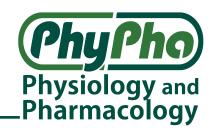


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Original Article



The blockade of D₁/D₂-like dopamine receptors in the lateral periaqueductal gray region affects morphine self-administration with and without exercise in rats





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ABSTRACT

Introduction: The periaqueductal gray (PAG) region plays an essential role in the modulation of nociception. Also, lateral PAG (IPAG) is involved in reward circuitry by the dopaminergic system in addiction. The present study investigated the blockade of D_1/D_2 -like dopamine receptors in the lateral PAG region affects morphine self-administration with and without exercise.

Methods: Rats were divided into six groups. The rats were initially trained to receive small pellets of food by pressing an active lever in the self administration apparatus. Exercise groups were run on a treadmill at 20m/min, 5 days/week, for 4 weeks before the surgery. Then rats were bilaterally implanted with cannulae in IPAG. The SCH23390 and sulpiride were microinjected into the IPAG, 5min before receiving morphine. Afterward, the animals were allowed to self administer morphine in 2h sessions over 11 consecutive days. At last, the numbers of lever pressing, infusion times and withdrawal symptoms were measured.

Results: The results showed the number of active lever pressing was significantly increased in the morphine group compared to other groups in self-infusion during 11 days. Exercise significantly reversed the detrimental effects of morphine self-administration after five days. However, the synergistic effect of injected sulpiride into the IPAG region with exercise training was more pronounced on the amelioration of morphine than on the combinatory effect of SCH23390 with exercise.

Conclusion: The findings suggested that the D₂ dopamine receptor in the IPAG region was involved in the morphine addiction via the dopaminergic system and exercise training in combination with antagonists could reduce the rewarding properties of morphine.

Keywords:

Periaqueductal gray Self-administration Exercise Morphine

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Introduction

Several studies have localized the antinociceptive effects of morphine to periaqueductal gray (PAG) and other midbrain areas and have shown that interference with neural transmission within PAG alters the antinociceptive effects of systemically administered morphine (Meyer et al., 2009). In addition, morphine has been demonstrated to increase dopamine release in several brain areas, including the nucleus accumbens and prefrontal cortex and probably PAG. However, it is unknown whether morphine causes an increase in dopamine release within ventral PAG (Tobaldini et al., 2018). Although opioid-mediated mechanisms in ventral PAG have been explored extensively, much less is known about the role of other neurotransmitters. For example, a subpopulation of dopaminergic neurons within ventral PAG projects either locally or in forebrain regions is known to be involved in endogenous pain modulation (Li et al., 2016). Dopamine D₁-like and D2-like receptors are expressed within ventral PAG (Voulalas et al., 2017).

Indeed, dopamine is the central neurotransmitter in the morphine reward system of the brain (Hyman et al., 2006). Some studies conducted on the addiction process have elucidated the functionality of dopaminergic systems in animal experimental models such as selfadministration and conditioned place preference (Ahmadi et al., 2018; Hosseini et al., 2009b). They found that the family of dopamine receptors was directly involved in the reinforcement properties of drugs (reward behavior) (Lynch et al., 2013; Schetz and Sibley, 2007). Dopamine receptors are classified into two families based on their pharmacological features: the D₁-like receptor including D₁ and closely-related D₅ receptors and the D2-like receptor consisting of D2 and closely-related D₃ and D₄ receptors (Avila-Luna et al., 2015). As presynaptic auto receptors, D₂-like dopamine receptors (D₂Rs) inhibit the dopamine release (Avila-Luna et al., 2015). Accordingly, it is possible that the D, receptor contributes to the induction of pleasure and stimulates the desire of drug consumption in drug abusers (Silva and Heyman, 2001).

Preclinical studies report that physical activity, especially aerobic exercise (in running wheel, treadmill and swimming) decreased the self-administration of morphine, nicotine, cocaine, methamphetamine and heroine (Ahmadi et al., 2018; Charmchi et al., 2016)...

Moreover, aerobic activity produces some alteration in central dopamine and opioid systems (Charmchi et al., 2016; Liu et al., 2019). Furthermore, most researches have demonstrated the effect of exercise on self-administration of drugs in different areas of the brain (Charmchi et al., 2016). Nevertheless, few studies have examined the effects of D_1 and D_2 antagonists on the latera PAG (lPAG) region with and without exercise on morphine addiction. Hence, exercise could be used as a valuable approach to help individuals to recover from addiction (Smith and Lynch, 2011). Therefore, the present study was designed to clarify the blockade of D_1 and D_2 -like receptors modifying morphine addiction (self-administration) via involvement of the lPAG region with and without exercise in rats.

Material and methods

Animals and experimental design

Eighty male Wistar rats obtained from Pasteur Institute, Tehran, Iran, with an initial body weight of 250-300g. The rats were housed under standard laboratory conditions with 12h light/dark cycle (07:00 - 19:00) under controlled temperature (22±2°C) and humidity (50±5%) with available food and water ad libitum. The rats were allowed to adapt themselves to the laboratory condition for at least 1 week before surgery. The Ethics Committee of Animal Use at Isfahan University of Medical Sciences approved the study (IR. MUI.MED.REC. 193083) and all experiments were conducted in compliance with the National Institute of Health Guide for the Care and Use of Laboratory Animals (NIH Publications No. 80-23, revised 2011). In total, eighty rats were used for these experiment that twenty rats were excluded due to misplaced cannulas and sixty reminding rats were randomly selected and divided into six groups (n=10):

1- Morphine (M) group: rats received morphine (5mg/ml) in saline solution during self-administration sessions; 2- D_1 antagonist+ morphine (D_1 +M) group: rats received D_1 antagonist ($4\mu g/0.5\mu l$ in DMSO solution within IPAG), 5min before receiving morphine during self-administration sessions; 3- D_2 antagonist+ morphine (D_2 +M) group: rats received D_2 antagonist ($4\mu g/0.5\mu l$ in saline solution within IPAG), 5min before receiving morphine during self-administration sessions; 4- exercise+ morphine (Ex+M) group: the rats run on the treadmill for 4 weeks and upon a 30min break, they

received morphine during self-administration sessions; 5- exercise+ D_1 antagonist+ morphine (Ex+ D_1 +M) group: rats run on the treadmill for 4 weeks and upon a 30min break, they received D_1 antagonist and morphine during self-administration sessions; 6- exercise+ D_2 antagonist+ Morphine (Ex+ D_2 +M) group: rats run on the treadmill for 4 weeks and upon a 30min break, they received D_2 antagonist and morphine during self-administration sessions.

All animals at the end of experiment (11 days after morphine administration) received naloxone (2mg/kg, SC) and withdrawal signs were recorded for 30min (Figure 1).

Drugs

In the present study, the following drugs were used: morphine sulfate (Temad, Tehran Iran) and SCH-23,390 (Tocris Bioscience, Bristol, UK), a D₁ receptor antagonist, which were dissolved in sterile saline (0.9%). Sulpiride (Tocris Bioscience, Bristol, UK), a D₂ receptor antagonist, was dissolved in 10% DMSO (Sigma-Aldrich, Germany). Naloxone hydrochloride (Sigma Co., USA) was dissolved in saline 0.9%.

Treadmill running

The rats were familiarized with the treadmill running 7 days before the experiments how to run on the treadmill. This also helped use to identify sick or lazy rats, which would be excluded from the experiment (20 male rats). The remaining rats (60 rats) underwent the intravenous cannulation and allowed to recover from the surgery for 7 days. The running period for all the groups consisted of 5–60min of running, five days per week, for 4 weeks on a flat treadmill at a rate of 5–20m/min. After surgery and 7 days of recovery, due to maintaining aerobic capacity the rats run on the treadmill for 15min at a rate of 15m/min. After a 30min break, they were placed in the selfadministration apparatus and continuous experiments were conducted for 11 days (Ahmadi et al., 2018). Indeed, the non-runners groups were put on

the treadmill without running for the same duration as runners (Hoveida et al., 2011).

Surgery (stereotaxic and jugular vein catheterization) The rats were anesthetized by intraperitoneal injection of ketamine (100mg/kg) and xylazine (10mg/kg) (Haghparast et al., 2013). The animals were placed into the stereotaxic device (Stoelting Co., USA). The scalp was retracted and the area of surrounding bergma was cleaned and dried. In addition, lidocaine with epinephrine (0.2ml) was injected in several locations around the incision (Haghparast et al., 2013). One stainless-steel guide cannula (23-gauge, 12mm) was unilaterally implanted 1mm above the intended site of injection (IPAG) according to the rat brain atlas (Paxinos and Watson, 2006). The coordinates for IPAG were determined by the rat brain atlas, anterior/posterior= -7.56mm caudal to bergma, medial/ lateral= +1mm and dorsal/ventral= -5.4mm ventral from the skull surface. The guide cannula was secured in place using two stainless steel screws anchored to the skull. Then, cannula was inserted into the jugular vein.

The cannula was guided subcutaneously up to the skull and connected to polyethylene tubing where it was fixed to a metal tube and secured to the skull with small screws, fixed with dental acrylic cement. After the cement was completely dried and hardened, two stainless steel stylets were used to occlude the guide cannula during the recovery period. Penicillin-G 200,000 IU/ml (0.2–0.3ml/rat, single dose, intramuscular) was administered immediately after surgery. After surgery, the animals were placed in individual home cages and they were allowed to recover from the operation for 7 days before performing experiments (Sadeghzadeh et al., 2015).

Drug administration

Microinjections were performed by lowering a stainless steel injector cannula (30-gauge needle) with a length of 1mm longer than the guide cannula into CA1. The injector cannula was connected to a 1-µl Hamilton syringe by polyethylene tubing (PE-20), then

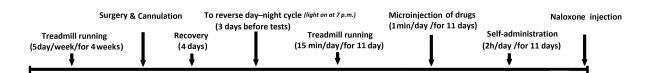


FIGURE 1. Experimental design for all groups.

drug solution or vehicle bilaterally was infused in a total volume of 0.5μ l/rat over 60s and was left for the 60s extra time and followed by replacement of the obturator (Sadeghzadeh et al., 2015).

Self-administration apparatus

The rats were initially trained to press a lever using food as reinforcement to acquire drug selfadministration before being surgically implanted with a chronic intravenous jugular catheter. Training and testing were carried out in standard operant conditioning cages (21×21×28cm) placed in a sound-attenuated room, ventilated with fans, based on the bothers of the previously-used method (Alaei et al., 2005; Sahraei et al., 2004) with minor modifications. The apparatus was equipped with active and passive levers, 2cm above the floor, with a red light located 4cm above the active lever. The intravenous cannula was connected to an infusion pump via a swivel, allowing the animal to move relatively freely (Hosseini et al., 2009a). Pressing of the active lever, marked by a light, resulted in a 10s infusion of 0.1ml fluid through an infusion pump. The fluid was saline in the morphine group (5mg/ml) in the other groups. Depression of the active lever during this time (10s) did not affect the infusion of the drug. Pressing of the passive lever had no programmed consequences. In this study, the number of lever pressing was regarded as a measure of the reinforcing action of the drug (Moradi et al., 2015; Reisi et al., 2014).

Training phase

Ten days before the experiments, the animals were transferred to a special room and the day-night cycle 12/12 hours were reversed into night-day cycle 12/12 hours (light on at 7 pm). The experiments were subsequently performed during the dark phase. Before the surgery, the training program was started after 24h of food restriction. The animals were placed in the selfadministration apparatus where a lever filled with food pellets was available. Each lever pressing resulted in the delivery of a 100mg pellet (Moradi et al., 2015). Each rat was allowed to have self-training until 40 pellets were received. Following acquisition of the lever pressing behavior, the rats were returned to ad libitum food and allowed to gain their weight for 3 days, before the surgery was performed (Hosseini et al., 2009a; Reisi et al., 2014).

Procedure

Seven days after recovery and following 24h of food restriction, the rats were placed in the operant chambers where a lever filled with food pellets was available. Each active lever pressing resulted in the delivery of a 100mg pellet. The jugular cannula of rats was connected to an infusion pump and the animals were placed in the self-administration apparatus for 2h each day on an FR-1 schedule (Hosseini et al., 2009a). The trained animals were allowed to press active and passive levers freely. By pressing the active lever, the rats received 0.1ml of morphine and small pellets in the first 6 days and morphine without pellets in the final 5 days of the experiment. Pressing the passive lever did not deliver fluid or food. In the first 6 days of self-administration period, the availability of food was restricted to reduce body weight by 15%, which has been shown to facilitate initiation of intravenous self-administration (McDougall et al., 2005). Changes of less than 15% in the number of injections in the last 3 days were regarded as baseline. In the next 5 days, the animals had free access to their ad libitum food. The catheters were flushed daily with 0.1ml saline containing heparin sulfate (50U/ml) during the recovery period as well as before and after self-administration sessions. All operant sessions were conducted during the animals' dark cycle. Catheter potency was tested by the injection of 0.1ml of sodium pentobarbital solution (10mg/ml) into the catheter and observation of animal behavior. Animals with patent catheters exhibit prominent signs of anesthesia (loss of muscle tone) a few seconds after the administration (Hosseini et al., 2009a).

Withdrawal syndrome signs

The withdrawal syndrome was precipitated with an intraperitoneal injection of 2.0mg/kg naloxone HCL, dissolved in saline. Withdrawal signs (climbing, jumping, grooming, defecation, teeth chattering, stretch, genital grooming and shaking) were recorded within 30min after the injection of naloxone at the last day of the experiment.

Histological verification

Immediately after the last session, all rats were anesthetized with a high dose of ether and perfused transcardiacally with a phosphate-buffered saline solution (pH= 7.4) (Hasue and Shammah-Lagnado,

2002). The brains were removed and placed in formaldehyde (4%). The brains were removed and kept in a 4% fresh formalin solution for 3 days and sliced coronal in $50\mu m$ sections through cannula placement sections. The neuroanatomical locations of cannula tips were confirmed using the rat brain atlas. The data reported here are only from animals whose placements of cannula tips were histologically verified (Figure 2).

Statistical analysis

Data have been presented as mean \pm SEM. The number of active and passive levers pressings of final 5 days was compared between different groups. The number of passive lever pressing between different groups (summed over ten sessions) was compared using one-way analysis of variance (ANOVA) and Turkey's post hoc test. In addition, the ANOVA test was used to analyze withdrawal signs. Then number of active and passive lever pressings every session and the number of self-infusion between different groups were compared to each other using repeated-measures one-way analysis of variance (ANOVA) and Turkeys' post hoc test. The criterion for statistical significance was P<0.05.

Results

Initiation of drug self-administration

The number of active levers pressing was significantly lower in (D_1 +M and D_2 +M) groups than in the morphine (M) group, showing that D_1 and D_2 antagonists reduced morphine self-administration (P<0.05 to P<0.001 on

all the experimental days; Figure 3). No significant (P>0.05) difference was found between D_1+M and D_2+M groups in the number of reinforcement lever pressing (Figure 3).

No significant reduction was found in the number of self-infusion (SI) between D_1+M and D_2+M groups. The number of self-infusion was significantly lower in D_1+M and D_2+M groups than in the M group during all days (P<0.05 and P<0.001; respectively), indicating that D_1 and D_2 antagonists could decrease the consumption of morphine (Figure 4).

Figure 5 presents the numbers of reinforcement lever pressing in the final 5 days. This number of levers pressing were significantly (P<0.001) lower in the Ex+M group than in the M group, demonstrating that exercise could reduce morphine self-administration. The number of active lever pressing was significantly decreased in Ex+D₁+M and Ex+D₂+M groups than in the Ex+M group (P<0.001 in both of them; Figure 5).

As Figure 5 shows, a significant (P<0.05) difference was found in the number of reinforcement lever pressing in the Ex+D₁+M group than in the Ex+D₂+M group, revealing that D₂ antagonist associated with exercise was more effective than D₁ antagonist associated with exercise in the reduction of morphine self-administration. The number of reinforcement lever pressing was significantly reduced in both Ex+D₁+M and Ex+D₂+M groups compared to the M group (P<0.05 and P<0.001; respectively).

Figure 6 presents the number of self-infusions in the

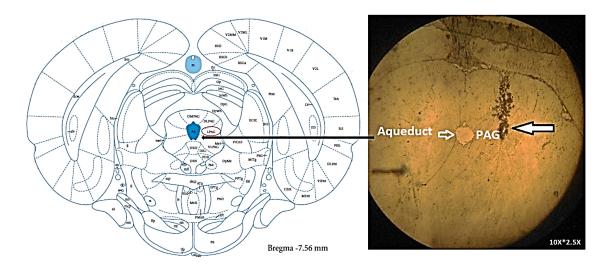


FIGURE 2. The position of the injection cannula tips in the lateral periaqueductal gray (LPAG) regions for all rats included in the statistical analysis. All histological section of brain were $50\mu m$ (10x*2.5x).

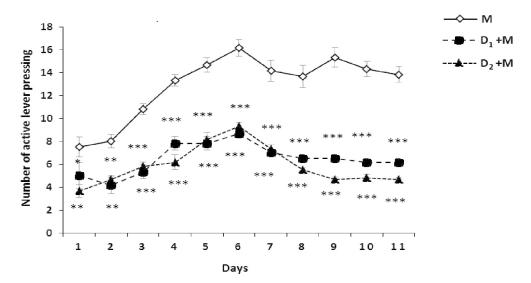


FIGURE 3. Comparison of the number of reinforcement lever pressing between three groups during 11 days (M, D_1+M and D_2+M). The numbers of the RL were compared in different groups using the repeated measure the one-way ANOVA followed by Tukey's post hoc test. Values are expressed as the mean±SEM. No significant difference was observed between D_1+M and D_2+M groups.

M: Morphine group; D_1+M : D_1 antagonist+ Morphine group; D_2+M : D_2 antagonist+ Morphine group; $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$ compared to the M group.

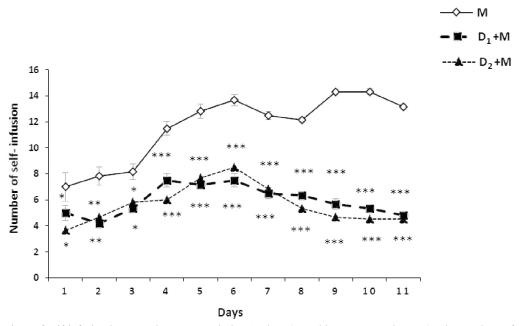
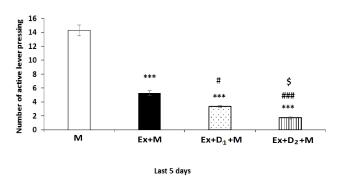
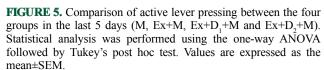


FIGURE 4. Comparison of self-infusion between three groups during 11 days (Morphine, D_1+M and D_2+M). The numbers of self- infusion were compared in different groups using the repeated measure the one-way ANOVA followed by Tukey's post hoc test. Values are expressed as the mean \pm SEM. No significant difference in the number of self-infusion was observed between D_1+M and D_2+M groups. M: Morphine group; D_1+M : D_1 antagonist+ Morphine group; D_2+M : D_2 antagonist+ Morphine group. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$ compared to the M group.

last 5 days of morphine self-administration. Running activity significantly (P<0.001) decreased the number of self-infusions during the past 5 days in the M+E group than in the M group, so that the running activity reduced self-administration of morphine. The number of self-infusion was significantly (P<0.001) decreased in Ex+D₁+M and Ex+D₂+M groups than in the Ex+M group (Figure 6). The number of self-infusion was significantly (P<0.05) lower in the Ex+D₂+M group

than in the $Ex+D_1+M$ group, showing that D_2 antagonist associated with exercise was more effective than D_1 antagonist associated with exercise in the presentation of morphine self-administration. The number of self-infusion in the final 5 days was significantly (P<0.001) lower in both $Ex+D_1+M$ and $Ex+D_2+M$ groups than in the M group (Figure 6). No significant (P>0.05) difference was found in the number of passive lever pressing between all groups during 11 days (Figure 7).





M: Morphine group; Ex+M: Exercise+ Morphine group; Ex+D₁+M: Exercise+ D₁ antagonist+ Morphine group; Ex+D₂+M: Exercise+ D₂ antagonist+ Morphine group.

**** $P \le 0.001$ compared to the M group; $^{\#}P \le 0.05$, $^{\#\#}P \le 0.001$ compared to the Ex+M group; $^{\$}P \le 0.05$ compared to the Ex+D₁+M group.

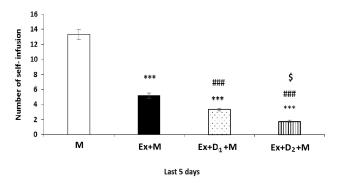


FIGURE 6. Comparison of the number of self-infusion between the four groups in the last 5 days (M, Ex+M, Ex+D₁+M and Ex+D₂+M). Statistical analysis was conducted using the one-way ANOVA followed by Tukey' post hoc test. Values are expressed as the mean±SEM.

M: Morphine group; Ex+M: Exercise+ Morphine group; Ex+D₁+M: Exercise+ D₁ antagonist+ Morphine group; Ex+D₂+M: Exercise+ D₂ antagonist+ Morphine group.

**** $P \le 0.001$ compared to the M group; $^{*}P \le 0.05$, $^{###}P \le 0.001$ compared to the Ex+M group; $^{\$}P \le 0.05$ compared to the Ex+D₁+M group.

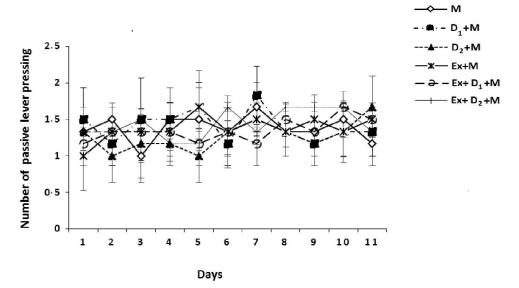


FIGURE 7. Comparison of passive lever pressing between all groups of animals during 11 days (M, Ex+M, Ex+D₁+M and Ex+D₂+M). The numbers of passive lever were compared in different groups using the repeated measure the one- way ANOVA followed by Tukey's post hoc test. Values are expressed as the mean±SEM. No significant difference in the number of passive lever pressing was observed between the groups.

M: Morphine group; D_1+M : D_1 antagonist+ Morphine group; D_2+M : D_2 antagonist+ Morphine group; $Ex+D_1+M$: Exercise+ D_1 antagonist+ Morphine group; $Ex+D_2+M$: Exercise+ D_2 antagonist+ Morphine group.

Withdrawal syndrome signs

Table 1 shows the effects of exercise associated with drugs (morphine, D_1 and D_2 antagonists) on withdrawal syndrome signs. All of the withdrawal signs (except jumping) were significantly (P<0.001) attenuated in the M+Ex group than in the M group. The withdrawal syndrome signs (climbing, grooming, stretch and genital grooming) were significantly lower in the D_2 +M+Ex group than in the D_1 +M group (P<0.05 to P<0.001). Most of the withdrawal signs, especially genital

grooming, jumping and shaking significantly (P<0.05 to P<0.001) were decreased in the D_2 +M+Ex group than in the D_2 +M group.

Discussion

This study showed that the injection blockade of D_1 - and D_2 -like receptors within the lateral periaqueductal gray region reduced morphine self-administration with and without exercise in rats. According to our results, D_1 and D_2 antagonists reduced morphine active lever

IABLE 1: Comparison of morphine withdrawal syndrome signs (including climbing, teeth chattering, jumping, grooming and defecation) between all groups during 30min after naloxone injection (2mg/kg IP) in the last day.

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Group Withdrawal sings	/Climbing Standing	Grooming	Stretch	Defection	Teething	Jumping	Genital grooming	Shaking
Morphine (M)	28±0.93	19.5±0.43	7.17±0.40	3.17±0.17	43.67±1.02	1.5±0.22	2.5±0.34	6.17±0.83
D _i antagonist+Morphine (D _i +M)	14.3±0.98	10±0.73	3.33±0.33	1±0.00	1.76±0.33	1 ±0.00	0.00±0.00	00-0-0
D_2 antagonist+Morphine (D_2+M)	10±0.73	10.3±0.61	1.33±0.21	1±0.00	3.3±0.00	0.67±0.21	2±0.00	1±0.00
Exercise+ Morphine (Ex+M)	8±0.73	5±0.26	2±0.00	1.67±0.21	10±0.00	1±0.00	1.67±0.21	1.33±0.21
Exercise+D ₁ antagonist +Morphine (Ex+D ₁ +M)	6.17±0.47	3.67±0.21 EEESSS ¥¥¥###***	0.83±0.16	1±0.00	2.5±0.22	0.67±0.21	2.17±0.16	0±0.00
Exercise+D ₂ antagonist +Morphine (Ex+D,+M)	5.83±0.82	3.5±0.22 \$\$\$¥¥###***	0.67±0.21	1±0.00	2.5±0.43	0.5±0.22	1.67±0.21	0±0.00

* $P \le 0.05$, *** $P \le 0.001$ compared to the morphine group. * $P \le 0.05$, *** $P \le 0.001$ compared to the D₁+M group. * $P \le 0.05$, *** $P \le 0.001$ compared to the Ex+M group. Statistical analysis was conducted using the one-way ANOVA followed by Tukey's post hoc test. Values are expressed as the mean±SEM.

pressing compared to the morphine group, and the number of self-infusion was significantly decreased in D_1 and D_2 , suggesting the attenuation reinforcement effects of morphine. Furthermore, no significant difference in the number of passive lever pressing between the morphine group and the other groups, proving that the animal behavior was directed to obtain the reward effects of morphine. In agreement with these results, it was demonstrated that morphine induced euphoria through

transmission during sensitization development (Meyer et al., 2009; Smith and Lynch, 2011). In animal studies, some neuro-adaptive changes were also observed in dopamine D_1 and D_2 receptors during sensitization (Łupina et al., 2018).

Previous findings also revealed that analgesia induced

μ opioid receptors activation and increased dopamine

by morphine was mediated by D_1 and D_2 -like dopamine receptors (Nestler, 2001; Tobaldini et al., 2018). The

present results and other researchers suggested that D_1 and D_2 dopamine receptors were involved in morphine-induced analgesia and inhibited behavioral sensitization to morphine through the release of dopamine from dopaminergic neuron by stimulation of these receptors in nucleus accumbens (Charmchi et al., 2016) and blockade of these receptors in IPAG.

According to these connections and the key role of dopamine in drug reward, IPAG could participate in opiate reward. Microinjection of dopamine antagonists into IPAG reduced the reinforcement effects of morphine. Consistent with these findings, other studies reported that PAG dopamine network was involved in the rewarding and sensitizing properties of heroin (Lane et al., 2005), suggesting that repeated opiate drug exposure induces biochemical changes in specific brain regions to mediate the reinforcing actions of drugs, mostly the mesolimbic dopamine system (Flores et al., 2004). Previous studies revealed that the direct injection of dopamine antagonists into the PAG attenuated the ant nociceptive effects of morphine and heroin (Meyer et al., 2007). This dopaminergic modulation of nociception within ventral PAG is likely involved in the development of opioid tolerance and drug seeking. Ventral PAG was known to play an important role in tolerance to opioids (Boecker et al., 2008; Meyer et al., 2009). In addition, the development of opioid mediated reinforcement and sensitization has been shown to be dependent on ventral PAG dopamine (Meyer et al., 2007). In the current study, the microinjection of dopamine receptors antagonists into IPAG also reduced the properties of morphine. Overall, these experiments indicated an interaction between morphine and dopamine within IPAG. The previous research also found that anti-nociception of dopamine was induced by activation of D, receptors not by D₁ (Liu et al., 2016), while the present study could not find significant differences between D₁ or D₂ antagonists in morphine addiction. Others reported that many of the behavioral alterations after using morphine were associated with the activity of D₂-expressing in the brain (Lane et al., 2005; Yoshida et al., 2019).

Previous studies indicated that exercise induced euphoria, especially endurance exercise significantly increased the level of euphoria (Greenwood and Fleshner, 2011). The release of endogenous opioids also occurred in front-limbic brain regions after sustained physical exercise and changes of the central opioid receptor

binding were identified in prefrontal regions after 2h of long-distance running (Greenwood and Fleshner, 2011). Furthermore, treadmill running increased the effect of D, antagonist within IPAG to prevent effects of dopamine on morphine-induced sensitization. Others showed that neurotransmitters and endogenous opioids were evoked in the brain by exercise (Robison et al., 2018), similar to those induced by substances of abuse. For example, the levels of beta-endorphins, dopamine and norepinephrine were increased by exercise and drugs, and abuse of drugs was reduced by combination of exercise and morphine (Smith et al., 2018). Others also reported that disturbances of the brain dopaminergic activity (including low striatal D, receptor levels) induced compulsive behaviors in substance abuse (Lane et al., 2005; Nestler, 2001). Similarly, treadmill running significantly reduced morphine sensitization through D, receptors within IPAG. The evidence shows that exercise induces higher level of D₂R binding in striatum and plays a crucial role in addiction (Volkow and Baler, 2014). In our results, no significant differences were found in D₁ and D₂ dopamine receptors antagonists after injection within IPAG to reduce reinforcement effects of morphine in rats, but aerobic exercise, along with injection of D₂ antagonist increased this reduction into IPAG. In agreement with the results, it has been demonstrated that resistance exercise reduces heroin self-administration via the down-regulation of D₁ versus D, receptors in the nucleus accumbens core, but no significant differences were found between exercising and sedentary heroin-exposed rat groups in dopamine receptor mRNA expression at nucleus accumbens shell (Robison et al., 2018). Therefore, these differences in effects of exercise on drug abuse are probably related to the kind of drug abuse and site injection of dopamine receptors antagonists. Moreover, exercise can enhance dopamine and may be able to provide even more benefits in suppressing the reward from substance use (Volkow and Baler, 2014).

Conclusion

In conclusion, the results of the present study demonstrated that treadmill running reduced morphine sensitization behavior and induced euphoria in rats. Previous studies revealed the effects of physical activity exert by the release of endogenous opioid and dopamine neurotransmitters in different parts of the brain. Moreover, we found that the microinjection of D_1 and D_2 antagonists into the IPAG region decreased the reinforcement effects of morphine. Considering the association of aerobic exercise with dopamine antagonism, the role of the D_2 -receptor in the sensitization of morphine was more pronounced than that of the D_1 receptor. It is likely that the running activity can protect against addiction via the release of dopamine by activating D_2 dopamine receptors in the IPAG area.

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Conflict of interest

The authors declare no conflict of interests.

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