

Original Article

Ferulic acid, a phenolic compound with therapeutic effects in neuropsychiatric disorders, stimulates the production of nerve growth factor and endocannabinoids in rat brain

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Abstract

Introduction: Ferulic acid, a phenolic phytochemical with neuroprotective, anti-inflammatory, and antioxidant properties, has shown promising antidepressant-like effects in behavioral studies; however, its mechanism(s) of action have not been fully understood. Based on the contribution of nerve growth factor (NGF) and endocannabinoid signaling (eCBs) to the emotional or antidepressant activity and their interaction, we aimed to evaluate whether ferulic acid affects NGF or eCB contents in the brain regions involved in the modulation of emotions.

Methods: Following single and four-week once-daily intraperitoneal injections of ferulic acid (50, 100, 130 and 150 mg/kg), amitriptyline (2.5, 5, 8 and 10 mg/kg) or lorazepam (2, 5, 8 and 10 mg/kg) into male Wistar rats, NGF and eCB levels were quantified by Bio-Rad protein assay and isotope-dilution liquid chromatography/mass spectrometry. In the case of significant alteration of brain NGF or eCB content, the effects of pre-treatment with cannabinoid CB₁ or CB₂ receptor antagonist (AM251 or SR144528) were investigated.

Results: Four-week treatment with the highest doses of ferulic acid or amitriptyline led to a significant and sustained enhancement of eCB and NGF contents in brain region-specific fashion. Neither acute nor four-week treatment with lorazepam affected NGF or eCB levels. Pre-treatment with AM251 (3 mg/kg), but not SR144528, prevented the elevation of NGF levels. AM251 showed no effect by itself.

Conclusion: Ferulic acid similar to the conventional antidepressant, amitriptyline, affects brain eCB and NGF signaling. CB₁ receptors mediate the production of brain NGF.

Keywords:

Ferulic acid;
Nerve growth factor;
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Introduction

The high prevalence of neuropsychiatric disorders

including the depression has raised increasing concerns among the health authorities (Baumeister and Harter, 2007). Conventional therapeutic agents are usually associated with limited efficiency,

resistance, significant side effects and drug-drug or drug-food interactions (Ereshefsky, 2009; Santarsieri and Schwartz, 2015). In recent years, research efforts have been attracted towards the identification of nutraceuticals or phytochemicals with antidepressant properties (Nemeroff, 2007; van der Watt et al., 2008). Ferulic acid (4-hydroxy-3-methoxycinnamic acid), a natural phenolic compound, has shown a wide range of pharmacological activities including the effects against the oxidative stress, inflammation, aging and neurological disorders such as the spinal cord injury, stroke, Alzheimer's disease and epilepsy (Cheng et al., 2008; Gohil et al., 2012; Hassanzadeh et al., 2017; Srinivasan et al., 2007; Wu et al., 2014; Yan et al., 2001). Furthermore, this bioflavonoid via the enhancement of monoamine neurotransmitter levels in the brain, modulation of oxidative/nitrosative stress and activities of protein kinase A, protein kinase C, mitogen-activated protein kinase/extracellular signal-regulated kinase, Ca²⁺/calmodulin-dependent protein kinase II and phosphatidylinositol-3-kinase signalling pathways has shown therapeutic effects against the depression-like behaviors (Chen et al., 2015; Zeni et al., 2012; Zhang et al., 2011). Over the last decade, implication of the neurotrophins in antidepressant activity has been well-documented (Aloe et al., 2015; Berton and Nestler, 2006; Tanis and Duman, 2007). Neurotrophins which are critically involved in the neuronal growth, differentiation and function (Huang and Reichardt, 2001), have shown neuroprotective effects against the neuropsychiatric disorders (Castren et al., 2007; Shaltiel et al., 2007). The prototypical neurotrophin, nerve growth factor (NGF), which regulates stress and cognitive function (Ebendal, 1992; Lad et al., 2003), is implicated in the mechanisms of action of a variety of psychotropic agents (Aloe et al., 2015; Berton and Nestler, 2006). This, prompted us to evaluate the effect of ferulic acid on NGF production in the brain regions including the frontal cortex, hippocampus, amygdala, olfactory bulb and brain stem which are involved in the processing of emotional information. As a positive control, we selected the tricyclic antidepressant, amitriptyline, due to its neurotrophic activity (Chadwick et al., 2011; Jang et al., 2009). The benzodiazepine, lorazepam, was also used for comparative purposes.

Based on the modulatory effects of the endocannabinoid system (eCBs) on a variety of

pathophysiological or physiological processes including the emotional reactivity (Bambico et al., 2009; Gobbi et al., 2005; Hassanzadeh and Rostami, 2014; McLaughlin and Gobbi, 2012; Pacher et al., 2006; Viveros et al., 2005) and its interaction with neurotrophin signaling (Hassanzadeh, 2010; Khaspekov et al., 2004; Williams et al., 2003), we focused on the eCBs in order to identify a potential mechanism by which brain NGF production may be regulated by ferulic acid, amitriptyline or lorazepam. For further assessment of the role of the eCB transmission in the mechanisms of action of the abovementioned drugs, the brain contents of two major endocannabinoids, N-arachidonyl ethanolamine (anandamide, AEA) and 2-arachidonylglycerol (2-AG), were measured after the acute and four-week treatment.

Materials and methods

Animals

Randomly assigned male Wistar rats weighing 250-280 g (Pasteur Institute, Tehran, Iran) were housed three per cage on a 12-h light/dark cycle under the standard laboratory conditions (temperature: 22 ± 2 °C, humidity: 55–65%). Food pellets and water were available *ad libitum* and animals were allowed to acclimatize to the housing conditions for one week prior to the experiments. The maintenance and care of experimental animals complies with National Institutes of Health guidelines for the humane use of laboratory animals and has been approved by Institutional Ethics Committee.

Treatments

Ferulic acid (Sigma Aldrich, Germany) was dissolved in 0.5% dimethyl sulfoxide (DMSO, Sigma Aldrich, Germany) and administered at doses of 50, 100, 130 and 150 mg/kg (Koh, 2012a and b; Lenzi et al., 2015). Amitriptyline hydrochloride (Sigma Aldrich, Germany) was dissolved in 0.9% saline and injected at doses of 2.5, 5, 8 and 10 mg/kg (Jang et al., 2009; di Matteo et al., 2000). Lorazepam (Sigma Aldrich, Germany) was dissolved in 3 to 4 drops of Tween 80 (Sigma-Aldrich, Germany) prior to the addition of 0.9% saline and injected at doses of 2, 5, 8 and 10 mg/kg (Jackson et al., 2003; Mehrotra and Jadhav, 2006). In order to compare the effects of acute treatment and cumulative doses, animals received

once daily injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicle between 9:00 to 10:00 am for either one day or 28 consecutive days ($n=6/\text{group}$). In the case of any significant alteration of brain NGF contents following the treatment with ferulic acid, amitriptyline or lorazepam, the cannabinoid CB₁ receptor antagonist, AM251 (Tocris Bioscience, UK), at doses of 1, 2 and 3 mg/kg (Dono and Currie, 2012; Femenia et al., 2010) or CB₂ receptor antagonist, SR144528 (N-[(1S)-endo-1,3,3-trimethylbicyclo [2.2.1] heptan-2-yl] 5-(4-choro-3-methylphenyl)-1-(4-methyl benzyl pyrazole-3-carboxamide, Sigma Aldrich, Germany), at doses of 1, 2.5 and 5 mg/kg (Abalo et al., 2010) was dissolved in DMSO, Tween-80 and 0.9% saline in a 1:1:8 ratio and injected either alone or 30 min prior to the injection of ferulic acid, amitriptyline or lorazepam ($n=6/\text{group}$). All injections were intraperitoneal in a volume of 1 ml/kg.

NGF quantification

At 24, 48 or 72 h after the last injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicles (Gwinn et al., 2002; Parikh et al., 2004), the brain regional contents of NGF were measured. Animals were decapitated without anaesthesia and the brain of each animal was quickly and carefully removed from the skull. Using the Paxinos and Watson atlas (2007), the brain regions including the frontal cortex, hippocampi, amygdalae, olfactory bulbs and brain stem were dissected on a frozen pad taken from a -80 °C freezer. Then, the brain regions were weighed [frontal cortex: 0.26 ± 0.04 , hippocampi: 0.19 ± 0.02 , amygdalae: 0.08 ± 0.006 , olfactory bulbs: 0.05 ± 0.004 and brain stem: 0.38 ± 0.02 (mean \pm SEM, in grams)] and kept frozen at -80 °C. Each tissue sample was homogenized on ice in 5-6 vol of 0.25 M sucrose and 10 mM HEPES (pH 7.0) containing 10 mM dithiothreitol and immediately frozen in a dry ice/acetone bath and stored at -80 °C until NGF analysis. Noteworthy, the total brain NGF content is several fold higher than generally reported as the pellet which is usually discarded, contains a higher amount of NGF than the corresponding supernatant (Hoener et al., 1996). In this respect, the homogenates were centrifuged at 10,000 g (Hettich, Germany) for 10 min at 15 °C and the remaining pellets were each dissolved in 750 μ l NGF homogenization buffer, treated with ultrasound (Vibra

Cell; Bioblock Scientific, France) for 3 min and processed for further quantification of NGF by a fluorometer (Titertek Fluoroskan II, Germany) as previously described in detail (Hellweg et al., 1989; Hellweg et al., 2001; Hellweg et al., 2002; Hoener et al., 1996). The measured and recovery-corrected NGF contents are expressed as ng NGF per g protein in the resuspended NGF homogenate quantified by Bradford (1976) protein assay using a spectrophotometer (UV-1601, Shimadzu, Japan) at 595 nm. The measurements were performed in duplicate and analyzed by an investigator blind to the experimental set-up.

Extraction and analysis of the endocannabinoids

In order to evaluate the involvement of eCB transmission in the mechanisms of action of ferulic acid, amitriptyline or lorazepam, the contents of two major endocannabinoids, AEA and 2-AG, were measured in the aforementioned brain regions following both acute and four-week administration of the drugs or corresponding vehicles. Briefly, animals ($n=6/\text{group}$) were killed 1, 5 and 12 h after the last injection (de Lago et al., 2005) and the brains were quickly removed, dissected, weighed and stored frozen at -80 °C until the lipid extraction. The contents of AEA and 2-AG within the lipid extracts were determined by isotope-dilution liquid chromatography/mass spectrometry (LC-MS) using a Shimadzu LC-10ADVP HPLC apparatus (Shimadzu, Japan) coupled to a Shimadzu quadrupole MS (LCMS-2010) via a Shimadzu atmospheric pressure chemical ionization (APCI) interface (Koga et al., 1997; Patel et al., 2003). The measurements were performed in duplicate and analyzed by an investigator blind to the experimental set-up. AEA and 2-AG are expressed as pM and nM per g of wet tissue extracted, respectively.

Statistical analysis

Shapiro-Wilk test was used to verify the normal distribution of the data. The brain regional contents of AEA, 2-AG and NGF were analyzed by three-way ANOVA (treatment, dose and time point as the fixed factors and AEA, 2-AG or NGF as the dependent variable) followed by Tukey's post hoc analysis. Data are presented as mean \pm SEM (6 animals per group). The level of significance was set at $P<0.05$.

Table 1: The brain regional levels of NGF at baseline and after acute treatment with the highest doses of ferulic acid, amitriptyline, or lorazepam

Time point (h)	Brain regions	Baseline values	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid	Amitriptyline	Lorazepam
24	Frontal cortex	34.17 ± 2.43	32.58 ± 1.97	34.03± 2.94	32.17 ± 2.43	37.15 ± 3.24	35.82 ± 3.07	33.47± 1.97
	Hippocampi	77.32 ± 6.27	74.15 ± 4.69	72.28 ± 6.43	76.09 ± 5.17	75.46 ± 4.58	79.43± 5.88	75.27± 6.27
	Amygdalae	31.75 ± 3.03	33.45 ± 2.84	32.47 ± 3.04	33.17 ± 2.45	33.78 ± 2.23	44.32 ± 2.75	39.48± 2.52
	Olfactory bulbs	33.27 ± 1.94	35.03 ± 2.17	33.27 ± 2.66	31.46 ± 2.77	37.19 ± 1.89	32.67 ± 1.93	37.19 ± 1.86
	Brain stem	31.68 ± 2.66	33.42 ± 1.94	32.55 ± 3.11	35.08 ± 2.94	35.94± 2.17	39.53 ± 2.69	35.43 ± 3.09
48	Frontal cortex	34.17 ± 2.43	34.52 ± 2.89	32.27 ± 1.95	34.15 ± 3.04	33.79 ± 2.74	33.45± 2.63	35.43 ± 3.09
	Hippocampi	77.32 ± 6.27	75.11 ± 5.43	70.66 ± 5.77	72.33 ± 7.15	77.25 ± 6.15	72.19 ± 3.76	73.55 ± 4.11
	Amygdalae	31.75 ± 3.03	31.57 ± 1.86	32.79± 2.55	35.17 ± 2.33	34.28 ± 2.73	37.95 ± 1.79	37.46 ± 2.88
	Olfactory bulbs	33.27 ± 1.94	33.56 ± 2.93	31.97 ± 1.73	34.08 ± 3.24	35.86 ± 2.69	33.85 ± 2.63	39.17 ± 2.49
	Brain stem	31.68 ± 2.66	31.75 ± 3.07	34.16 ± 2.86	32.27 ± 1.88	32.48 ± 3.07	38.63 ± 1.83	31.56 ± 1.76
72	Frontal cortex	34.17 ± 2.43	34.49 ± 3.03	35.66 ± 2.79	36.03 ± 2.56	33.56 ± 1.77	31.76 ± 2.64	31.87 ± 2.43
	Hippocampi	77.32 ± 6.27	72.11 ± 7.08	75.03 ± 4.66	76.17 ± 5.37	72.48± 4.77	74.58 ± 7.05	73.48 ± 6.02
	Amygdalae	31.75 ± 3.03	35.23 ± 2.87	35.74 ± 1.73	31.84 ± 1.79	33.58± 3.09	39.41 ± 3.11	32.49 ± 2.75
	Olfactory bulbs	33.27 ± 1.94	35.11 ± 1.76	33.15 ± 2.27	32.57 ± 1.58	37.23± 2.83	31.87 ± 1.58	35.14 ± 2.56
	Brain stem	31.68 ± 2.66	35.47 ± 2.38	35.19± 3.11	35.13 ± 2.43	35.16 ± 1.49	32.96 ± 2.73	35.73± 1.82

Single injections of drugs even at the highest doses tested evoked no significant alterations in brain NGF contents as compared with the baseline values or vehicle-treated control groups. NGF levels are expressed as ng of NGF per g of protein in the resuspended NGF homogenate. Data are presented as mean±SEM (n=6/group). Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam respectively.

Results

The effects of ferulic acid, amitriptyline or lorazepam on brain regional levels of NGF

Single injection of ferulic acid, amitriptyline or lorazepam even at the highest dose tested did not affect brain regional contents of NGF at time points 24, 48 or 72 h following the injection as compared to the baseline values or vehicle-treated groups (Table 1, $P>0.05$). Four-week once-daily injection of ferulic acid, amitriptyline or lorazepam at lower doses did not affect brain NGF protein levels (Tables 2 and 3; $P>0.05$ vs. the vehicle-treated groups), however, 24 h after the last injection of ferulic acid (130 and 150 mg/kg) or amitriptyline (8 and 10 mg/kg), NGF protein levels were significantly increased in brain region-specific fashion (Table 4; $P<0.05$, $P<0.01$ and $P<0.001$). NGF contents remained significantly elevated up to 48 h (Table 4; $P<0.05$, $P<0.01$ and $P<0.001$) and 72 h (Table 4; $P<0.05$ and $P<0.01$). The main effects and interactions between the factors are as follows; {frontal cortex: [dose: $F(2,270)=140.48$, $P<0.001$], [treatment: $F(5,270)=69.89$, $P<0.001$], [time point: $F(2,270)=0.63$,

$P=0.54$], [dose × treatment: $F(10,270)=37.97$, $P<0.001$], [dose × time point × treatment: $F(20,270)=0.51$, $P=0.96$]; hippocampi: [dose: $F(2,270)=45.59$, $P<0.001$], [treatment: $F(5,270)=64.75$, $P<0.001$], [time point: $F(2,270)=0.07$, $P=0.93$], [dose × treatment: $F(10,270)=20.58$, $P<0.001$], [dose × time point × treatment: $F(20,270)=0.06$, $P=1$]; amygdalae: [dose: $F(2,270)=79.81$, $P<0.001$], [treatment: $F(5,270)=43.14$, $P<0.001$], [time point: $F(2,270)=0.17$, $P=0.85$], [dose × treatment: $F(10,270)=23.21$, $P<0.001$], [dose × time point × treatment: $F(20,270)=0.48$, $P=0.97$]; olfactory bulbs: [dose: $F(2,270)=37.01$, $P<0.001$], [treatment: $F(5,270)=34.95$, $P<0.001$], [time point: $F(2,270)=0.88$, $P=0.41$], [dose × treatment: $F(10,270)=19.94$, $P<0.001$], [dose × time point × treatment: $F(20,270)=0.45$, $P=0.98$]; brain stem: [dose: $F(2,270)=1.24$, $P=0.29$], [treatment: $F(5,270)=0.19$, $P=0.97$], [time point: $F(2,270)=2.18$, $P=0.12$], [dose × treatment: $F(10,270)=0.09$, $P=1$], [dose × time point × treatment: $F(20,270)=0.09$, $P=1$]. Four-week daily administration of lorazepam even at the highest doses tested (8 and 10 mg/kg) did not significantly affect brain NGF contents as compared to the

Table 2: The effects of four-week treatment with ferulic acid (50 mg/kg), amitriptyline (2.5 mg/kg) or lorazepam (2 mg/kg) on brain regional contents of NGF

Time point (h)	Brain regions	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid	Amitriptyline	Lorazepam
24	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.99 ± 3.11	38.64 ± 3.03	40.94 ± 3.04	36.17 ± 2.92
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	72.29 ± 5.56	79.82 ± 7.07	76.03 ± 6.82	71.63 ± 6.88
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	35.66 ± 2.31	37.84 ± 2.79	42.06 ± 2.91	35.29 ± 3.09
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.25 ± 4.03	41.06 ± 3.35	34.17 ± 2.79	35.17 ± 2.83
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	31.24 ± 3.66	39.13 ± 3.21	36.98 ± 3.07	32.69 ± 3.06
48	Frontal cortex	34.30 ± 3.16	35.87 ± 3.74	32.71 ± 2.78	37.13 ± 3.64	35.27 ± 3.02	32.92 ± 2.89
	Hippocampi	72.67 ± 5.89	74.20 ± 7.04	74.48 ± 6.45	80.04 ± 7.15	76.83 ± 5.97	71.32 ± 6.04
	Amygdalae	34.22 ± 2.83	34.14 ± 3.47	32.83 ± 3.43	37.18 ± 3.71	39.08 ± 3.18	32.75 ± 3.09
	Olfactory bulbs	32.94 ± 3.04	35.83 ± 2.85	32.53 ± 3.57	43.29 ± 4.01	38.45 ± 3.67	34.19 ± 2.94
	Brain stem	34.37 ± 3.24	32.77 ± 3.57	34.85 ± 3.93	38.17 ± 3.37	41.03 ± 3.79	35.12 ± 3.03
72	Frontal cortex	33.47 ± 2.73	34.85 ± 3.21	32.53 ± 2.84	37.18 ± 3.43	37.19 ± 3.68	34.16 ± 3.13
	Hippocampi	74.43 ± 5.48	72.98 ± 6.05	75.40 ± 6.47	76.13 ± 7.73	78.18 ± 6.71	76.03 ± 7.12
	Amygdalae	32.71 ± 3.47	33.28 ± 3.04	35.23 ± 2.74	37.03 ± 2.69	35.42 ± 3.14	35.19 ± 2.84
	Olfactory bulbs	34.87 ± 3.15	32.55 ± 2.79	34.19 ± 3.13	39.18 ± 3.23	35.15 ± 3.03	31.79 ± 3.39
	Brain stem	34.20 ± 3.17	31.49 ± 3.50	33.93 ± 3.53	38.51 ± 4.03	35.92 ± 3.75	37.06 ± 2.63

24, 48 and 72 h represent the time points at which NGF contents were measured after the last injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicles. NGF levels are expressed as ng of NGF per g of protein in the resuspended NGF homogenate. Data are presented as mean ± SEM (n=6/group). Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively.

Table 3: The effects of four-week administration of ferulic acid (100 mg/kg), amitriptyline (5 mg/kg) or lorazepam (5 mg/kg) on brain regional levels of NGF

Time point (h)	Brain regions	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid	Amitriptyline	Lorazepam
24	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.99 ± 3.11	41.13 ± 4.09	39.28 ± 3.11	34.29 ± 3.07
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	72.29 ± 5.56	78.94 ± 7.42	81.24 ± 6.93	74.15 ± 7.13
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	35.66 ± 2.31	41.13 ± 3.87	38.13 ± 3.15	32.98 ± 2.83
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.25 ± 4.03	38.72 ± 3.03	40.13 ± 3.82	37.11 ± 3.17
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	31.24 ± 3.66	40.13 ± 3.86	42.03 ± 3.94	34.13 ± 2.98
48	Frontal cortex	34.30 ± 3.16	35.87 ± 3.74	32.71 ± 2.78	39.87 ± 3.28	41.17 ± 2.94	32.96 ± 2.97
	Hippocampi	72.67 ± 5.89	74.20 ± 7.04	74.48 ± 6.45	81.29 ± 7.86	83.12 ± 8.02	72.19 ± 6.83
	Amygdalae	34.22 ± 2.83	34.14 ± 3.47	32.83 ± 3.43	42.34 ± 4.11	39.21 ± 3.17	34.05 ± 3.12
	Olfactory bulbs	32.94 ± 3.04	35.83 ± 2.85	32.53 ± 3.57	44.08 ± 3.83	37.81 ± 4.06	33.92 ± 2.74
	Brain stem	34.37 ± 3.24	32.77 ± 3.57	34.85 ± 3.93	40.29 ± 2.77	39.18 ± 2.79	32.45 ± 4.03
72	Frontal cortex	33.47 ± 2.73	34.85 ± 3.21	32.53 ± 2.84	39.15 ± 3.96	42.14 ± 3.08	32.17 ± 3.13
	Hippocampi	74.43 ± 5.48	72.98 ± 6.05	75.40 ± 6.47	77.39 ± 6.98	75.47 ± 7.13	75.23 ± 7.29
	Amygdalae	32.71 ± 3.47	33.28 ± 3.04	35.23 ± 2.74	37.57 ± 2.78	35.23 ± 3.07	36.02 ± 4.06
	Olfactory bulbs	34.87 ± 3.15	32.55 ± 2.79	34.19 ± 3.13	35.14 ± 3.06	38.21 ± 3.12	31.47 ± 2.81
	Brain stem	34.20 ± 3.17	31.49 ± 3.50	33.93 ± 3.53	37.16 ± 2.87	33.82 ± 3.23	35.14 ± 4.11

24, 48 and 72 h represent the time points at which NGF contents were measured following the last injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicles. NGF levels are expressed as ng of NGF per g of protein in the resuspended NGF homogenate. Data are presented as mean ± SEM (n=6/group). Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively.

Table 4: The effects of four-week administration of ferulic acid (130 and 150 mg/kg), amitriptyline (8 and 10 mg/kg) or lorazepam (8 and 10 mg/kg) on brain regional contents of NGF

Time point (h)	Brain regions	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid (130 mg/kg)	Ferulic acid (150 mg/kg)	Amitriptyline (8 mg/kg)	Amitriptyline (10 mg/kg)	Lorazepam (8 mg/kg)	Lorazepam (10 mg/kg)
24	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.99 ± 3.11	59.16 ± 2.47**	63.31 ± 3.23**	66.21 ± 2.32***	69.07 ± 3.09**	35.56 ± 2.77	38.14 ± 3.23
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	72.29 ± 5.56	124.05 ± 8.63**	129.17 ± 6.13***	113.81 ± 9.76*	109.75 ± 6.27**	73.01 ± 6.87	67.86 ± 3.45
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	35.66 ± 2.31	61.36 ± 3.03**	64.53 ± 3.09**	56.38 ± 2.59**	60.14 ± 2.17***	35.13 ± 2.99	40.57 ± 4.11
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.25 ± 4.03	58.52 ± 2.83**	52.19 ± 3.17*	64.33 ± 2.95***	72.25 ± 4.06**	32.46 ± 3.06	36.11 ± 3.28
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	31.24 ± 3.66	35.18 ± 3.61	31.44 ± 2.39	38.11 ± 3.68	33.97 ± 2.55	33.93 ± 3.29	31.42 ± 2.73
48	Frontal cortex	34.30 ± 3.16	35.87 ± 3.74	32.71 ± 2.78	62.86 ± 3.05**	58.03 ± 3.14*	60.85 ± 3.96**	63.72 ± 2.49***	37.49 ± 2.48	33.56 ± 2.75
	Hippocampi	72.67 ± 5.89	74.20 ± 7.04	74.48 ± 6.45	111.71 ± 9.91*	117.48 ± 7.11**	115.42 ± 10.61*	103.97 ± 7.14**	75.97 ± 7.62	79.11 ± 5.44
	Amygdalae	34.22 ± 2.83	34.14 ± 3.47	32.83 ± 3.43	59.94 ± 2.28**	53.12 ± 2.56**	61.91 ± 3.15**	57.86 ± 2.33**	35.19 ± 3.07	31.76 ± 2.83
	Olfactory bulbs	32.94 ± 3.04	35.83 ± 2.85	32.53 ± 3.57	66.41 ± 3.11**	70.37 ± 3.03***	59.12 ± 2.62**	63.29 ± 3.11**	37.29 ± 2.92	33.63 ± 3.07
	Brain stem	34.37 ± 3.24	32.77 ± 3.57	34.85 ± 3.93	42.04 ± 4.35	37.24 ± 2.97	39.05 ± 3.26	41.38 ± 2.28	36.99 ± 2.76	43.07 ± 4.13
72	Frontal cortex	33.47 ± 2.73	34.85 ± 3.21	32.53 ± 2.84	57.98 ± 3.01*	52.13 ± 2.87*	51.19 ± 3.13*	55.86 ± 2.37**	32.58 ± 3.21	37.12 ± 2.67
	Hippocampi	74.43 ± 5.48	72.98 ± 6.05	75.40 ± 6.47	104.85 ± 7.76*	98.75 ± 4.13**	108.48 ± 6.05**	96.74 ± 4.88**	75.98 ± 6.07	71.65 ± 4.12
	Amygdalae	32.71 ± 3.47	33.28 ± 3.04	35.23 ± 2.74	51.82 ± 3.14*	49.68 ± 2.11*	56.93 ± 2.49*	51.49 ± 3.09*	31.19 ± 2.75	36.29 ± 3.05
	Olfactory bulbs	34.87 ± 3.15	32.55 ± 2.79	34.19 ± 3.13	58.84 ± 3.09*	63.29 ± 3.53*	53.78 ± 2.51*	57.26 ± 2.66*	35.64 ± 3.86	32.73 ± 2.16
	Brain stem	34.20 ± 3.17	31.49 ± 3.50	33.93 ± 3.53	36.17 ± 2.19	33.74 ± 2.36	37.31 ± 2.87	34.31 ± 2.87	32.39 ± 2.95	37.06 ± 3.17

24, 48 and 72 h represent the time points at which NGF contents were measured after the last injection of ferulic acid, amitriptyline, lorazepam, or the corresponding vehicles. NGF levels are expressed as ng of NGF per g of protein in the resuspended NGF homogenate. The limits of detection and quantification (LOD and LOQ) values were approximately 0.001 and 0.0028 ng/ml, respectively. Data are presented as mean ± SEM (n=6/group). Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. the corresponding vehicle.

Table 5: The effects of AM251 on the enhanced brain NGF contents induced by ferulic acid (130 and 150 mg/kg) or amitriptyline (8 and 10 mg/kg)

AM251 (mg/kg)	Brain region	Vehicle 1	Vehicle 2	Vehicle 3	AM251	AM/fer 130	AM/fer 150	AM/ami 8	AM/ami 10
1	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.45 ± 3.40	32.76 ± 2.55	61.53 ± 3.34*	63.26 ± 2.19***	59.97 ± 3.04**	54.33 ± 2.97*
	Hippocampi	73.48 ± 5.87	75.73 ± 7.17	74.09 ± 6.92	77.42 ± 4.87	111.93 ± 8.52*	107.93 ± 6.52**	115.05 ± 9.25*	109.05 ± 7.28**
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	34.18 ± 2.78	32.55 ± 1.96	57.12 ± 2.43**	59.12 ± 3.09*	60.23 ± 2.87**	65.23 ± 3.11*
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.26 ± 3.78	31.74 ± 2.48	59.26 ± 3.07*	53.26 ± 2.76**	56.49 ± 3.05*	54.12 ± 3.03*
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	32.70 ± 3.34	36.04 ± 1.79	37.48 ± 4.19	33.57 ± 2.79	35.29 ± 3.17	38.11 ± 2.86
2	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.45 ± 3.40	37.11 ± 2.56	58.51 ± 2.47**	59.15 ± 3.15*	61.64 ± 2.16***	63.75 ± 2.31**
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	74.09 ± 6.92	71.53 ± 4.33	107.86 ± 7.43*	103.12 ± 4.49**	110.17 ± 9.12*	103.79 ± 6.17**
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	34.18 ± 2.78	32.96 ± 1.66	59.90 ± 2.15**	61.27 ± 3.32*	56.41 ± 3.09*	60.39 ± 3.11*
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.26 ± 3.73	37.05 ± 2.63	56.78 ± 2.53**	50.69 ± 2.11**	53.37 ± 2.65*	51.37 ± 2.44*
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	32.70 ± 3.34	35.19 ± 1.85	35.22 ± 2.90	35.13 ± 2.23	37.91 ± 3.08	34.52 ± 2.97
3	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	34.45 ± 3.40	31.74 ± 1.69	44.38 ± 4.84	47.66 ± 2.75	49.81 ± 3.21	54.13 ± 3.37
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	74.09 ± 6.92	69.73 ± 4.77	90.28 ± 9.42	95.43 ± 7.66	88.60 ± 6.43	92.43 ± 8.72
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	34.18 ± 2.78	32.27 ± 3.03	42.93 ± 3.13	51.74 ± 3.43	45.59 ± 3.19	43.63 ± 2.36
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.26 ± 3.78	37.42 ± 1.94	42.10 ± 4.14	44.59 ± 3.17	46.85 ± 2.97	45.86 ± 4.05
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	32.70 ± 3.34	35.11 ± 2.37	38.55 ± 2.67	33.77 ± 3.24	35.63 ± 3.23	39.05 ± 2.78

Daily pre-treatment with CB₁ receptor antagonist, AM251 (1 or 2 mg/kg), did not significantly affect the elevated NGF contents, while, 3 mg/kg of AM251 showed a preventive effect ($P > 0.05$ vs. the vehicle-treated groups). AM251 showed no effect by itself ($P > 0.05$ vs. the vehicle group). NGF levels are expressed as ng of NGF per g of protein in the re-suspended NGF homogenate. Vehicles 1, 2 and 3 are related to ferulic acid, amitriptyline and AM251, respectively. Data are expressed as mean ± SEM of $n = 6$ /group. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. the corresponding vehicle.

(AM/fer 130, AM/fer 150: injection of AM251 prior to the administration of ferulic acid 130 and 150 mg/kg, AM/ami 8, AM/ami 10: injection of AM251 before the administration of amitriptyline 8 and 10 mg/kg).

Table 6: The effects of SR144528 on the increased brain NGF levels induced by ferulic acid (130 and 150 mg/kg) or amitriptyline (8 and 10 mg/kg)

SR144528 (mg/kg)	Brain region	Vehicle 1	Vehicle 2	Vehicle 3	SR144528	SR/fer 130	SR/fer 150	SR/ami 8	SR/ami 10
1	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	35.13 ± 2.55	34.27 ± 3.05	63.28 ± 2.75**	59.86 ± 2.65**	61.23 ± 2.54**	57.19 ± 2.13**
	Hippocampi	73.48 ± 5.87	75.73 ± 7.17	69.74 ± 4.83	73.07 ± 6.17	117.33 ± 6.43**	102.13 ± 7.55*	119.11 ± 6.37**	104.59 ± 7.08*
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	32.45 ± 1.56	35.23 ± 3.03	50.06 ± 2.27*	55.24 ± 2.16*	62.36 ± 2.79*	63.11 ± 2.15*
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	36.13 ± 1.73	34.29 ± 1.77	54.11 ± 2.15*	50.17 ± 2.04*	53.07 ± 2.17*	57.09 ± 2.23*
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	31.55 ± 2.78	33.54 ± 2.33	35.24 ± 3.09	31.23 ± 1.95	39.27 ± 2.58	30.45 ± 1.23
2.5	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	32.44 ± 1.87	32.26 ± 2.64	53.13 ± 2.13*	52.28 ± 2.23*	64.19 ± 2.35**	60.32 ± 2.17**
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	73.11 ± 4.56	68.94 ± 6.13	113.45 ± 6.37*	105.65 ± 5.38*	113.45 ± 5.42**	116.23 ± 6.52*
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	33.52 ± 3.15	35.43 ± 2.76	57.43 ± 2.37*	64.09 ± 3.07*	50.17 ± 2.33*	63.56 ± 2.54*
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	30.15 ± 2.67	33.27 ± 1.83	49.97 ± 1.54*	53.17 ± 2.18*	49.23 ± 2.06*	55.14 ± 2.16*
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	36.05 ± 1.73	32.56 ± 2.44	37.13 ± 1.79	31.76 ± 3.03	33.45 ± 1.76	37.19 ± 3.07
5	Frontal cortex	34.75 ± 2.03	32.49 ± 2.75	36.11 ± 2.76	35.04 ± 2.69	56.83 ± 2.25*	58.43 ± 2.14*	60.56 ± 3.08*	57.23 ± 2.25*
	Hippocampi	73.48 ± 5.87	75.74 ± 7.17	70.13 ± 7.03	75.13 ± 5.73	109.65 ± 5.18**	97.59 ± 6.23*	119.14 ± 4.23**	107.54 ± 5.65*
	Amygdalae	31.57 ± 3.17	33.35 ± 2.99	36.42 ± 3.08	33.25 ± 2.85	51.39 ± 1.93*	70.13 ± 4.15*	55.36 ± 2.95*	59.25 ± 2.42*
	Olfactory bulbs	34.98 ± 2.96	31.84 ± 3.07	34.52 ± 1.55	33.53 ± 3.04	57.56 ± 1.25**	59.17 ± 2.67*	52.77 ± 3.17*	59.84 ± 1.87**
	Brain stem	32.96 ± 3.14	34.03 ± 2.89	30.87 ± 2.52	36.07 ± 3.17	33.29 ± 2.93	30.27 ± 1.76	39.13 ± 2.73	32.94 ± 3.18

Daily pre-treatment with CB₂ receptor antagonist, SR144528, did not significantly affect the elevated NGF contents induced by ferulic acid or amitriptyline ($P > 0.05$ vs. ferulic acid- or amitriptyline-treated group). SR144528 showed no effect by itself ($P > 0.05$ vs. the vehicle group). NGF levels are expressed as ng of NGF per g of protein in the re-suspended NGF homogenate. Vehicles 1, 2 and 3 are related to ferulic acid, amitriptyline and SR144528, respectively. Data are expressed as mean ± SEM of $n = 6$ /group. * $P < 0.05$ and ** $P < 0.01$ vs. the corresponding vehicle (SR/fer 130, SR/fer 150: injection of SR144528 prior to the administration of ferulic acid 130 and 150 mg/kg, SR/ami 8, SR/ami 10: injection of SR144528 before the administration of amitriptyline 8 and 10 mg/kg).

Table 7: The brain regional levels of endocannabinoids at baseline and after acute treatment with the highest dose of ferulic acid, amitriptyline, or lorazepam

Endocannabinoids	Brain regions	Baseline values	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid	Amitriptyline	Lorazepam
A AEA (pM/g tissue)	Frontal cortex	9.27 ± 0.64	7.94 ± 0.56	8.25 ± 0.62	7.63 ± 0.42	11.74± 0.96	9.53± 0.72	10.36± 0.87
	Hippocampi	27.44 ±2.13	24.93 ±2.43	25.29± 1.79	27.05 ±2.17	31.43 ± 2.55	33.06 ± 1.85	26.08 ±1.63
	Amygdalae	8.56 ± 0.41	7.82 ± 0.52	8.04 ± 0.74	7.83 ± 0.46	9.17 ±0.56	8.54 ± 0.97	11.32± 1.07
	Olfactory bulbs	7.13 ± 0.39	7.63 ± 0.47	8.15 ± 0.64	8.09 ± 0.53	7.36±0.32	6.82 ± 0.54	8.19 ± 0.74
	Brain stem	6.92 ± 0.47	6.29 ± 0.37	6.54± 0.58	7.03 ± 0.68	6.53± 0.47	9.67 ± 0.43	7.53 ± 0.38
2-AG (nM/g tissue)	Frontal cortex	5.38 ± 0.33	5.07 ± 0.42	4.96 ± 0.32	5.32 ± 0.47	4.13 ±0.28	6.73 ±0.62	6.13 ± 0.47
	Hippocampi	9.22 ± 0.67	8.79± 0.74	8.73± 0.65	7.96 ± 0.38	11.42± 0.96	9.46 ±0.45	9.08± 0.77
	Amygdalae	7.59 ± 0.28	8.03 ± 0.55	7.86 ± 0.47	9.04 ± 0.77	9.46 ± 0.53	10.55 ± 0.63	7.94 ± 0.63
	Olfactory bulbs	5.49 ± 0.43	5.36 ± 0.37	5.07± 0.26	4.85 ± 0.32	4.29 ± 0.35	9.37± 0.44	7.42 ± 0.36
	Brain stem	4.61 ± 0.37	4.97± 0.53	5.11 ± 0.32	4.74 ± 0.55	6.45 ± 0.42	4.64 ± 0.53	5.76 ± 0.67
B AEA (pM/g tissue)	Frontal cortex	9.27 ± 0.64	9.13 ± 0.57	8.73 ± 0.44	6.83 ± 0.47	8.35 ±0.44	12.43 ± 0.93	9.32 ± 0.65
	Hippocampi	27.44 ±2.13	25.34 ±1.73	27.13 ±1.86	25.36 ±2.06	23.87± 1.27	24.56 ± 1.35	27.85 ±2.13
	Amygdalae	8.56 ± 0.41	7.68 ± 0.66	8.13 ± 0.64	9.28 ± 0.53	10.44± 0.94	9.72 ± 0.47	9.38± 0.73
	Olfactory bulbs	7.13 ± 0.39	8.32 ± 0.77	7.45 ± 0.37	9.43 ± 0.55	7.63 ±0.63	9.35 ± 0.77	11.34± 1.07
	Brain stem	6.92 ± 0.47	6.49 ± 0.37	7.03 ± 0.56	7.11 ± 0.34	9.03 ± 0.35	10.09 ± 0.54	9.32± 0.87
2-AG (nM/g tissue)	Frontal cortex	5.38 ± 0.33	5.23 ± 0.42	4.96 ± 0.33	4.63 ± 0.56	9.86± 0.35	4.93 ± 0.32	5.85 ± 0.63
	Hippocampi	9.22 ± 0.67	9.36 ± 0.65	8.72 ± 0.48	8.72 ± 0.44	12.33± 0.96	9.23 ± 0.45	8.33 ± 0.46
	Amygdalae	7.59 ± 0.28	7.53 ± 0.27	8.96 ± 0.57	9.07 ± 0.38	10.07 ± 0.63	11.08 ± 0.62	9.48± 0.53
	Olfactory bulbs	5.49 ± 0.43	5.11 ± 0.43	4.73 ± 0.26	4.79 ± 0.58	9.44 ±0.42	7.54 ± 0.47	6.77 ± 0.28
	Brain stem	4.61 ± 0.37	4.37 ± 0.35	4.88 ± 0.53	5.03 ± 0.42	5.23 ± 0.38	4.58 ± 0.53	4.38± 0.58
C AEA (pM/g tissue)	Frontal cortex	9.27 ± 0.64	8.93 ± 0.67	9.17 ± 0.85	9.03 ± 0.76	8.55 ±0.32	11.03 ± 0.94	9.32 ± 0.48
	Hippocampi	27.44 ±2.13	25.58 ±1.42	27.44 ±2.57	27.15 ±1.63	25.86 ± 2.29	24.48 ± 1.56	27.05 ±2.11
	Amygdalae	8.56 ± 0.41	8.13 ± 0.59	7.79 ± 0.38	7.13 ± 0.54	10.77 ± 1.43	9.87 ± 0.45	9.39 ± 0.83
	Olfactory bulbs	7.13 ± 0.39	7.04 ± 0.45	6.58± 0.29	6.83 ± 0.36	9.48 ± 0.66	7.13 ± 0.38	9.13 ± 0.75
	Brain stem	6.92 ± 0.47	6.85 ± 0.39	7.13 ± 0.72	7.05 ± 0.68	8.03 ± 0.72	7.68 ± 0.27	7.26 ± 0.63
2-AG (nM/g tissue)	Frontal cortex	5.38 ± 0.33	4.97 ± 0.36	5.13 ± 0.45	4.97 ± 0.25	9.33 ± 0.75	11.47 ± 1.18	7.11 ± 0.84
	Hippocampi	9.22 ± 0.67	9.13 ± 0.43	7.94 ± 0.66	9.33 ± 0.63	13.55 ± 1.07	9.84 ± 0.77	10.23± 0.49
	Amygdalae	7.59 ± 0.28	9.54 ± 0.66	8.1 ± 0.72	7.92 ± 0.38	11.37± 1.05	9.43 ± 0.59	7.36 ± 0.52
	Olfactory bulbs	5.49 ± 0.43	4.83 ± 0.29	5.32 ± 0.65	4.85 ± 0.57	8.42 ± 0.89	9.49 ± 1.04	5.13 ± 0.63
	Brain stem	4.61 ± 0.37	5.09 ± 0.62	4.88 ± 0.42	5.39 ± 0.44	5.63 ± 0.47	7.08 ± 0.58	4.26 ± 0.32

Data have been obtained 1 h (A), 5 h (B), or 12 h (C) after the injection. As shown, single injections of drugs even at the highest doses tested evoked no significant changes in the brain endocannabinoid contents as compared to the baseline values or vehicle-treated control groups. Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively. Data are expressed as mean±SEM of n=6/group.

Table 8: The effects of four-week treatment with lower doses of ferulic acid, amitriptyline or lorazepam on brain regional levels of AEA and 2-AG

Endocannabinoids	Brain regions	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid (50 mg/kg)	Ferulic acid (100 mg/kg)	Amitriptyline (2.5 mg/kg)	Amitriptyline (5 mg/kg)	Lorazepam (2 mg/kg)	Lorazepam (5 mg/kg)
A AEA (pM/g tissue)	Frontal cortex	8.23 ± 0.68	8.79 ± 0.83	9.13 ± 0.57	10.37 ± 1.45	8.17 ± 1.97	11.48 ± 0.94	9.64 ± 0.55	7.45 ± 0.73	10.53 ± 0.49
	Hippocampi	26.97 ± 1.83	28.17 ± 2.15	25.38 ± 1.79	28.34 ± 2.07	30.07 ± 2.96	25.49 ± 1.04	28.77 ± 2.07	29.11 ± 1.76	26.33 ± 2.15
	Amygdalae	7.41 ± 0.59	7.84 ± 0.63	8.23 ± 0.57	9.76 ± 0.87	9.39 ± 0.85	9.87 ± 0.67	7.93 ± 0.47	10.05 ± 0.65	8.45 ± 0.93
	Olfactory bulbs	8.27 ± 0.62	8.03 ± 0.49	7.59 ± 0.83	10.29 ± 0.94	8.92 ± 0.88	10.72 ± 0.93	11.04 ± 1.16	7.19 ± 0.32	10.23 ± 0.79
	Brain stem	6.74 ± 0.67	7.13 ± 0.59	6.63 ± 0.74	7.34 ± 0.45	9.56 ± 0.77	7.53 ± 0.86	9.32 ± 0.75	6.55 ± 0.67	9.87 ± 0.83
2-AG (nM/g tissue)	Frontal cortex	4.73 ± 0.37	5.29 ± 0.44	5.08 ± 0.39	5.48 ± 0.29	9.76 ± 0.64	7.55 ± 0.47	9.97 ± 0.76	8.03 ± 0.54	8.27 ± 0.94
	Hippocampi	9.12 ± 0.63	9.36 ± 0.87	8.55 ± 0.47	12.03 ± 2.37	11.32 ± 0.85	9.72 ± 0.63	12.46 ± 1.85	10.23 ± 0.92	11.47 ± 1.03
	Amygdalae	7.59 ± 0.39	8.13 ± 0.58	7.68 ± 0.73	10.39 ± 0.86	11.78 ± 1.03	10.07 ± 1.13	7.34 ± 0.53	10.07 ± 1.22	9.56 ± 0.83
	Olfactory bulbs	5.27 ± 0.41	5.49 ± 0.47	4.72 ± 0.46	8.42 ± 0.62	8.94 ± 0.38	9.15 ± 0.86	5.33 ± 0.62	7.54 ± 0.73	5.77 ± 0.42
	Brain stem	4.65 ± 0.28	4.77 ± 0.52	4.16 ± 0.33	5.63 ± 0.44	7.92 ± 0.45	5.76 ± 0.33	4.86 ± 0.45	5.33 ± 0.29	4.69 ± 0.57
B AEA (pM/g tissue)	Frontal cortex	8.77 ± 0.64	9.13 ± 0.48	8.94 ± 0.93	12.07 ± 2.11	10.22 ± 0.97	8.49 ± 0.76	12.03 ± 1.96	9.17 ± 0.72	7.38 ± 0.45
	Hippocampi	28.11 ± 1.57	28.07 ± 2.94	26.89 ± 2.46	30.14 ± 2.87	27.45 ± 1.56	25.48 ± 2.04	29.33 ± 1.79	30.25 ± 3.07	28.37 ± 1.82
	Amygdalae	8.06 ± 0.75	7.59 ± 0.37	7.63 ± 0.55	7.52 ± 0.46	10.86 ± 1.07	7.75 ± 0.53	9.23 ± 0.38	8.74 ± 0.93	7.25 ± 0.36
	Olfactory bulbs	7.85 ± 0.69	8.23 ± 0.49	8.05 ± 0.72	11.03 ± 0.95	8.11 ± 0.67	10.07 ± 0.93	7.39 ± 0.93	10.82 ± 1.03	9.38 ± 0.94
	Brain stem	7.14 ± 0.53	6.76 ± 0.33	6.49 ± 0.57	7.55 ± 0.72	8.53 ± 0.29	6.55 ± 0.35	6.23 ± 0.77	7.15 ± 0.82	6.53 ± 0.79
2-AG (nM/g tissue)	Frontal cortex	4.68 ± 0.29	5.27 ± 0.52	5.08 ± 0.43	9.23 ± 0.67	10.03 ± 1.85	7.45 ± 0.73	9.33 ± 0.54	7.64 ± 0.35	5.79 ± 0.44
	Hippocampi	8.75 ± 0.82	8.58 ± 0.73	9.11 ± 0.68	9.85 ± 0.95	9.43 ± 0.76	11.32 ± 1.07	7.42 ± 0.76	11.06 ± 0.94	7.85 ± 0.65
	Amygdalae	8.26 ± 0.55	7.34 ± 0.49	8.06 ± 0.72	12.38 ± 1.23	9.05 ± 0.43	9.06 ± 0.94	11.29 ± 1.45	7.42 ± 0.48	11.36 ± 1.27
	Olfactory bulbs	4.69 ± 0.48	5.48 ± 0.53	5.13 ± 0.27	7.24 ± 0.35	6.78 ± 0.65	6.45 ± 0.37	8.11 ± 0.43	7.13 ± 0.85	6.56 ± 0.83
	Brain stem	4.07 ± 0.45	4.27 ± 0.26	4.76 ± 0.39	6.03 ± 0.67	5.13 ± 0.27	4.38 ± 0.25	8.57 ± 0.92	9.08 ± 0.97	4.77 ± 0.49
C AEA (pM/g tissue)	Frontal cortex	9.14 ± 0.59	9.03 ± 0.73	8.79 ± 0.84	11.74 ± 0.94	9.77 ± 0.65	11.43 ± 1.67	9.45 ± 0.83	9.38 ± 0.54	10.24 ± 0.86
	Hippocampi	27.03 ± 2.65	27.19 ± 3.11	25.88 ± 2.97	26.13 ± 1.86	30.11 ± 1.77	27.13 ± 2.69	24.33 ± 1.87	29.04 ± 2.11	27.12 ± 1.55
	Amygdalae	7.49 ± 0.34	8.14 ± 0.44	8.22 ± 0.78	10.43 ± 0.87	12.07 ± 1.69	9.66 ± 0.83	10.23 ± 0.97	9.36 ± 0.88	8.53 ± 0.43
	Olfactory bulbs	6.94 ± 0.53	7.25 ± 0.49	7.97 ± 0.72	6.75 ± 0.45	10.04 ± 1.64	7.44 ± 0.57	9.11 ± 0.83	7.86 ± 0.52	6.49 ± 0.27
	Brain stem	7.26 ± 0.69	7.97 ± 0.44	6.35 ± 0.29	9.22 ± 0.82	7.12 ± 0.48	10.06 ± 0.86	6.44 ± 0.35	9.17 ± 0.74	10.03 ± 0.94
2-AG (nM/g tissue)	Frontal cortex	5.27 ± 0.42	4.79 ± 0.23	4.58 ± 0.55	6.32 ± 0.57	5.48 ± 0.39	6.22 ± 0.78	6.43 ± 0.65	4.32 ± 0.28	4.98 ± 0.63
	Hippocampi	7.94 ± 0.78	7.69 ± 0.49	8.17 ± 0.74	10.21 ± 1.83	7.28 ± 0.47	7.13 ± 0.43	10.22 ± 1.08	7.87 ± 0.93	9.15 ± 0.97
	Amygdalae	8.22 ± 0.79	7.97 ± 0.65	8.06 ± 0.69	7.43 ± 0.76	9.13 ± 0.57	9.42 ± 0.89	9.34 ± 0.94	10.16 ± 1.13	7.56 ± 0.63
	Olfactory bulbs	5.58 ± 0.45	4.78 ± 0.53	5.33 ± 0.28	8.07 ± 0.93	5.87 ± 0.33	7.64 ± 0.82	7.45 ± 0.86	8.23 ± 0.95	4.92 ± 0.43
	Brain stem	4.89 ± 0.24	4.62 ± 0.37	5.17 ± 0.52	4.82 ± 0.58	7.13 ± 0.86	5.75 ± 0.64	4.96 ± 0.32	4.65 ± 0.88	6.04 ± 0.86

Data have been obtained 1 h (A), 5 h (B), or 12 h (C) after the last injection in four-week once-daily injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicles. Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively. Data are presented as mean ± SEM (n=6/group).

Table 9: The effects of four-week treatment with ferulic acid (130 and 150 mg/kg), amitriptyline (8 and 10 mg/kg) or lorazepam (8 and 10 mg/kg) on brain regional contents of AEA and 2-AG

Endocannabinoids	Brain regions	Vehicle 1	Vehicle 2	Vehicle 3	Ferulic acid (130 mg/kg)	Ferulic acid (150 mg/kg)	Amitriptyline (8 mg/kg)	Amitriptyline (10 mg/kg)	Lorazepam (8 mg/kg)	Lorazepam (10 mg/kg)
A AEA (pM/g tissue)	Frontal cortex	8.23 ± 0.68	8.79 ± 0.83	9.13 ± 0.57	20.44 ± 2.34**	23.17 ± 2.11***	23.31 ± 2.17***	20.76 ± 1.83***	10.58 ± 0.77	13.78 ± 1.23
	Hippocampi	26.97 ± 1.83	28.17 ± 2.15	25.38 ± 1.79	41.17 ± 3.09**	39.94 ± 2.38**	38.45 ± 2.76**	43.27 ± 2.12***	25.39 ± 2.41	21.94 ± 2.03
	Amygdalae	7.41 ± 0.59	7.84 ± 0.63	8.23 ± 0.57	30.57 ± 3.17**	32.86 ± 3.21**	27.81 ± 2.64**	25.11 ± 1.93***	8.45 ± 0.72	10.05 ± 1.11
	Olfactory bulbs	8.27 ± 0.62	8.03 ± 0.49	7.59 ± 0.83	24.04 ± 2.93***	21.49 ± 1.78***	25.15 ± 3.07**	29.83 ± 2.26***	9.11 ± 0.68	7.89 ± 0.43
	Brain stem	6.74 ± 0.67	7.13 ± 0.59	6.63 ± 0.74	9.24 ± 0.49	7.73 ± 0.33	9.77 ± 0.93	7.48 ± 0.55	7.19 ± 0.74	9.42 ± 0.66
2-AG (nM/g tissue)	Frontal cortex	4.73 ± 0.37	5.29 ± 0.44	5.08 ± 0.39	15.84 ± 1.39***	13.45 ± 1.33**	17.08 ± 1.42***	15.42 ± 0.94***	7.45 ± 0.67	5.97 ± 0.38
	Hippocampi	9.12 ± 0.63	9.36 ± 0.87	8.55 ± 0.47	21.18 ± 3.17*	25.07 ± 2.83***	19.77 ± 1.25***	22.14 ± 1.33***	8.36 ± 0.49	10.28 ± 0.73
	Amygdalae	7.59 ± 0.39	8.13 ± 0.58	7.68 ± 0.73	16.29 ± 1.28**	19.36 ± 1.67**	17.39 ± 2.18**	15.48 ± 1.57**	9.03 ± 0.75	7.36 ± 0.49
	Olfactory bulbs	5.27 ± 0.41	5.49 ± 0.47	4.72 ± 0.46	13.58 ± 1.94**	11.92 ± 1.26**	16.08 ± 1.33***	13.63 ± 1.44**	4.47 ± 0.27	6.19 ± 0.23
	Brain stem	4.65 ± 0.28	4.77 ± 0.52	4.16 ± 0.33	4.09 ± 0.37	4.68 ± 0.23	5.21 ± 0.48	5.77 ± 0.29	5.26 ± 0.44	8.03 ± 0.29
B AEA (pM/g tissue)	Frontal cortex	8.77 ± 0.64	9.13 ± 0.48	8.94 ± 0.93	21.39 ± 2.86**	18.86 ± 1.92***	23.92 ± 2.43***	27.13 ± 1.97***	9.87 ± 0.45	8.43 ± 0.79
	Hippocampi	28.11 ± 1.57	28.07 ± 2.94	26.89 ± 2.46	42.07 ± 3.17**	40.83 ± 2.98**	39.28 ± 2.26*	42.28 ± 1.43***	29.42 ± 2.87	25.11 ± 1.65
	Amygdalae	8.06 ± 0.75	7.59 ± 0.37	7.63 ± 0.55	20.17 ± 2.83**	25.34 ± 2.11***	24.32 ± 2.03***	26.06 ± 2.18***	8.13 ± 0.73	11.09 ± 0.66
	Olfactory bulbs	7.85 ± 0.69	8.23 ± 0.49	8.05 ± 0.72	20.08 ± 3.14*	23.31 ± 2.65***	16.23 ± 1.95*	14.87 ± 0.68**	9.82 ± 0.67	13.25 ± 1.32
	Brain stem	7.14 ± 0.53	6.76 ± 0.33	6.49 ± 0.57	9.13 ± 0.83	7.45 ± 0.64	9.37 ± 0.49	6.89 ± 0.35	11.59 ± 0.93	8.62 ± 0.73
2-AG (nM/g tissue)	Frontal cortex	4.68 ± 0.29	5.27 ± 0.52	5.08 ± 0.43	18.24 ± 1.29***	22.19 ± 2.76**	15.86 ± 1.97**	20.06 ± 2.47**	4.98 ± 0.43	7.13 ± 0.69
	Hippocampi	8.75 ± 0.82	8.58 ± 0.73	9.11 ± 0.68	17.84 ± 1.93**	20.67 ± 1.23***	21.18 ± 2.83**	18.76 ± 1.59**	9.48 ± 0.72	7.21 ± 0.33
	Amygdalae	8.26 ± 0.55	7.34 ± 0.49	8.06 ± 0.72	16.48 ± 1.76**	14.93 ± 1.59*	15.43 ± 2.27*	18.38 ± 1.23**	9.56 ± 0.63	13.42 ± 1.15
	Olfactory bulbs	4.69 ± 0.48	5.48 ± 0.53	5.13 ± 0.27	14.43 ± 1.28***	19.21 ± 2.35**	15.08 ± 2.18*	12.08 ± 1.73*	5.33 ± 0.35	8.12 ± 0.75
	Brain stem	4.07 ± 0.45	4.27 ± 0.26	4.76 ± 0.39	5.77 ± 0.45	4.82 ± 0.53	4.92 ± 0.49	6.38 ± 0.37	4.48 ± 0.53	6.19 ± 0.27
C AEA (pM/g tissue)	Frontal cortex	9.14 ± 0.59	9.03 ± 0.73	8.79 ± 0.84	16.83 ± 1.28**	13.42 ± 0.94*	15.69 ± 1.53*	13.94 ± 0.78*	9.13 ± 0.72	8.47 ± 0.45
	Hippocampi	27.03 ± 2.65	27.19 ± 3.11	25.88 ± 2.97	37.44 ± 2.27*	39.21 ± 1.43**	39.26 ± 2.54*	32.72 ± 1.44**	27.95 ± 3.07	23.62 ± 1.39
	Amygdalae	7.49 ± 0.34	8.14 ± 0.44	8.22 ± 0.78	17.76 ± 2.13*	23.03 ± 1.87**	19.72 ± 2.87*	22.58 ± 2.32**	9.11 ± 0.66	7.69 ± 0.53
	Olfactory bulbs	6.94 ± 0.53	7.25 ± 0.49	7.97 ± 0.72	16.23 ± 1.86**	14.79 ± 1.54*	13.94 ± 2.42*	17.09 ± 1.13**	10.25 ± 1.03	7.25 ± 0.45
	Brain stem	7.26 ± 0.69	7.97 ± 0.44	6.35 ± 0.29	8.73 ± 0.56	6.63 ± 0.48	7.13 ± 0.59	9.27 ± 0.84	6.55 ± 0.27	8.19 ± 0.32
2-AG (nM/g tissue)	Frontal cortex	5.27 ± 0.42	4.79 ± 0.23	4.58 ± 0.55	10.48 ± 0.64*	14.29 ± 0.55**	11.69 ± 0.92*	13.27 ± 1.09*	9.17 ± 0.48	6.33 ± 0.75
	Hippocampi	7.94 ± 0.78	7.69 ± 0.49	8.17 ± 0.74	17.27 ± 1.79*	15.49 ± 0.43**	14.87 ± 1.16*	18.03 ± 0.92**	7.16 ± 0.39	11.08 ± 0.66
	Amygdalae	8.22 ± 0.79	7.97 ± 0.65	8.06 ± 0.69	11.52 ± 0.94*	15.27 ± 0.88*	14.29 ± 1.09*	12.08 ± 0.96*	7.99 ± 0.42	9.28 ± 0.67
	Olfactory bulbs	5.58 ± 0.45	4.78 ± 0.53	5.33 ± 0.28	10.27 ± 0.99*	13.73 ± 0.57*	12.69 ± 1.22**	16.29 ± 1.58**	6.07 ± 0.73	8.13 ± 0.79
	Brain stem	4.89 ± 0.24	4.62 ± 0.37	5.17 ± 0.52	7.13 ± 0.66	4.47 ± 0.29	6.24 ± 0.43	5.38 ± 0.37	7.24 ± 0.39	4.93 ± 0.43

Data have been obtained 1 h (A), 5 h (B), or 12 h (C) after the last injection in four-week once-daily injection of ferulic acid, amitriptyline, lorazepam or the corresponding vehicles. Vehicle 1, 2 and 3 are related to ferulic acid, amitriptyline and lorazepam, respectively. Data are presented as mean ± SEM (n=6/group). * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ vs. the respective controls.

vehicle-treated group (Table 4; $P>0.05$).

The effect of CB₁ or CB₂ receptor antagonist on the elevated brain NGF levels induced by ferulic acid or amitriptyline

Following daily pre-treatment with 1 or 2 mg/kg AM251, brain NGF contents remained significantly elevated (Table 5; $P<0.05$, $P<0.01$ and $P<0.001$), while, 3 mg/kg of AM251 prevented the enhancement of NGF induced by ferulic acid or amitriptyline (Table 5; $P>0.05$ vs. the vehicle-treated groups). AM251 (3 mg/kg) showed no effect by itself (Table 5, $P>0.05$ vs. the vehicle group). Pre-application of the CB₂ receptor antagonist, SR144528, did not significantly affect the elevated brain NGF contents induced by ferulic acid or amitriptyline (Table 6, $P>0.05$ vs. ferulic acid- or amitriptyline-treated group). SR144528 showed no effect by itself (Table 6, ($P>0.05$ vs. the vehicle group).

The effects of ferulic acid, amitriptyline or lorazepam on brain regional levels of AEA and 2-AG

one hour after the acute treatment with ferulic acid, amitriptyline or lorazepam, the brain regional eCB contents did not significantly differ from those of baseline values or vehicle-treated groups (Table 7, $P>0.05$). Similar findings were obtained 5 or 12 h after the acute treatment (Table 7, $P>0.05$). Four-week once-daily administration of the lower doses of ferulic acid, amitriptyline or lorazepam did not significantly affect brain eCB contents at any time point tested (Table 8, $P>0.05$), while, four-week exposure to 130 and 150 mg/kg of ferulic acid or 8 and 10 mg/kg of amitriptyline led to a brain region-specific elevation of AEA and 2-AG contents at 1 h from the last injection (Table 9A, $P<0.01$ and $P<0.001$) that lasted up to 5 h (Table 9B, $P<0.05$, $P<0.01$ and $P<0.001$) and 12 h (Table 9C, $P<0.05$ and $P<0.01$). The main effects and interaction between the factors are as follows: {frontal cortex (AEA): [dose: $F(2,270)=123.51$, $P<0.001$], [treatment: $F(5,270)=36.09$, $P<0.001$], [time point: $F(2,270)=0.16$, $P=0.85$], [dose \times treatment: $F(10,270)=40.31$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.13$, $P=1$]; frontal cortex (2-AG): [dose: $F(2,270)=108.27$, $P<0.001$], [treatment: $F(5,270)=99.81$, $P<0.001$], [time point: $F(2,270)=2.04$, $P=0.13$], [dose \times treatment: $F(10,270)=27.65$,

$P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.93$, $P=0.56$]; hippocampi (AEA): [dose: $F(2,270)=240.11$, $P<0.001$], [treatment: $F(5,270)=22.93$, $P<0.001$], [time point: $F(2,270)=0.27$, $P=0.93$], [dose \times treatment: $F(10,270)=13.16$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.17$, $P=0.94$]; hippocampi (2-AG): [dose: $F(2,270)=80.74$, $P<0.001$], [treatment: $F(5,270)=27.30$, $P<0.001$], [time point: $F(2,270)=0.71$, $P=0.49$], [dose \times treatment: $F(10,270)=27.79$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=1.36$, $P=0.14$]; amygdalae (AEA): [dose: $F(2,270)=126.24$, $P<0.001$], [treatment: $F(5,270)=39.28$, $P<0.001$], [time point: $F(2,270)=0.43$, $P=0.65$], [dose \times treatment: $F(10,270)=37.07$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.89$, $P=0.59$]; amygdalae (2-AG): [dose: $F(2,270)=99.59$, $P<0.001$], [treatment: $F(5,270)=17.31$, $P<0.001$], [time point: $F(2,270)=2.09$, $P=0.13$], [dose \times treatment: $F(10,270)=15.67$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=1.19$, $P=0.27$]; olfactory bulbs (AEA): [dose: $F(2,270)=81.17$, $P<0.001$], [treatment: $F(5,270)=20.75$, $P<0.001$], [time point: $F(2,270)=0.39$, $P=0.67$], [dose \times treatment: $F(10,270)=16.39$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.58$, $P=0.84$]; olfactory bulbs (2-AG): [dose: $F(2,270)=162.15$, $P<0.001$], [treatment: $F(5,270)=60.17$, $P<0.001$], [time point: $F(2,270)=1.09$, $P=0.34$], [dose \times treatment: $F(10,270)=31.79$, $P<0.001$], [dose \times time point \times treatment: $F(20,270)=0.49$, $P=0.97$]; brain stem (AEA): [dose: $F(2,270)=2.36$, $P=0.09$], [treatment: $F(5,270)=1.25$, $P=0.29$], [time point: $F(2,270)=1.89$, $P=0.15$], [dose \times treatment: $F(10,270)=0.88$, $P=0.55$], [dose \times time point \times treatment: $F(20,270)=0.40$, $P=0.99$]; brain stem (2-AG): [dose: $F(2,270)=0.06$, $P=0.94$], [treatment: $F(5,270)=0.99$, $P=0.43$], [time point: $F(2,270)=1.23$, $P=0.19$], [dose \times treatment: $F(10,270)=0.27$, $P=0.92$], [dose \times time point \times treatment: $F(20,270)=0.47$, $P=0.99$]. Lorazepam even at the highest doses tested did not significantly affect brain eCB contents (Table 9, $P>0.05$ vs. the vehicle-treated group).

Discussion

Research efforts on the health benefits of herbs have shown the ability of plants to synthesize a variety of

compounds which interact with biological pathways (Tapsell et al., 2006). Based on the limited efficacy or significant side effects of the classical antidepressants (Ereshfsky, 2009; Santarsieri and Schwartz, 2015), increasing research efforts have been attracted towards the development or identification of novel therapeutic agents including the natural antidepressants. Over the last decade, involvement of the neurotrophic factors in the mechanism of action of antidepressants has been the focus of intense research (Berton and Nestler, 2006; Yanpallewar et al., 2010; Shaltiel et al., 2007). In this study, we have found that ferulic acid similar to the conventional antidepressant, amitriptyline, is able to induce a sustained enhancement of NGF protein levels in brain region-specific fashion (Table 3). The elevation of brain NGF levels after four-week treatment suggests that the occurrence of neurotrophic effects is a slow-developing process that is in agreement with Yanpallewar et al. (2010) who have suggested that the increased expression of a trophic factor is a slow-onset adaptive change. The sustained enhancement of NGF contents in the brain regions involved in the regulation of emotional activity may be considered as a mechanism by which ferulic acid like amitriptyline exerts its antidepressant therapeutic effect. According to the stimulatory effect of NGF on cell proliferation (Cheng et al., 2009), ferulic acid- or amitriptyline-induced elevation of NGF in the frontal cortex might be of therapeutic value against the stress-induced reduction in cell proliferation in the frontal cortex (Banasr et al., 2007). As shown in Table 3, four-week administration of amitriptyline or ferulic acid resulted in a significant enhancement of NGF content in the hippocampus. Since NGF is critically involved in the hippocampal plasticity and neurogenesis (Conner et al., 2009) as well as cognitive function (Winkler et al., 2000), it seems that ferulic acid or amitriptyline via the elevation of hippocampal NGF induces hippocampal neurogenesis that may lead to the improved cognitive function. Ferulic acid or amitriptyline significantly elevated NGF protein level in the amygdala after four-week daily treatment (Table 3). This, may result in the improved cognitive performance due to the facilitatory role of NGF on the cholinergic neurotransmission between the nucleus basalis and amygdala (Moises et al., 1995). Ferulic acid like amitriptyline significantly increased

NGF level in the olfactory bulbs (Table 3). According to the pivotal role of NGF in the maintenance or development of the olfactory system (Miwa et al., 2002), ferulic acid or amitriptyline may significantly affect the function of this system. As shown in Table 3, neither ferulic acid nor amitriptyline affected NGF production in the brain stem suggesting that NGF signaling in this brain region is not implicated in the mechanisms of action of these antidepressants.

Four-week administration of lorazepam even at the highest dose tested did not significantly alter the brain regional levels of NGF (Table 3) suggesting that the ineffectiveness of lorazepam against the depression is, at least in part, due to its inability to affect brain NGF.

In an attempt to identify a mechanism by which ferulic acid or amitriptyline affects the brain NGF levels, we focused on the implication of the eCBs which its role in the treatment of neuropsychiatric diseases and interaction with neurotrophic factors have been well-documented (Bambico et al., 2009; Hassanzadeh and Hassanzadeh, 2011; Khaspekov et al., 2004; Marsicano and Lutz, 2006; Williams et al., 2003). Pretreatment with CB₁ receptor antagonist, AM251 (3 mg/kg), prevented the elevation of brain NGF contents induced by ferulic acid or amitriptyline (Table 4), while, CB₂ receptor antagonist, SR144528, failed to show preventive effects (not shown). These findings indicate that ferulic acid or amitriptyline elevates brain NGF content under the regulatory drive of CB₁ receptors.

In order to further explore the implication of eCB transmission in the mechanisms of action of ferulic acid, amitriptyline, or lorazepam, the brain regional levels of two major endocannabinoids, AEA and 2-AG, were measured at various time points after acute and four-week treatment with abovementioned drugs. As shown in Table 6, four-week once-daily treatment with the highest doses of ferulic acid or amitriptyline led to a sustained enhancement of AEA and 2-AG levels in the brain regions implicated in the modulation of emotional behaviour and synaptic plasticity. Our findings demonstrate the involvement of the intrinsic endocannabinoid tone -with a well-established regulatory role in the emotional states and neuroplasticity (Marsicano and Lutz, 2006; Viveros et al., 2007)-, in the mechanism of action of ferulic acid or amitriptyline. Four-week daily administration of the benzodiazepine and lorazepam

even at the highest doses tested did not significantly alter brain AEA and 2-AG contents (Table 6) suggesting that the eCB transmission is not involved in the mechanism of action of lorazepam.

As may be compared in Tables 3 and 6, NGF and endocannabinoids were elevated in the same brain regions following treatment with ferulic acid or amitriptyline. It appears that a balance between the endocannabinoid and NGF signalling should exist for the activities of these compounds.

As aforementioned, various molecular targets and intracellular signalling pathways mediate the therapeutic effects of ferulic acid. Besides its intrinsic radical scavenging activity (Sultana et al., 2005), ferulic acid down-regulates the expression of inducible nitric oxide (NO) synthase gene and inhibits the production of NO (Koh, 2012a), pro-inflammatory cytokines (Sadar et al., 2016) and elevates the activity of heme oxygenase (Ma et al., 2011), an enzyme which plays an important role in the neuroprotective processes (Chen, 2014). Regarding the antidepressant-like effect of ferulic acid, involvement of the norepinephrine, serotonergic and antioxidant defense systems, extracellular signal-regulated kinase cascade has been previously shown (Chen et al., 2015; Lenzi et al., 2015; Zeni et al., 2012). Based on our findings, brain NGF and eCB signalling are also involved in the mechanism of action of ferulic acid.

Conclusion

In conclusion, we have found that ferulic acid like the classical antidepressant, amitriptyline, is able to stimulate the production of brain NGF and endocannabinoids that might be of therapeutic significance in a variety of diseases which are associated with abnormal eCB or NGF signalling. Furthermore, the cannabinoid CB₁ receptors mediate the brain NGF production suggesting, once again, NGF-eCBs interplay and the potential applicability of this interaction to develop more effective therapeutic agents against the neuropsychiatric disorders.

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Conflict of interest

None of the authors has any conflict of interest to disclose.

References

- Abalo R, Cabezos PA, Vera G, Fernandez-pujol R, Martin MI. The cannabinoid antagonist SR144528 enhances the acute effect of WIN 55,212-2 on gastrointestinal motility in the rat. *Neurogastroenterol Motil* 2010; 22: 694-e206.
- Aloe L, Rocco ML, Balzamino BO, Micera A. Nerve growth factor: a focus on neuroscience and therapy. *Curr Neuropharmacol* 2015; 13: 294-303.
- Bambico FR, Duranti A, Tontini A, Tarzia G, Gobbi G. Endocannabinoids in the treatment of mood disorders: evidence from animal models. *Curr Pharm Des* 2009; 15: 1623-46.
- Banasr M, Valentine GW, Li XY, Gourley SL, Taylor JR, Duman RS. Chronic unpredictable stress decreases cell proliferation in the cerebral cortex of the adult rat. *Biol Psychiatry* 2007; 62: 496-504.
- Baumeister H, Harter M. Prevalence of mental disorders based on general population surveys. *Soc Psychiatry Psychiatr Epidemiol* 2007; 42: 537-46.
- Berton O, Nestler EJ. New approaches to antidepressant drug discovery: Beyond monoamines. *Nat Rev Neurosci* 2006; 7: 137-51.
- Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976; 72: 248-54.
- Castren E, Voikar V, Rantamaki T. Role of neurotrophic factors in depression. *Curr Opin Pharmacol* 2007; 7: 18-21.
- Chadwick W, Mitchell N, Carroll J, Zhou Y, Park SS, Wang L, et al. Amitriptyline-mediated cognitive enhancement in aged 3xTg Alzheimer's disease mice is associated with neurogenesis and neurotrophic activity. *PLoS One* 2011; 6: e21660.
- Chen J, Lin D, Zhang C, Li G, Zhang N, Ruan L, et al. Antidepressant-like effects of ferulic acid: involvement of serotonergic and norepinephrine systems. *Metab Brain Dis* 2015; 30: 129-36.
- Chen J. Heme oxygenase in neuroprotection: from mechanisms to therapeutic implications. *Rev Neurosci* 2014; 25: 269-80.
- Cheng CY, Ho TY, Lee EJ, Su SY, Tang NY, Hsieh CL. Ferulic acid reduces cerebral infarct through its antioxidative and anti-inflammatory effects following transient focal cerebral ischemia in rats. *Am J Chin Med* 2008; 36: 1105-19.
- Cheng S, Ma M, Ma Y, Wang Z, Xu G, Liu X. Combination therapy with intranasal NGF and electroacupuncture enhanced cell proliferation and survival in rats after stroke. *Neurol Res* 2009; 31: 753-58.
- Conner JM, Franks KM, Titterness AK, Russell K, Merrill

- DA, Christie BR, et al. NGF is essential for hippocampal plasticity and learning. *J Neurosci* 2009; 29: 10883-89.
- de Lago E, Petrosino S, Valenti M, Morera E, Ortega-Gutierrez S, Fernandez-Ruiz J, et al. Effect of repeated systemic administration of selective inhibitors of endocannabinoid inactivation on rat brain endocannabinoid levels. *Biochem Pharmacol* 2005; 70: 446-52.
- Di Matteo V, Di Mascio M, Di Giovanni G, Esposito E. Acute administration of amitriptyline and mianserin increases dopamine release in the rat nucleus accumbens: possible involvement of serotonin 2C receptors. *Psychopharmacology (Berl)* 2000; 150: 45-51.
- Dono LM, Currie PJ. The cannabinoid receptor CB₁ inverse agonist AM251 potentiates the anxiogenic activity of urocortin I in the basolateral amygdala. *Neuropharmacology* 2012; 62: 192-99.
- Ebendal T. Function and evolution in the NGF family and its receptors. *J Neurosci Res* 1992; 32: 461-70.
- Ereshfsky L. Drug-drug interactions with the use of psychotropic medications: questions and answers. *CNS Spectr* 2009; 14: 1-8.
- Femenia T, Garcia-Gutierrez MS, Manzanares J. CB₁ receptor blockade decreases ethanol intake and associated neurochemical changes in fawn-hooded rats. *Alcohol Clin Exp Res* 2010; 34: 131-41.
- Gobbi G, Bambico FR, Mangieri R, Bortolato M, Campolongo P, Solinas M, et al. Antidepressant-like activity and modulation of brain monoaminergic transmission by blockade of anandamide hydrolysis. *Proc Natl Acad Sci USA* 2005; 102: 18620-25.
- Gohil KJ, Kshirsagar SB, Sahane RS. Ferulic acid-A comprehensive pharmacology of an important bioflavonoid. *Int J Pharm Sci Res* 2012; 3: 700-10.
- Gwinn RP, Kondratyev A, Gale K. Time-dependent increase in basic fibroblast growth factor protein in limbic regions following electroshock seizures. *Neuroscience* 2002; 114: 403-09.
- Hassanzadeh P. The endocannabinoid system: critical for the neurotrophic action of psychotropic drugs. *Biomed Rev* 2010; 21: 31-46.
- Hassanzadeh P, Hassanzadeh A. The role of the endocannabinoids in suppression of the hypothalamic-pituitary-adrenal axis activity by doxepin. *Iran J Basic Med Sci* 2011; 14: 414-21.
- Hassanzadeh P, Rostami F. CB₁ cannabinoid receptors are involved in neuroleptic-induced enhancement of brain neurotensin. *Iran J Basic Med Sci* 2014; 17: 181-88.
- Hassanzadeh P, Arbabi E, Atyabi F, Dinarvand R. Ferulic acid exhibits antiepileptogenic effect and prevents oxidative stress and cognitive impairment in the kindling model of epilepsy. *Life Sci* 2017; 179: 9-14.
- Hellweg R, Hock C, Hartun, HD. An improved rapid and highly sensitive enzyme immunoassay for nerve growth factor. *Technique J Meth Cell Mol Biol* 1989; 1: 43-48.
- Hellweg R, Thomas H, Arnswald A, von Richthofen S, Kay S, Fink H, et al. Serotonergic lesion of median raphe nucleus alters nerve growth factor content and vulnerability of cholinergic septohippocampal neurons in rat. *Brain Res* 2001; 907: 100-08.
- Hellweg R, Lang UE, Nagel M, Baumgartner A. Subchronic treatment with lithium increases nerve growth factor content in distinct brain regions of adult rats. *Mol Psychiatry* 2002; 7: 604-08.
- Hoener MC, Hewitt E, Conner JM, Costello JW, Varon S. Nerve growth factor (NGF) content in adult rat brain tissues is several-fold higher than generally reported and is largely associated with sedimentable fractions. *Brain Res* 1996; 728: 47-56.
- Huang EJ, Reichardt LF. Neurotrophins: roles in neuronal development and function. *Ann Rev Neurosci* 2001; 24: 677-736.
- Jackson A, Duka T, Stephens DN. Effects of alcohol and lorazepam during extinction of alcohol self-administration in rats. *J Psychopharmacol* 2003; 17: 293-99.
- Jang SW, Liu X, Chan CB, Weinschenker D, Hall RA, Xiao G, et al. The antidepressant amitriptyline is a TrkA and TrkB receptor agonist that promotes TrkA/TrkB heterodimerization and has potent neurotrophic activity. *Chem Biol* 2009; 16: 644-56.
- Khaspekov LG, Brenz Verca MS, Frumkina LE, Hermann H, Marsicano G, Lutz B. Involvement of brain-derived neurotrophic factor in cannabinoid receptor-dependent protection against excitotoxicity. *Eur J Neurosci* 2004; 19: 1691-98.
- Koga D, Santa T, Fukushima T, Homma H, Imai K. Liquid chromatographic-atmospheric pressure chemical ionization mass spectrometric determination of anandamide and its analogues in rat brain and peripheral tissues. *J Chromatogr B Biomed Sci Appl* 1997; 690: 7-13.
- Koh PO. Ferulic acid modulates nitric oxide synthase expression in focal cerebral ischemia. *Lab Anim Res* 2012a; 28: 273-78.
- Koh PO. Ferulic acid prevents the cerebral ischemic injury-induced decreases of astrocytic phosphoprotein PEA-15 and its two phosphorylated forms. *Neurosci Lett* 2012b; 511: 101-05.
- Lad SP, Neet KE, Mufson EJ. Nerve growth factor: structure, function and therapeutic implications for Alzheimer's disease. *Curr Drug Targets CNS Neurol Disord* 2003; 2: 315-34.
- Lenzi J, Rodrigues AF, Rós Ade S, de Castro AB, de Lima DD, Magro DD, et al. Ferulic acid chronic treatment exerts antidepressant-like effect: role of antioxidant defense system. *Metab Brain Dis* 2015; 30: 1453-63.
- Li FQ, Su H, Wang J, Liu JY, Zhu QG, Fei YB, et al. Preparation and characterization of sodium ferulate entrapped bovine serum albumin nanoparticles for liver targeting. *Int J Pharm* 2008; 349: 274-82.
- Ma ZC, Hong Q, Wang YG, Liang QD, Tan HL, Xiao CR, et al. Ferulic acid induces heme oxygenase-1 via activation of ERK and Nrf2. *Drug Discov Ther* 2011; 5: 299-305.
- Marsicano G, Lutz B. Neuromodulatory functions of the

- endocannabinoid system. *J Endocrinol Invest* 2006; 29: 27-46.
- McLaughlin RJ, Gobbi G. Cannabinoids and emotionality: a neuroanatomical perspective. *Neuroscience* 2012; 204: 134-44.
- Mehrotra N, Jadhav RN. Effects of lorazepam on brain of rat in subacute doses. *J Anat Soc India* 2006; 55: 11-14.
- Miwa T, Moriizumi T, Horikawa I, Uramoto N, Ishimaru T, Nishimura T, et al. Role of nerve growth factor in the olfactory system. *Microsc Res Tech* 2002; 58: 197-203.
- Moises HC, Womble MD, Washburn MS, Williams LR. Nerve growth factor facilitates cholinergic neurotransmission between nucleus basalis and the amygdala in rat: an electrophysiological analysis. *J Neurosci* 1995; 15: 8131-42.
- Nemeroff CB. The burden of severe depression: a review of diagnostic challenges and treatment alternatives. *J Psychiatr Res* 2007; 41: 189-206.
- Pacher P, Batkai S, Kunos G. The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol Rev* 2006; 58: 389-62.
- Parikh V, Khan MM, Alvin T, Sahebarao PM. Differential effects of typical and atypical antipsychotics on nerve growth factor and choline acetyltransferase expression in the cortex and nucleus basalis of rats. *J Psychiatr Res* 2004; 38: 521-29.
- Patel S, Rademacher DJ, Hillard CJ. Differential regulation of the endocannabinoids anandamide and 2-arachidonylglycerol within the limbic forebrain by dopamine receptor activity. *J Pharmacol Exp Ther* 2003; 306: 880-88.
- Paxinos G, Watson C. *The rat brain in stereotaxic coordinates*. 2007; San Diego: Academic press.
- Sadar SS, Vyawahare NS, Bodhankar SL. Ferulic acid ameliorates TNBS-induced ulcerative colitis through modulation of cytokines, oxidative stress, iNOS, COX-2, and apoptosis in laboratory rats. *EXCLI J* 2016; 15: 482-99.
- Santarsieri D, Schwartz TL. Antidepressant efficacy and side-effect burden: a quick guide for clinicians. *Drugs Context* 2015; 4: 212290.
- Shaltiel G, Chen G, Manji HK. Neurotrophic signalling cascades in the pathophysiology and treatment of bipolar disorder. *Curr Opin Pharmacol* 2007; 7: 22-26.
- Srinivasan M, Sudheer AR, Menon VP. Ferulic Acid: therapeutic potential through its antioxidant property. *J Clin Biochem Nutr* 2007; 40: 92-100.
- Sultana R, Ravagna A, Mohammad-Abdul H, Calabrese V, Butterfield DA. Ferulic acid ethyl ester protects neurons against amyloid beta-peptide(1-42)-induced oxidative stress and neurotoxicity: relationship to antioxidant activity. *J Neurochem* 2005; 92: 749-58.
- Tanis KQ, Duman RS. Intracellular signaling pathways pave roads to recovery for mood disorders. *Ann Med* 2007; 39: 531-44.
- Tapsell LC, Hemphill I, Cobiac L, Patch CS, Sullivan DR, Fenech M, et al. Health benefits of herbs and spices: the past, the present, the future. *Med J Aust* 2006; 185: S4-24.
- Viveros MP, Marco EM, File SE. Endocannabinoid system and stress and anxiety responses. *Pharmacol Biochem Behav* 2005; 81: 331-42.
- Viveros MP, Marco EM, Liorente R, Lopez-Gallardo M. Endocannabinoid system and synaptic plasticity: implication for emotional response. *Neural Plast* 2007; 2007: 52908.
- van der Watt G, Laugharne J, Janca A. Complementary and alternative medicine in the treatment of anxiety and depression. *Curr Opin Psychiatry* 2008; 21: 37-42.
- Williams EJ, Walsh FS, Doherty P. The FGF receptor uses the endocannabinoid signaling system to couple to an axonal growth response. *J Cell Biol* 2003; 160: 481-86.
- Winkler J, Ramirez GA, Thal LJ, Waite JJ. Nerve growth factor (NGF) augments cortical and hippocampal cholinergic functioning after p75NGF receptor-mediated deafferentation but impairs inhibitory avoidance and induces fear-related behaviors. *J Neurosci* 2000; 20: 834-44.
- Wu W, Lee SY, Wu X, Tyler JY, Wang H, Ouyang Z, et al. Neuroprotective ferulic acid (FA)-glycol chitosan (GC) nanoparticles for functional restoration of traumatically injured spinal cord. *Biomaterials* 2014; 35: 2355-64.
- Yan JJ, Cho JY, Kim HS, Kim KL, Jung JS, Huh SO, et al. Protection against b-amyloid peptide toxicity in vivo with long-term administration of ferulic acid. *Br J Pharmacol* 2001; 133: 89-96.
- Yanpallewar SU, Fernandes K, Marathe SV, Vadodaria KC, Jhaveri D, Rommelfanger K, et al. α 2-adrenoceptor blockade accelerates the neurogenic, neurotrophic, and behavioral effects of chronic antidepressant treatment. *J Neurosci* 2010; 30: 1096-109.
- Zeni LB, Zomkowski DE, Maraschin M, Rodrigues LS, Tasca I. Involvement of PKA, CaMKII, PKC, MAPK/ERK and PI3K in the acute antidepressant-like effect of ferulic acid in the tail suspension test. *Pharmacol Biochem Behav* 2012; 103: 181-86.
- Zhang YJ, Huang X, Wang Y, Xie Y, Qiu XJ, Ren P, et al. Ferulic acid-induced anti-depression and prokinetics similar to Chaihu-Shugan-San via polypharmacology. *Brain Res Bull* 2011; 86: 222-28.