effects of a high-fat diet on HB9 expression are limited; nonetheless, according to the above-mentioned findings and the results of the present study, there is a possibility that a high-fat diet has an effect on HB9 expression. The presence of free fatty acids in a HF diet and increased plasma corticosterone levels, as occurred in the present study, can both lead to elevated levels of free fatty acids in plasma, which can affect the expression of transcription factors, including HB9. Regarding the fatty acids in the high-fat diet could alter the corticosterone circadian rhythm (Teeple et al., 2023), which may lead in turn to changes in gene expression (Paula et al., 2020), the increase in HPA axis activity is likely linked to modifications in HB9 protein expression. That might be accomplished by means of epigenetic modifications (Beaudry and Riddell 2012; Sadeghimahalli et al., 2021). We did not analyze the corticosterone circadian cycle, but the plasma corticosterone level among HF group animals

increased significantly, which might be due to the influence of high-fat food as a psychophysical stressor (Benite-Ribeiro et al., 2015; Desai et al., 2014; Namvar et al., 2016). Thus, the heightened corticosterone levels in the HF group might have resulted in enhanced HB9 protein expression in pancreatic tissue (Fine et al., 2018; Udagawa et al., 2023).

In this study, the HF animals exhibited no significant differences in fasting plasma glucose and insulin levels, nor in HOMA-β when compared to the N group. On the other hand, it has been reported that a high-fat diet, rich in saturated fats, can lead to increased plasma glucose levels and decreased insulin sensitivity, which possibly compromises the body's ability to respond to insulin and raises the risk of type 2 diabetes as a consequence (von Frankenberg et al., 2017). It is noteworthy that the elevated levels of glucocorticoids can inhibit insulin secretion in a concentration-dependent manner. On the other hand, as they participate in tissue metabolism and stimulate lipolysis and protein degradation, they can provide the necessary substrates for insulin secretion by enhancing the levels of FFAs and amino acids (Lambillotte et al., 1997). Thus, in the current study, the dual role of glucocorticoids in insulin secretion could have contributed to the lack of change in insulin plasma levels observed in the animals of the HF group. Additionally, it is possible that the elevated HB9 expression observed in the HF group of the current study plays a role in maintaining beta cell function, which helps to stabilize insulin plasma levels and prevent a rise in glucose plasma concentration (Harrison et al., 1999a; Kaneto and Matsuoka 2015).

Although the high-fat diet of this study was high in

calories, the animals fed a high-fat diet did not exhibit a significant difference in body weight. However, they did show a reduction in food consumption compared to the control group, which aligns with the results of the Namvar study (Namvar et al., 2016). In contrast, several studies have reported different results regarding the effect of a high-fat diet on food and calorie intake as well as body weight (Bhandari et al., 2011; Lasker et al., 2019; Marques et al., 2016). Variations in the results observed in distinct studies could be a result of either the duration or the type of high-fat food consumed. The similarity in calorie intake between the HF group animals and the control group could be the result of lower food consumption observed in the HF group. It seems that a self-regulation mechanism has occurred to maintain calorie intake and body weight in the HF group.

In this regard, some evidence indicates that elevated corticosterone levels may decrease food intake by modifying the gene expression of orexigenic (like ghrelin) or anorexigenic (such as proopiomelanocortin) peptides in the hypothalamus (Ans et al., 2018). This relationship is intricate and can be affected by both the fat content and the length of the diet (de Moura e Dias et al., 2021; Higa et al., 2014; Wang et al., 2020). As a result, other studies have also observed contrasting outcomes. For example, in a study, the animals who fed a cafeteria diet showed a disrupted self-regulation mechanism, which led to an increase in food consumption (Higa et al., 2014). Moreover, some studies have reported that oxytocin mRNA expression in the hypothalamus increases following high-fat diet consumption, which could be involved in reducing food intake in the HF group (Manti et al., 2018; Sullivan et al., 2012).

Conclusion

Long-term consumption of high-fat foods before and during pregnancy, as well as during lactation, can stimulate the HPA axis and elevate corticosterone levels in the dams. This increase may promote HB9 protein expression, which could help prevent disruptions in glucose regulation. Nonetheless, continued adherence to this dietary habit threatens glucose homeostasis, emphasizing the importance of conducting more research.

Limitations and future suggestions

Considering the limitations of this study, the following suggestions are proposed: Evaluating the hypothalamic

factors that influence anorexia and orexia affecting the food intake. Analyzing plasma and pancreatic fatty acid levels. Exploring potential epigenetic modifications of the HB9 gene. Performing HB9-specific knockout or inhibitor experiments to clarify the role of this protein in regulating glucose metabolism.

Acknowledgements

We extend our sincere gratitude to all project participants. Approval and support for this project have been provided by the Department of Physiology, School of Medicine, Shahid Beheshti University of Medical Sciences in Tehran, Iran.

Conflict of interest

The authors affirm that they have no competing interests.

Ethics approval

This article was reviewed by the Department of Physiology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. The authors upheld ethical standards in animal research, and the study received approval from the Ethics Committee of Shahid Beheshti University of Medical Sciences under ethical code IR.SBMU.MSP.REC.1398.173.

References

Acosta-Montaño P, García-González V. Effects of dietary fatty acids in pancreatic beta cell metabolism, implications in homeostasis. Nutrients 2018; 10: 393. https://doi.org/10.3390/nu10040393

Ans A H, Anjum I, Satija V, Inayat A, Asghar Z, Akram I, et al. Neurohormonal regulation of appetite and its relationship with stress: A mini literature review. Cureus 2018; 10. https://doi.org/10.7759/cureus.3032

Armoni M, Harel C, Bar-Yoseph F, Milo S, Karnieli E. Free fatty acids repress the GLUT4 gene expression in cardiac muscle via novel response elements. Journal of Biological Chemistry 2005; 280: 34786-34795. https://doi.org/10.1074/jbc.M502740200

Barnett A, Martino E, Knibbs L D, Shaw J E, Dunstan D W, Magliano D J, et al. The neighbourhood environment and profiles of the metabolic syndrome. Environ Health 2022; 21: 1-18. https://doi.org/10.1186/s12940-022-00894-4

Beaudry J L, Riddell M C. Effects of glucocorticoids and exercise on pancreatic β-cell function and diabetes develop-

- ment. Diabetes/metabolism research and reviews 2012; 28: 560-573. https://doi.org/10.1002/dmrr.2310
- Benite-Ribeiro S A, Santos J M, Duarte J A R. Does high fat diet have the stress-like effect on animals? Diabetes & Clinical Diagnosis. 2015. https://doi.org/10.15344/2394-1499/2015/112
- Bhandari U, Kumar V, Khanna N, Panda B P. The effect of high-fat diet-induced obesity on cardiovascular toxicity in Wistar albino rats. Hum Exp Toxicol 2011; 30: 1313-1321. https://doi.org/10.1177/0960327110389499
- Busceti C L, Ferese R, Bucci D, Ryskalin L, Gambardella S, Madonna M, et al. Corticosterone upregulates gene and protein expression of catecholamine markers in organotypic brainstem cultures. Int J Mol Sci 2019; 20: 2901. https://doi.org/10.3390/ijms20122901
- de Moura e Dias M, Dos Reis S A, da Conceição L L, Sediyama C M N d O, Pereira S S, de Oliveira L L, et al. Diet-induced obesity in animal models: points to consider and influence on metabolic markers. Diabetol Metab Syndr 2021; 13: 1-14. https://doi.org/10.1186/s13098-021-00647-2
- Desai M, Jellyman J K, Han G, Beall M, Lane R H, Ross M G. Maternal obesity and high-fat diet program offspring metabolic syndrome. Am J Obstet Gynecol 2014; 211: 237. e1-237. e13. https://doi.org/10.1016/j.ajog.2014.03.025
- Elsakr J M, Zhao S K, Ricciardi V, Dean T A, Takahashi D L, Sullivan E, et al. Western-style diet consumption impairs maternal insulin sensitivity and glucose metabolism during pregnancy in a Japanese macaque model. Scientific Reports 2021; 11: 12977. https://doi.org/10.1038/s41598-021-92464-w
- Fenichel P. Lifestyle and environmental factors in metabolic diseases; endocrine disruptors: new diabetogens? 19th European Congress of Endocrinology 2017; 49. https://doi.org/10.1530/endoabs.49.S21.1
- Fine N H, Doig C L, Elhassan Y S, Vierra N C, Marchetti P, Bugliani M, et al. Glucocorticoids reprogram β-cell signaling to preserve insulin secretion. Diabetes 2018; 67: 278-290. https://doi.org/10.2337/db16-1356
- Gawlińska K, Gawliński D, Filip M, Przegaliński E. Relationship of maternal high-fat diet during pregnancy and lactation to offspring health. Nutrition Reviews 2021; 79: 709-725. https://doi.org/10.1093/nutrit/nuaa020
- Gosadi I M . Assessment of the environmental and genetic factors influencing prevalence of metabolic syndrome in Saudi Arabia. Saudi Med J 2016; 37: 12. https://doi.org/10.15537/smi.2016.1.12675
- Harrison K A, Thaler J, Pfaff S L, Gu H, Kehrl J H. Pancreas

- dorsal lobe agenesis and abnormal islets of Langerhans in Hlxb9-deficient mice. Nat Genet 1999a; 23: 71-75. https://doi.org/10.1038/12674
- Harrison K A, Thaler J, Pfaff S L, Gu H, Kehrl J H. Pancreas dorsal lobe agenesis and abnormal islets of Langerhans in Hlxb9-deficient mice. Nat Genet 1999b; 23: 71-75. https://doi.org/10.1038/12674
- Higa T S, Spinola A V, Fonseca-Alaniz M H, Sant F, Evangelista A. Comparison between cafeteria and high-fat diets in the induction of metabolic dysfunction in mice. International journal of physiology, pathophysiology and pharmacology 2014; 6: 47.
- Izadi M S, Eskandari F, Binayi F, Salimi M, Rashidi F S, Hedayati M, et al. Oxidative and endoplasmic reticulum stress develop adverse metabolic effects due to the high-fat high-fructose diet consumption from birth to young adult-hood. Life Sci 2022; 309: 120924. https://doi.org/10.1016/j. lfs.2022.120924
- Janssen J A J. New insights into the role of insulin and hypothalamic-pituitary-adrenal (hpa) axis in the metabolic syndrome. Int J Mol Sci 2022; 23: 8178. https://doi.org/10.3390/ijms23158178
- Ji F, Ning F, Duan H, Kaprio J, Zhang D, Zhang D, et al. Genetic and environmental influences on cardiovascular disease risk factors: a study of Chinese twin children and adolescents. Twin Research and Human Genetics 2014; 17: 72-79. https://doi.org/10.1017/thg.2014.5
- Kaneto H, Matsuoka T-a. Role of pancreatic transcription factors in maintenance of mature β-cell function. Int J Mol Sci 2015; 16: 6281-6297. https://doi.org/10.3390/ijms16036281
- Khalil W J, Akeblersane M, Khan A S, Moin A S M, Butler A E J. Environmental pollution and the risk of developing metabolic disorders: Obesity and diabetes. Int J Mol Sci 2023; 24: 8870. https://doi.org/10.3390/ijms24108870
- Kruger N J. The Bradford method for protein quantitation. The protein protocols handbook 2009: 17-24. https://doi.org/10.1007/978-1-59745-198-7 4
- Lam T K, Carpentier A, Lewis G F, van de Werve G, Fantus I G, Giacca A. Mechanisms of the free fatty acid-induced increase in hepatic glucose production. Am J Physiol Endocrinol Metab 2003; 284: E863-E873. https://doi.org/10.1152/ajpendo.00033.2003
- Lambillotte C, Gilon P, Henquin J-C. Direct glucocorticoid inhibition of insulin secretion. An in vitro study of dexamethasone effects in mouse islets. J Clin Invest 1997; 99: 414-423. https://doi.org/10.1172/JCI119175

- Lasker S, Rahman M M, Parvez F, Zamila M, Miah P, Nahar K, et al. High-fat diet-induced metabolic syndrome and oxidative stress in obese rats are ameliorated by yogurt supplementation. Scientific Reports 2019; 9: 20026. https://doi.org/10.1038/s41598-019-56538-0
- Leotta C G, Federico C, Brundo M V, Tosi S, Saccone S. HLXB9 gene expression, and nuclear location during in vitro neuronal differentiation in the SK-N-BE neuroblastoma cell line. PLoS One 2014; 9: e105481. https://doi.org/10.1371/journal.pone.0105481
- Li S-W, Yu H-R, Sheen J-M, Tiao M-M, Tain Y-L, Lin I-C, et al. A maternal high-fat diet during pregnancy and lactation, in addition to a postnatal high-fat diet, leads to metabolic syndrome with spatial learning and memory deficits: beneficial effects of resveratrol. Oncotarget 2017; 8: 111998. https://doi.org/10.18632/oncotarget.22960
- Manti M, Fornes R, Qi X, Folmerz E, Lindén Hirschberg A, de Castro Barbosa T, et al. Maternal androgen excess and obesity induce sexually dimorphic anxiety-like behavior in the offspring. The FASEB Journal 2018; 32: 4158-4171. https://doi.org/10.1096/fj.201701263RR
- Marques C, Meireles M, Norberto S, Leite J, Freitas J, Pestana D, et al. High-fat diet-induced obesity Rat model: a comparison between Wistar and Sprague-Dawley Rat. Adipocyte 2016; 5: 11-21. https://doi.org/10.1080/21623945.2 015.1061723
- Mendes-da-Silva C, Giriko C Á, Mennitti L V, Hosoume L F, Souto T d S, Silva A V d. Maternal high-fat diet during pregnancy or lactation changes the somatic and neurological development of the offspring. Arquivos de neuro-psiquiatria 2014; 72: 136-144. https://doi.org/10.1590/0004-282X20130220
- Mosser R E, Maulis M F, Moullé V S, Dunn J C, Carboneau B A, Arasi K, et al. High-fat diet-induced β-cell proliferation occurs prior to insulin resistance in C57Bl/6J male mice. Am J Physiol Endocrinol Metab 2015; 308: E573-E582. https://doi.org/10.1152/ajpendo.00460.2014
- Namvar S, Gyte A, Denn M, Leighton B, Piggins H D, Integrative, Physiology C. Dietary fat and corticosterone levels are contributing factors to meal anticipation. Am J Physiol Regul Integr Comp Physiol 2016; 310: R711-R723. https://doi.org/10.1152/ajpregu.00308.2015
- O'Dowd J F, Stocker C J. Endocrine pancreatic development: impact of obesity and diet. Front Physiol 2013; 4: 170. https://doi.org/10.3389/fphys.2013.00170
- Oh Y S, Bae G D, Baek D J, Park E-Y, Jun H-S. Fatty acid-induced lipotoxicity in pancreatic beta-cells during develop-

- ment of type 2 diabetes. Frontiers in endocrinology 2018; 9: 384. https://doi.org/10.3389/fendo.2018.00384
- Paula A B R, de Coutinho Miranda D, Nogueira F T, de Lauro Castrucci A M, Isoldi M C. Does a high-fat diet affect the circadian clock, or is it the other way around? A systematic review. Nutrition Research 2020; 84: 1-13. https://doi.org/10.1016/j.nutres.2020.10.003
- Roat R, Rao V, Doliba N M, Matschinsky F M, Tobias J W, Garcia E, et al. Alterations of pancreatic islet structure, metabolism and gene expression in diet-induced obese C57BL/6J mice. PLoS One 2014; 9: e86815. https://doi.org/10.1371/journal.pone.0086815
- Sadeghimahalli F, Karbaschi R, Salimi M, Khodagholi F, Zardooz H J. Biochemistry. Pancreatic HB9 protein level is affected by early life stress in young adult rats: possible involvement of TNF-α and corticosterone. Arch Physiol Biochem 2021; 127: 406-413. https://doi.org/10.1080/138 13455.2019.1645699
- Saengnipanthkul S, Noh H L, Friedline R H, Suk S, Choi S, Acosta N K, et al. Maternal exposure to high-fat diet during pregnancy and lactation predisposes normal weight offspring mice to develop hepatic inflammation and insulin resistance. Physiological Reports 2021; 9: e14811. https:// doi.org/10.14814/phy2.14811
- Sahoo K, Sahoo B, Choudhury A K, Sofi N Y, Kumar R, Bhadoria A S. Childhood obesity: causes and consequences. J Family Med Prim Care 2015; 4: 187. https://doi. org/10.4103/2249-4863.154628
- Satokar V V, Vickers M H, Reynolds C M, Ponnampalam A P, Firth E C, Garg M L, et al. Fish oil supplementation of rats fed a high fat diet during pregnancy improves offspring insulin sensitivity. Front Nutr 2022; 9: 968443. https://doi.org/10.3389/fnut.2022.968443
- Sharma R B, Alonso L C. Lipotoxicity in the pancreatic beta cell: not just survival and function, but proliferation as well? Current diabetes reports 2014; 14: 1-9. https://doi.org/10.1007/s11892-014-0492-2
- Shi K, Parekh V I, Roy S, Desai S S, Agarwal S K. The embryonic transcription factor Hlxb9 is a menin interacting partner that controls pancreatic β-cell proliferation and the expression of insulin regulators. Endocr Relat Cancer 2013; 20: 111. https://doi.org/10.1530/ERC-12-0077
- Sullivan E L, Nousen E K, Chamlou K A, Grove K L. The impact of maternal high-fat diet consumption on neural development and behavior of offspring. International journal of obesity supplements 2012; 2: S7-S13. https://doi.org/10.1038/ijosup.2012.15

- Teeple K, Rajput P, Gonzalez M, Han-Hallett Y, Fernández-Juricic E, Casey T. High fat diet induces obesity, alters eating pattern and disrupts corticosterone circadian rhythms in female ICR mice. PLoS One 2023; 18: e0279209. https://doi.org/10.1371/journal.pone.0279209
- Udagawa H, Funahashi N, Nishimura W, Uebanso T, Kawaguchi M, Asahi R, et al. Glucocorticoid receptor-NECAB1 axis can negatively regulate insulin secretion in pancreatic β-cells. Scientific Reports 2023; 13: 17958. https://doi.org/10.1038/s41598-023-44324-y
- von Frankenberg A D, Marina A, Song X, Callahan H S, Kratz M, Utzschneider K M. A high-fat, high-saturated fat diet decreases insulin sensitivity without changing intra-abdominal fat in weight-stable overweight and obese adults. Eur J Nutr 2017; 56: 431-443. https://doi.org/10.1007/s00394-015-1108-6
- Wali J A, Jarzebska N, Raubenheimer D, Simpson S J, Rodionov R N, O'Sullivan J F. Cardio-metabolic effects of

- high-fat diets and their underlying mechanisms-A narrative review. Nutrients 2020; 12: 1505. https://doi.org/10.3390/nu12051505
- Wali J A, Ni D, Facey H J, Dodgson T, Pulpitel T J, Senior A M, et al. Determining the metabolic effects of dietary fat, sugars and fat-sugar interaction using nutritional geometry in a dietary challenge study with male mice. Nat Commun 2023; 14: 4409. https://doi.org/10.1038/s41467-023-40039-w
- Wang L, Xu F, Song Z, Han D, Zhang J, Chen L, et al. A high fat diet with a high C18: 0/C16: 0 ratio induced worse metabolic and transcriptomic profiles in C57BL/6 mice. Lipids Health Dis 2020; 19: 1-13. https://doi.org/10.1186/s12944-020-01346-z
- Ye R, Onodera T, Scherer P E. Lipotoxicity and β cell maintenance in obesity and type 2 diabetes. J Endocr Soc 2019; 3: 617-631. https://doi.org/10.1210/js.2018-00372