



# Antioxidant and anti-inflammatory effects of *Cinnamomum* species and their bioactive compounds: An updated review of the molecular mechanisms

 Farzaneh Davoudi<sup>1</sup>, Elham Ramazani<sup>2</sup> 

1. Department of Cell and Molecular Biology, Kosar University of Bojnord, Bojnord, Iran

2. Department of Biology, Faculty of Science, Yazd University, Yazd, Iran

## ABSTRACT

**Introduction:** The genus *Cinnamomum* (cinnamon) is one of the well-known aromatic spices throughout the world with numerous medicinal applications. Several beneficial pharmacological properties of cinnamon have been evaluated, including antioxidant, anti-inflammatory, anti-diabetic, anticancer, cardiovascular-disease-lowering, and neurological disorder-improving effects. This review critically evaluates studies regarding the molecular mechanisms underlying the antioxidant and anti-inflammatory properties of cinnamon species.

**Methods:** Using three online literature databases (PubMed, Scopus, Science Direct), we identified studies describing the antioxidant and anti-inflammatory properties of cinnamon species. A literature search was carried out using a combination of keywords such as (“*Cinnamomum*,”) AND (“antioxidant” OR “anti-inflammatory”) or other related words. In this review, we evaluated new findings regarding the molecular mechanisms of antioxidant and anti-inflammatory effects of *Cinnamomum* species published from 2005 until December 2022. A total of 38 papers were selected to describe the antioxidant and anti-inflammatory properties of cinnamon species.

**Results:** Cinnamon species possess antioxidant effects by reducing ROS, MDA, and NO levels, and depleting GSH, decreasing MPO activity, and enhancing the growth of SOD and CAT. Additionally, the suppression of caspase-3 and caspase-9 activity and the upregulation of bcl-2 expression determine the anti-apoptotic effects of cinnamon. Their anti-inflammatory effects are mainly related to the reduction of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-18, IL-10, iNOS, MCP-1, and COX-2, and the inhibition of NF- $\kappa$ B, ERK1/2, p38, and JNK activation.

**Conclusion:** This review highlighted the antioxidant and anti-inflammatory effects of genus cinnamon and can provide a suitable basis for further pharmacologic surveys and efficient clinical research on cinnamon to obtain new evidence on its benefits for human health.

## Keywords:

*Cinnamon*

Antioxidant

Anti-inflammatory

\* Corresponding author: Elham Ramazani, el.ramazani@um.ac.ir

Received 2 October 2023; Revised from 8 January 2024; Accepted 17 January 2024

**Citation:** Davoudi F, Ramazani E. Antioxidant and anti-inflammatory effects of *Cinnamomum* species and their bioactive compounds: An updated review of the molecular mechanisms. Physiology and Pharmacology 2024; 28: 99-116. <http://dx.doi.org/10.61186/phypha.28.2.99>

## Introduction

The genus *Cinnamomum* (cinnamon), as a member of the Lauraceae family, comprises about 250 species distributed mostly in Asia and some areas in South and Central America, as well as Australia (Dassanayake and Larsen, 1996; Ramazani et al., 2021; Ramazani et al., 2020). Cinnamon is one of the most economically important spices worldwide and has been used as a traditional herbal medicine for thousands of years (Mahmoodnia et al., 2017). Mostly, cinnamon can be categorized into two main types including true or Ceylon cinnamon (*Cinnamomum verum* and *C. zeylanicum*) and cassia cinnamon (*C. aromaticum* and *C. burmanii*) (Shalaby and Saifan, 2014; Ramazani et al., 2020). Cinnamon is considered an essential spice due to its significant amounts of manganese, iron, calcium, and fiber (Chan et al., 2014; Tulunay et al., 2015). The most common method for extracting cinnamon essential oil is water vapor distillation (Wong et al., 2014). Cinnamon essential oil is a rich source of bioactive compounds including eugenol (10.5%),  $\alpha$ -phellandrene (7.41%), cinnamyl acetate (7.13%), terpinolene (4.49%), saffrole (5.69%), cinnamaldehyde (4.25%),  $\alpha$ -terpineol (3.21%), Linalol (2.96%) and  $\beta$ -pinene (2.51%), which are listed in Table 1 (Mahmoodnia et al., 2017; Jayaprakasha et al., 2006; Plata-Rueda et al., 2018). Cinnamon bark contains phenolic compounds such as tannin, flavonoid, volatiles, and phenolics, which significantly contribute to its antioxidant activity (Peng et al., 2008; Ervina et al., 2016). Additionally, cinnamon leaf essential oil contains  $\beta$ -cadinene (12.45%), chavicol (6.46%),  $\alpha$ -muurolene (8.79%), cis- $\beta$ -ocimene (5.17%), citral (5.33%), bornyl acetate (4.15%),  $\gamma$ -muurolene (4.34%), linalool (3.31%),  $\alpha$ -copaene (3.78%), 3-allyl-6-methoxyphenol (3.17%) methyl cinnamate (3.28%), and geraniol (3.31%), that show anti-inflammatory properties via suppressing the release of inflammatory mediators (Hao et al., 2019). Numerous studies have been reported a wide range of pharmacological activities for cinnamon, including antioxidant, anti-inflammatory, antimicrobial, antifungal, antitumor, ant-diabetic, anti-cholesterol, amelioration of unpleasant menopausal symptoms, gastrointestinal healing, and neuroprotection (Wang, et al., 2005; Jindarat et al., 2006; Elshafie et al., 2012; Csikós et al., 2020; Ramazani et al., 2021; Ramazani et al., 2020). Recently, the ability of cinnamon and its compounds to slow the SARS-CoV-2 progression has been examined (Yakhch-

ali et al., 2021). It should be mentioned that, According to the Food and Drug Administration (FDA), the use of different types of cinnamon in typical amounts in food is generally considered safe (Anderson, 2008). Taken together, the present study is designed to review the biomedical applications of cinnamon as potential pharmacological agents based on the latest research findings.

### Pharmacological effects

Among the numerous pharmacological properties of cinnamon, substantial evidence highlights its antioxidant and anti-inflammatory effects. Several *in vivo* and *in vitro* experiments have documented these impacts. Table 2 and Figure 1 summarize the main features of published studies focusing on these pharmacological properties of cinnamon.

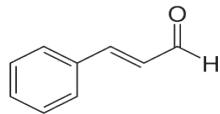
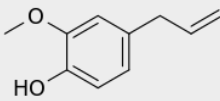
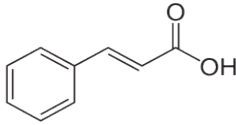
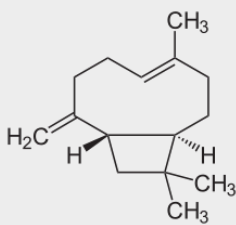
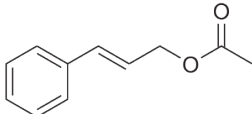
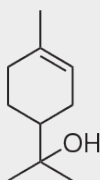
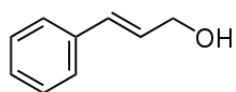
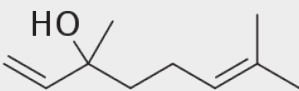
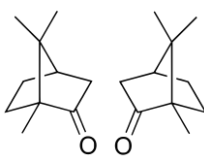
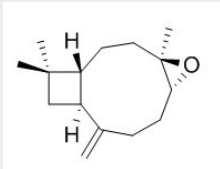
### Antioxidant effects of cinnamon

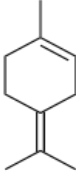
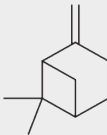
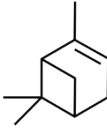
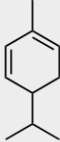
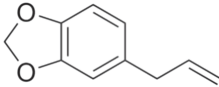
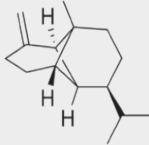
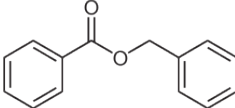
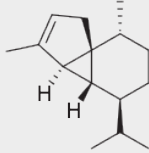
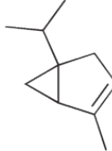
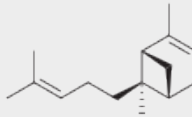
ROS encompass free radicals such as the hydroxyl radical ( $\bullet$ OH) and superoxide anion ( $O_2^{\bullet-}$ ), as well as non-radical molecules like hydrogen peroxide ( $H_2O_2$ ) and singlet oxygen ( $^1O_2$ ). The accumulation of ROS concentrations, induced by various environmental stresses, poses significant harm to organisms, leading to oxidative damage to lipids, proteins, and DNA. This oxidative damage can result in alterations to intrinsic membrane properties, including fluidity and ion transport, as well as the loss of enzyme activity, protein crosslinking, inhibition of protein synthesis, DNA damage, and ultimately, cell death (Sharma et al., 2012). Internal and external antioxidants can reduce both ROS levels and cellular oxidation (Gilgun-sherki et al., 2001).

### Clinical studies

Few Clinical studies have shown the antioxidant effects of cinnamon supplementation. In a study (25 type 2 diabetic patients for 12-weeks) showed that 1000 mg of cinnamon significantly increased the level of serum superoxide dismutase (SOD) and decreased malondialdehyde (MDA) level (Sahib, 2016). In addition, Borzoei et al. (2018) reported that 500 mg of cinnamon for 8 weeks in obese polycystic ovary syndrome (PCOS) patients exhibited antioxidant activity via reducing the level of MDA and serum total antioxidant capacity (TAC) (Borzoei et al., 2018). Also, in a double-blind study, the administration of 250 mg of cinnamon extract for 12 weeks demonstrated a considerable reduction in plasma

**TABLE 1:** Some of the main chemical constituents of *Cinnamomum* species and their bioactive compounds.

| NO. | Name                                            | Formula           | Structure                                                                             |
|-----|-------------------------------------------------|-------------------|---------------------------------------------------------------------------------------|
| 1   | Cinnamaldehyde                                  | $C_9H_8O$         |    |
| 2   | Eugenol<br>2-Methoxy-4-(prop-2-en-1-yl) phenol  | $C_{10}H_{12}O_2$ |    |
| 3   | Cinnamic acid<br>(2E)-3-Phenylprop-2-enoic acid | $C_9H_8O_2$       |    |
| 4   | (-)-β-caryophyllene                             | $C_{15}H_{24}$    |   |
| 5   | Cinnamyl acetate                                | $C_{11}H_{12}O_2$ |  |
| 6   | Terpineol                                       | $C_{10}H_{18}O$   |  |
| 7   | Cinnamyl alcohol<br>(2E)-3-Phenylprop-2-en-1-ol | $C_9H_{10}O$      |  |
| 8   | Linalool                                        | $C_{10}H_{18}O$   |   |
| 9   | Camphore                                        | $C_{10}H_{16}O$   |  |
| 10  | Caryophyllene oxide                             | $C_{15}H_{24}O$   |  |

| NO. | Name                               | Formula           | Structure                                                                             |
|-----|------------------------------------|-------------------|---------------------------------------------------------------------------------------|
| 11  | terpinolene<br>$\delta$ -Terpinene | $C_{10}H_{16}$    |    |
| 12  | beta-Pinene                        | $C_{10}H_{16}$    |    |
| 13  | $\alpha$ -pinene                   | $C_{10}H_{16}$    |    |
| 14  | $\alpha$ -Phellandrene             | $C_{10}H_{16}$    |    |
| 15  | Safrole                            | $C_{10}H_{10}O_2$ |  |
| 16  | Copaene                            | $C_{15}H_{24}$    |  |
| 17  | Benzyl benzoate<br>Ascabin         | $C_{14}H_{12}O_2$ |  |
| 18  | $\alpha$ -Cubebene                 | $C_{15}H_{24}$    |  |
| 19  | $\alpha$ -Thujene                  | $C_{10}H_{16}$    |  |
| 20  | $\alpha$ -trans-bergamotene        | $C_{15}H_{24}$    |  |

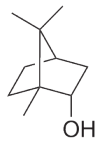
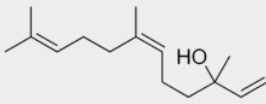
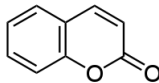
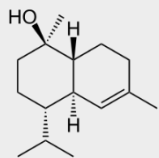
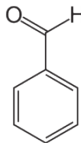
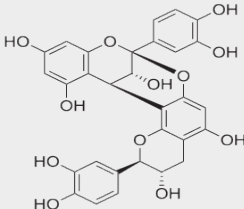
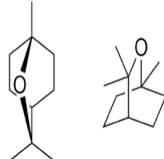
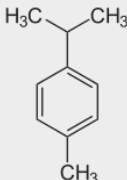
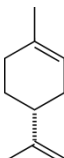
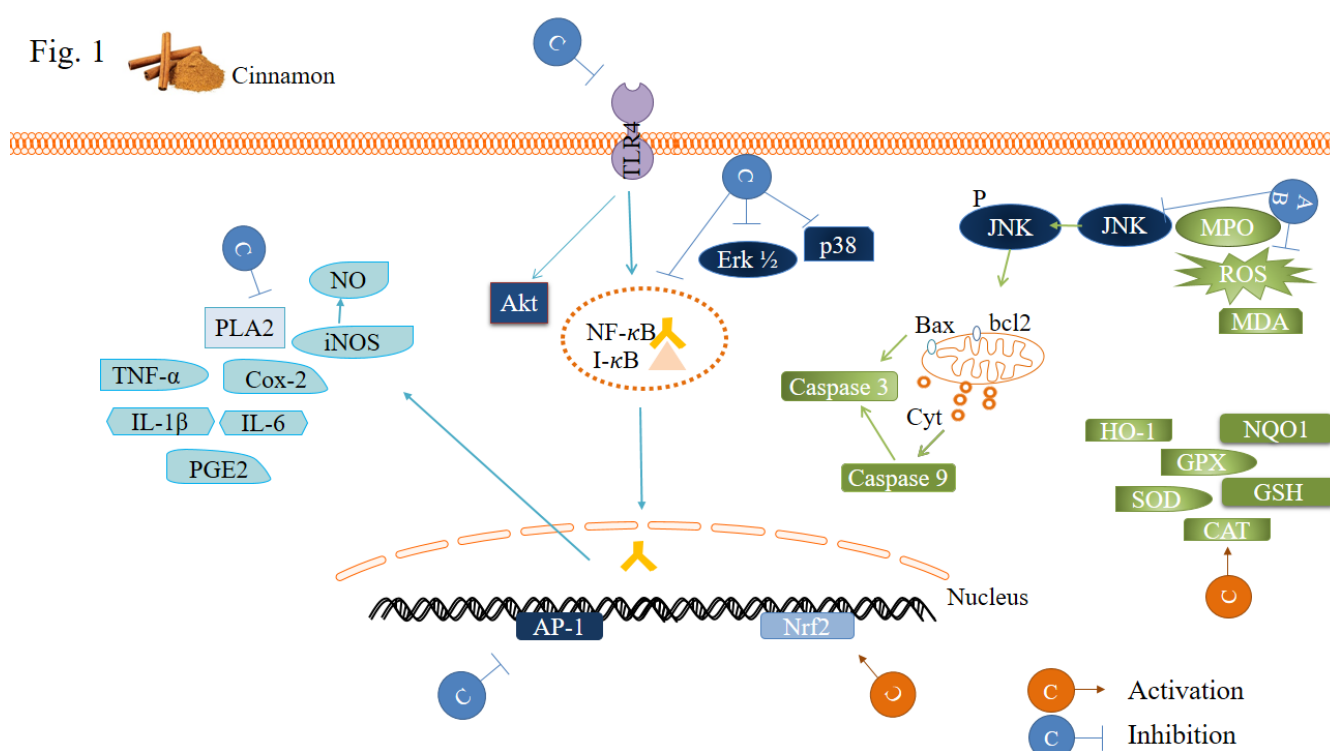
| NO. | Name              | Formula              | Structure                                                                             |
|-----|-------------------|----------------------|---------------------------------------------------------------------------------------|
| 21  | Borneol           | $C_{10}H_{18}O$      |    |
| 22  | Nerolidol         | $C_{15}H_{26}O$      |    |
| 23  | Coumarin          | $C_9H_6O_2$          |    |
| 24  | $\alpha$ -Cadinol | $C_{15}H_{26}O$      |    |
| 25  | Benzaldehyde      | $C_7H_6O$            |  |
| 26  | Procyanidin A1    | $C_{30}H_{24}O_{12}$ |  |
| 27  | 1,8-Cineole       | $C_{10}H_{18}O$      |  |
| 28  | p-Cymene          | $C_{10}H_{14}$       |  |
| 29  | Limonene          | $C_{10}H_{16}$       |  |

Fig. 1  Cinnamon



**FIGURE 1.** Intracellular signaling pathway through antioxidant (green), anti-inflammatory (blue), activity of *Cinnamomum* species and their bioactive compounds. Akt1, Protein kinase B1; AP-1, Activator protein 1; Bax, Bcl-2 associated X; Bcl-2, B-cell lymphoma 2; CAT, Catalase; COX-2, Cyclooxygenase-2; Erk1/2, Extracellular signal-regulated kinases 1/2; GSH-Px, Glutathione peroxidase; IL-6, Interleukin 6; IL1- $\beta$ , interleukin 1-b; iNOS, Inducible nitric oxide synthase; JNK, c-Jun N-terminal kinases; MDA, Malondialdehyde; NF- $\kappa$ B, Nuclear factor kappa B; NO, Nitric oxide; NQO1, NADPH-dehydrogenase-quinone-1; Nrf2, Nuclear factor erythroid 2-related factor 2; PL, Phospholipase; PG, Prostaglandin; PGE2, Prostaglandin E2; PLA2, Phospholipase A2; ROS, Reactive oxygen species; SOD, Superoxide dismutase; TNF- $\alpha$ , Tumour necrosis factor- $\alpha$ ; TLR, Toll-like receptor.

MDA and an increase in plasma sulfhydryl (SH) groups of plasma which all prove the antioxidant potential of this tropical herb in overweight or obese people (Rous-sel et al., 2009).

### *In vivo and in vitro studies*

Mathew and Abraham confirmed that the antioxidant activity of cinnamon is dose-dependent. In their study, they used the methanolic extract of *C. Verum* leaf (12.5–50 µM) and found that higher concentrations of the methanolic leaf extracts were directly related to reduced free radicals. They demonstrated that the methanolic extract of *C. Verum* leaf showed antioxidant activity through scavenging free radicals such as hydroxyl radicals and superoxide anions, and also inhibited lipid peroxidation against 1,1-diphenyl-2-picrylhydrazyl (DPPH) radicals and 2,2'-azinobis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) radicals (Mathew and Abraham, 2006). Similar results were reported by Udayaprakash et al. They found that internal methanolic extract of cinna-

mon leaves exhibited higher antioxidant capacity than butylated hydroxyanisole (BHA) (Udayaprakash et al., 2015). Similar to these results, the essential oil extracted from *C. glaucescens* showed a higher free radical-scavenging activity than butylated hydroxytoluene (BHT) (Prakash et al., 2013). Numerous pharmacological studies confirmed the antioxidant activities of cinnamon on human health. Tuzcu et al. observed that cinnamon polyphenol extract (100 mg/kg b.wt.) for 12 weeks had antioxidant effects through modulation of transcription factors including nuclear factor kappa B (NF- $\kappa$ B), and nuclear factor erythroid 2-related factor 2 (Nrf2) in high fat diet (HFD) rat liver. In another *in vivo* study by Noori et al. (2012), oral consumption of 10 mg/kg cinnamon aqueous extract for 13 consecutive days, significantly increased the level of glutathione (GSH) in rats (Noori et al., 2012). Likewise, Seyed Ahmadi et al. (2019) indicated that topical administration of cinnamon oil on infected wounds significantly reduced the high content of MDA along with increasing the antioxidant capacity

**TABLE 2:** The potential pharmacological effects of *Cinnamomum* species and their bioactive compounds.

| Subject                                                                                      | Dose (Duration)                                                                                  | Mechanism                                                                                                                                                                                                      | Ref.                        |
|----------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------|
| <i>Cinnamon and Oxidative stress</i>                                                         |                                                                                                  |                                                                                                                                                                                                                |                             |
| Type 2 diabetes patients                                                                     | 1000 mg (12-weeks)                                                                               | Increased serum SOD levels and decreased MDA levels                                                                                                                                                            | (Sahib, 2016)               |
| Obese polycystic ovary Syndrome (PCOS) patients                                              | 500 mg/day (8weeks)                                                                              | Increased antioxidant activity, reduced MDA and TAC levels                                                                                                                                                     | (Borzoei et al., 2018)      |
| Diabetic subjects                                                                            | 250 g/day (12 weeks)                                                                             | Decreased plasma MDA and increase in plasma thiol (SH) groups of plasma, increased antioxidant activity                                                                                                        | (Roussel et al., 2009)      |
| HFD-induced rats (liver)                                                                     | 100 mg/kg b.wt (12 weeks)                                                                        | Modulated NF- $\kappa$ B and Nrf2, reduced NF- $\kappa$ B p65 expression levels, induced expression of HO-1                                                                                                    | (Tuzcu et al., 2017)        |
| Human colon cancer cells (HCT116, HT29) and non-immortalized primary fetal colon cells (FHC) | 1.25-40 $\mu$ M (4 h)                                                                            | Increased Nrf2 protein levels and HO-1                                                                                                                                                                         | (Wondrak et al., 2010)      |
| Glucose-treated mice                                                                         | 10 $\mu$ M                                                                                       | Inhibited ROS generation and NO level, up-regulated Nrf2 expression, HO-1, NQO1, Catalase and Gpx1                                                                                                             | (Wang et al., 2015)         |
| <i>In vitro</i> study                                                                        | 12.5-150 $\mu$ g                                                                                 | Antioxidant activity by free radical scavenging including superoxide anions and hydroxyl radicals, inhibited lipid peroxidation against 1,1-diphenyl-2-picrylhydrazyl (DPPH) radicals and ABTS radicals action | (Mathew and Abraham, 2006)  |
| Rats                                                                                         | 10 mg/kg (13 days)                                                                               | Increased GSH                                                                                                                                                                                                  | Noori et al.,) (2012)       |
| <i>Wound model (mice)</i>                                                                    | 2 and 4% (2 and 4 g of C verum essential oil was added into 98 and 96 g of soft yellow paraffin) | Moderated MDA levels, increased the antioxidant capacity in wound tissue                                                                                                                                       | (Seyed Ahmadi et al., 2019) |
| Mice induced by APAP                                                                         | 200 mg/kg/day                                                                                    | Induced total antioxidant capacity and total oxidant status.                                                                                                                                                   | (Hussain et al., 2019)      |
| APAP-induced rats                                                                            | 100 and 200 mg/kg (14 days)                                                                      | Decreased MDA levels, increased GSH levels and antioxidant enzymes (CAT, SOD, GR and GPx) activities                                                                                                           | (Hussain et al., 2020)      |
| Rats                                                                                         | 200 mg/kg (14 days before gamma irradiation)                                                     | Restricted the TNF- $\alpha$ and NO as well as induced SOD                                                                                                                                                     | (Azab et al., 2011)         |
| Nephropathic mice                                                                            | 200 and 400 mg/kg (14 days)                                                                      | Decreased TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, increased SOD, GSH, and catalase                                                                                                                            | (Atsamo et al., 2021)       |
| HeLa and Raji cells                                                                          | 0.125 to 4 $\mu$ g/mL<br>12 to 0.375 $\mu$ g/mL                                                  | Increased antioxidant activity                                                                                                                                                                                 | (Kallel et al., 2019)       |
| Human colon cancer cells                                                                     | 75 $\mu$ M                                                                                       | Increased antioxidant activity                                                                                                                                                                                 | (Petrocelli et al., 2021)   |

| Subject              | Dose (Duration)                                                                       | Mechanism                                                   | Ref.                       |
|----------------------|---------------------------------------------------------------------------------------|-------------------------------------------------------------|----------------------------|
| Obese diabetic rats  | 200, 400 mg/kg (6 weeks)                                                              | Increased antioxidant activity                              | (Shalaby and Saifan, 2014) |
| BV2 microglial cells | 100, 150, 200 µg/mL                                                                   | Reduced NO and inflammatory cytokines IL-6, IL-18, IL-1β    | (Chen, Tang et al., 2020)  |
| LPS-induced mice     | 50 mg/kg (7 days)                                                                     | Inhibited IL-1β, MDA, and caspase-3 stimulated Nrf2 pathway | (El-ezz et al., 2018)      |
| PC12 cells           | 20 µg/ml ( <i>C. verum</i> and <i>C. cassia</i> ) 5 and 10 µM (cinnamaldehyde) (24 h) | ROS scavenging                                              | (Ramazani et al., 2020)    |

#### *Cinnamon and Inflammation*

|                                                         |                                     |                                                                                                                         |                            |
|---------------------------------------------------------|-------------------------------------|-------------------------------------------------------------------------------------------------------------------------|----------------------------|
| Type 2 diabetes patients                                | 3g/day (8weeks)                     | Reduced NF-κB level (Restricted inflammatory factors).                                                                  | (Davari et al., 2020)      |
| Caco-2 and RAW264.7                                     | 25, 50, and 100 µg/ml               | PGE2, IL-6, IL-8, TNF-α and suppressed the phosphorylation NF-κB pathways factors.                                      | (Kim and Kim, 2019)        |
| Acrylamide-intoxicated rats                             | 250 and 500 mg/kg/day (28 days)     | Induced TAC levels in the liver, lowered serum MDA and inflammatory cytokines (TNF-α, hs-CRP)                           | (Haidari et al., 2020)     |
| RAW264.7 cells                                          | 100 µg/ml                           | Decreased the IL-1β, IL-6, TNF-α, inhibited PGE2 and NO                                                                 | (Lee et al., 2006)         |
| LPS-induced mice                                        | 20, 100 and 500 mg/kg b.w. (6 days) | Reduced TNF-α and IL-6 levels, suppressed IκBα degradation and the activation of JNK, p38, and ERK.                     | (Hong et al., 2012)        |
| LPS-stimulated RAW 264.7 cells                          | 25 µg/ml and 10 µg/ml               | Inhibited production of NO and PGE2                                                                                     | (Tung et al., 2008)        |
| Raw cells                                               | 1 to 51 µg/ml                       | Reduced NO production                                                                                                   | (Lee et al., 2002)         |
| RAW 264.7 and J774A.1 macrophages                       | 0.5-1.25 mg/ml                      | Induced NO and TNF-α exhibited anti-inflammatory activity                                                               | (Gunawardena et al., 2015) |
| LPS-activated RAW264.7 cells and peritoneal macrophages | 100 µg/mL                           | Inhibited the activation of NF-κB, suppressed iNOS expression and NO production, lowered (COX)-2, TNF-α and PGE2 levels | (Yu et al., 2012)          |
| Rats                                                    | 8 mg/kg (12 to 21 days)             | Enhanced serum CRP levels                                                                                               | (Vetal et al., 2013)       |
| Colonic homogenate of colitis animals                   | 150 mg/kg/day (10 days)             | Decreased activity of MPO and the levels of NO, TNF-α, IL-1β, IL-6, and COX-2                                           | (Salamatian et al., 2019)  |

| Subject                                 | Dose (Duration)                 | Mechanism                                                                                                                            | Ref.                      |
|-----------------------------------------|---------------------------------|--------------------------------------------------------------------------------------------------------------------------------------|---------------------------|
| HDF3CGF, a human skin disease model     | 0.0012% v/v                     | Decreased monocyte chemoattractant protein-1, interferon gamma-induced protein 10, interferon-inducible T-cell alpha, and MIG        | (Han and Parker, 2017)    |
| Raw induced by LPS                      | 20, 50 mM                       | Reduced NF- $\kappa$ B, prevented transcription of COX-2 and IFN $\beta$                                                             | (Youn et al., 2008)       |
| APAP-induced rats                       | 50, 100 and 200 mg/kg (14 days) | Reduced IL-1 $\beta$ , IL-6, active caspase-3 and -9                                                                                 | (Hussain et al., 2020)    |
| Pneumonitis mice                        | 6.55 $\mu$ L/L                  | Restrained inflammatory activity                                                                                                     | (Csikós et al., 2020)     |
| THP-1 monocytes                         | 25 $\mu$ g/ml                   | Inhibited phosphorylation of Akt, I $\kappa$ B $\alpha$ , suppressed TLR $\epsilon$ and TLR $\gamma$                                 | (Schink et al., 2018)     |
| Atherosclerotic mice                    | 5, 10, 20 mg/kg (8 weeks)       | Repressed I $\kappa$ B $\alpha$ and p $\tau^{\Delta}$ NF- $\kappa$ B phosphorylation, decreased TNF- $\alpha$ , IL-6, ROS, and MCP-1 | (Li et al., 2019)         |
| LPS-induced human OA chondrocytes cells | (10, 20, and 50 mM) for 24h     | Suppressed expression of IL-1 $\beta$ , IL-6, and TNF- $\alpha$ .                                                                    | (Chen, Ruan et al., 2020) |

of the tissue (Seyed Ahmadi et al., 2019).

Also, it was found that cinnamon polyphenol decreased NF- $\kappa$ B expression levels, thereby suppressing the expression of pro-inflammatory cytokines, and increased the expression of genes encoding antioxidant proteins, resulting in upregulation of Nrf2 expression (Tuzcu et al., 2017). ROS are signal transduction pathways, and it has been found that one of the critical intracellular signaling pathways leading to NF- $\kappa$ B activation is the ROS (Ghosh and Karin, 2002). Similarly, cinnamaldehyde treatment showed antioxidant effects in the endothelium of mouse aortas by inhibiting ROS generation, protecting NO levels, and up-regulating Nrf2 expression, which led to increased downstream target proteins such as NADPH-dehydrogenase-quinone-1 (NQO1), heme oxygenase 1 (HO-1), glutathione peroxidase 1 (Gpx1), and catalase (CAT) under high glucose condition (Wang et al., 2015). It has been revealed that cinnamon bark aqueous extract at 200 mg/kg/day, for 14 days induced total antioxidant capacity by elevating serum TAC and total oxidant status (TOS) in mice and ameliorated the intensity of tissue damage induced by acetaminophen (APAP, 200 mg/kg) (Hussain et al., 2019). Similarly, in another study, cinnamon oil at 100 and 200 mg/kg b.w. for 14 days showed antioxidant activities via diminishing MDA levels, increasing

GSH levels, and enhancing antioxidant enzymes (CAT, SOD, glutathione reductase (GR), and GPx) activities (Hussain et al., 2020). Also, treatment with 200 mg/kg cinnamon aqueous extract for 14 days before gamma irradiation led to antioxidant effects and limited levels of tumor necrosis factor-alpha (TNF- $\alpha$ ) and liver NO, as well as inducing SOD activity in rats (weighing 100–120 g) (Azab et al., 2011). According to Atsamo et al. (2021), 200 mg/kg and 400 mg/kg of aqueous extract of *C. zeylanicum* for 14 consecutive days considerably reduced oxidative stress in nephropathic mice. The proposed antioxidant mechanism for the aqueous extract of *C. zeylanicum* is the reduction of oxidative stress markers including TNF- $\alpha$ , interleukin (IL)-6, and IL-1 $\beta$ , and enhancement of antioxidant parameters such as SOD, GSH, and CAT (Atsamo et al., 2021). Cinnamaldehyde significantly affects cancer signaling pathways. In an *in vivo* experiment, human colon cancer cells were treated with 75 $\mu$ M of cinnamaldehyde showed antioxidant effects (Petrocelli et al., 2021). Similarly, Shalaby and coworkers claimed that the treatment of animals with an aqueous extract of cinnamon (200, 400 mg/kg for 6 weeks) enhanced antioxidant activity in obese diabetic rats (Shalaby and Saifan, 2014). In another study, cinnamon administration (50 mg/kg for 7 days) stimulated the Nrf2 pathway, leading to induced levels of antioxidant

enzymes. It should be mentioned that cinnamon significantly suppressed the expression of MDA, IL-1 $\beta$ , and caspase-3 in lipopolysaccharide (LPS)-induced mice with neuroinflammation (El-ezz et al., 2018). Also, we found that hydro-alcohol extract and essential oil of *C. verum* and *C. cassia* along with its main bioactive component cinnamaldehyde, exhibited antioxidant activity via ROS scavenging in 6-OHDA-exposed PC12 cells, which serve as an in vitro model of Parkinson's disease (Ramazani et al., 2020).

Nrf2, a transcription factor, plays a crucial role in defending cells and tissues against oxidative stress by regulating the expression of antioxidant genes, including heme oxygenase 1 (HO-1) (Bendavit et al., 2016). Evaluation of the ethanolic extract and trans-cinnamic aldehyde (cinnamaldehyde) prepared from *C. cassia* bark on HCT116 and HT29 (human colon cancer cells) and FHC (non-immortalized primary fetal colon cells) demonstrated upregulated cellular Nrf2 protein levels, leading to increased HO-1 expression as part of the antioxidant response element (Wondrak et al., 2010).

A recent study has pointed out that cinnamon essential oil displayed significant antioxidant properties on human cell lines HeLa (0.125–4  $\mu\text{g/mL}$ ) and Raji (12–0.375  $\mu\text{g/mL}$ ) when compared with synthetic antioxidant BHT (butylated hydroxytoluene) and ascorbic acid (vitamin C) (Kallel et al., 2019). In addition, cinnamon essential oil showed cytotoxic activity toward PC-3, A549, and MCF-7 human tumor cell lines with concentrations of (0.012, 0.017, 0.076 v/v) (Zu et al., 2010). Pretreatment of BV2 microglial cells with essential oil from *C. camphora* (Linn.) for 48 h at different concentrations (100, 150, 200  $\mu\text{g/mL}$ ) caused a reduction of NO as well as to measure of cytokines including IL-1 $\beta$ , IL-6, and IL-18, induced by LPS (Chen, Tang et al., 2020). Overall, it seems that cinnamon chemical compositions reveal their antioxidant effects through lessening ROS levels, increasing SOD, GSH-Px, CAT, HO-1 and NQO1 activity, decreasing the MDA and TAC level. Additionally, the antioxidant effect of cinnamon leads to diminution in caspase-3, caspase-9, TNF- $\alpha$ , and NF- $\kappa\text{B}$  levels and enhancement of Erk1/2, Akt1, and JNK levels.

#### *Anti-inflammatory effects of cinnamon*

Inflammation is a vital defense mechanism that protects injured or infected cells *via* eliminating injurious factors and promoting the healing of damaged tissues.

Pro-inflammatory mediators, including cytokines such as TNF- $\alpha$ , IL-1, IL-6, IL-10, and type 1 interferons (IFNs), induce a cellular inflammatory process (Lawrence, 2009). IL-1 stimulates mRNA expression and protein synthesis of inducible nitric oxide synthase (iNOS), cyclooxygenase (COX-2), and phospholipase A2 (PLA2), which promote the formation of NO, platelet-activating factor, and prostaglandin E2 (PGE2) (Kany et al., 2019). NF- $\kappa\text{B}$  is an important regulator of inflammatory responses, activated by IL-1 $\beta$  and TNF- $\alpha$  (Somade et al., 2019). The NF- $\kappa\text{B}$  represents a family of transcription factors involved in regulating the expression of hundreds of genes, especially those related to inflammatory and immune responses. In unstimulated cells, NF- $\kappa\text{B}$  dimers exist in the cytoplasm in an inactive form bound to inhibitor of nuclear factor kappa B (I- $\kappa\text{B}$ ). Upon degradation of I- $\kappa\text{B}$ , NF- $\kappa\text{B}$  translocates into the nucleus, where it binds to DNA consensus sequences on gene promoters (Giuliani et al., 2018). Due to the increasing prevalence of diseases associated with inflammation, scientists are increasingly investing in bioactive compounds with potential biological activities found in traditional herbal medicine.

#### *Clinical studies*

Based on the results, 3 g/day cinnamon supplementation for 8 weeks did not significantly reduce NF- $\kappa\text{B}$ , IL-6, and TNF- $\alpha$  levels in patients with type 2 diabetes. However, it did show a significant reduction in NF- $\kappa\text{B}$  levels between groups (Davari et al., 2020).

#### *In vivo and in vitro studies*

Interestingly, administration of cinnamon extract at both 250 and 500 mg/kg/day for 28 days to acrylamide-intoxicated rats markedly augmented serum and liver TAC levels and lowered liver and serum MDA levels. It also ameliorated inflammatory responses *via* a reduction in the leptin serum levels (one of the major adipocytokines) and TNF- $\alpha$  (Haidari et al., 2020). It has been documented that oral administration of 20, 100, and 500 mg/kg of cinnamon water extract exhibited anti-inflammatory activities *in vivo* and *in vitro* LPS-induced models. After 6 days of oral administration, the serum levels of IL-6 and TNF- $\alpha$  significantly decreased in mice. Moreover, cinnamon water extract significantly reduced mRNA expression of TNF- $\alpha$ , suppressed I- $\kappa\text{B}\alpha$  degradation, and inhibited the activation

of extracellular signal-regulated kinases 1/2 (ERK1/2), c-Jun N-terminal kinases (JNK), and p38 in peritoneal macrophages from glycollate-injected mice (Hong et al., 2012). In 2019, Vallianou and collaborators reported that *C. Zeylanicum* supplementation enhanced serum C-reactive protein levels (Vallianou et al., 2019). C-reactive protein is an acute-phase inflammatory protein, and its increased levels are observed during the early stages of several chronic conditions including type 2 diabetes mellitus, pre-diabetes, obesity, coronary artery disease, rheumatoid arthritis, and nonalcoholic fatty liver disease (Taghizadeh et al., 2019). In a study by Vet al. the anti-inflammatory of TAPP of the bark of *C. zeylanicum* in rats was reported. Based on the result, administration 8 mg/kg, p.o., of TAPP daily (day-12 to day-21) significantly showed anti-inflammatory and anti-arthritis activities in adjuvant induced established arthritis (AIA) model. (Vet al., 2013). Besides, it was shown that *C. zeylanicum* extract (150 mg/kg/day for 10 days) treatment had antioxidant and anti-inflammatory effects in colonic homogenate of colitis animals. It decreased the activity of myeloperoxidase (MPO) and the levels of nitric oxide (NO), and significantly reduced the expression of various cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-6) and inflammatory enzymes (COX-2) (Salamatian et al., 2019). Acetaminophen (APAP) can stimulate inflammatory cytokines such as IL-1 $\beta$  and IL-6, as well as activate caspase-3 and -9, thereby increasing liver injury (Das et al., 2010; Woolbright and Jaeschke, 2018; Wu et al., 2019). cinnamon oil at doses of 50, 100, and 200 mg/kg b.w. for 14 days exerted anti-inflammatory effects by decreasing IL-1 $\beta$ , IL-6 levels, and caspase-3 and -9 expressions in APAP-induced rats (170–220 g), suggesting it could be an alternative treatment for liver damage (Hussain et al., 2020). Recent research reported that inhalation of cinnamon oil (6.55  $\mu$ L/L) restrained inflammatory airway hyperresponsiveness and reduced some of the cellular inflammatory markers in pneumonitis mice (Csikós et al., 2020). Furthermore, Li et al. reported the treatment of atherosclerotic mice with 5, 10, 20 mg/kg of cinnamaldehyde for 8 weeks reduced the phosphorylation level of I $\kappa$ B $\alpha$  and NF- $\kappa$ B, and significantly decreased inflammatory cytokines (TNF- $\alpha$ , IL-6, ROS, and monocyte chemoattractant protein-1 (MCP-1)) (Li et al., 2019). Given the influence of NF- $\kappa$ B activation on inflammation and ROS, it has been suggested that the anti-inflammatory process is sometimes con-

comitant with antioxidant activities (Flohé et al., 1997). Wondrak and colleagues revealed that cinnamon acts as an Nrf2 activator in cultured human colon cells, reducing the risk of colon cancer (Wondrak et al., 2010).

Kim et al. evaluated the effects of *C. japonicum* Sieb. subcritical water extract on anti-inflammation in the RAW264.7 and Caco-2 co-culture model of cellular intestinal inflammation. The results displayed that cinnamon subcritical water extract (CSWE) significantly reduced nitrite, PGE2, IL-6, IL-8, TNF- $\alpha$  levels, and NF- $\kappa$ B activity. They concluded that cinnamaldehyde (the major components of *C. japonicum*) and cinnamic acid suppressed the phosphorylation of factors in the NF- $\kappa$ B pathway (Kim and Kim, 2019). The evaluation of *C. camphora* extracts effects on RAW264.7 cells demonstrated the anti-inflammatory properties of methanol (MeOH), hexane, and ethyl acetate (EtOAc) extracts (100  $\mu$ g/ml) through suppressing the production of interleukins IL-1 $\beta$ , IL-6, and TNF- $\alpha$ . Additionally, treatment with hexane and EtOAc extracts (100  $\mu$ g/ml) induced inhibitory effects on NO production, while EtOAc and n-butanol (BuOH) extracts strongly inhibited PGE2 production in LPS/ IFN- $\gamma$ -activated macrophages. EtOAc and BuOH extracts also have antiplatelet effects when tested by the DPPH and xanthine oxide assays (Lee et al., 2006). The anti-inflammation activities of essential oil and its constituents from indigenous *C. osmophloeum* twigs in LPS-stimulated RAW 264.7 cells were evaluated by Tung et al. The results showed the inhibitory effect of *C. osmophloeum* twig essential oil on NO and PGE2 production at concentrations of 25  $\mu$ g/ml and 10  $\mu$ g/ml, respectively (Tung et al., 2008). Similar to these results, Lee et al. reported that *C. cassia* extract significantly reduced NO production with an IC<sub>50</sub> value between 0.1-10  $\mu$ g/ $\mu$ L (Lee et al., 2002). *Cinnamomum zeylanicum* and *C. cassia* extracts exhibited anti-inflammatory effects via decreasing LPS+IFN- $\gamma$  induced NO and TNF- $\alpha$  production in RAW 264.7 and J774A.1 macrophages (Gunawardena et al., 2015). Yu et al. confirmed that *C. cassia* ethanol extract (100  $\mu$ g/mL) suppressed NF- $\kappa$ B activation, which resulted in the suppression of iNOS, COX-2, and TNF- $\alpha$  mRNA expression in LPS-activated RAW264.7 cells and peritoneal macrophages. Also, they observed lowered levels of NO, TNF- $\alpha$ , and PGE2 production (Yu et al., 2012). In addition, following treatment with the fruit essential oil of *C. insularimontanum*, inhibitory

effects on the protein expression of I $\kappa$ B kinase, iNOS, and nuclear NF- $\kappa$ B reduction were documented (Lin et al., 2008). Similarly, TAPP isolated from *C. zeylanicum* bark can exhibit anti-inflammatory effects *via* inhibiting COX enzyme activity, and reducing the production of pro-inflammatory cytokines such as IL-1 $\beta$  and TNF- $\alpha$  (Ping et al., 2010; Miguel, 2010). Anti-inflammatory activities of cinnamaldehyde, eugenol, and terpene in cinnamon have been shown in various studies (Hsueh and Law, 1998; Lee et al., 2002; Penckofer, et al., 2002). New findings revealed that cinnamaldehyde can exert anti-inflammatory properties by activating peroxisome proliferator-activated receptors (PPARs), which results in attenuating NF- $\kappa$ B functioning (Miguel, 2010; Huang et al., 2011). Following treatment of a dermal fibroblast cell system, HDF3CGF, which serves as a human skin disease model, with 0.0012% v/v of *C. zeylanicum* bark essential oil, several inflammatory protein biomarkers like IFN- $\gamma$ -induced protein 10, monocyte chemoattractant protein-1, interferon-inducible T-cell alpha chemoattractant, and monokine induced by gamma interferon (MIG) were significantly diminished (Han and Parker, 2017). Pre-treating Raw cells with cinnamaldehyde (20 and 50 mM) for 1 hour disrupts the toll-like receptor 4 (TLR4) oligomerization process induced by LPS. Based on the results, cinnamaldehyde significantly reduces NF- $\kappa$ B induced by pro-inflammatory stimuli and prevents downstream gene transcription such as COX-2 and IFN $\beta$  (Youn et al., 2008). In 2018, Schink and colleagues found that cinnamon extract exerts anti-inflammatory effects (25  $\mu$ g/ml) against toll-like receptors (TLR4, TLR2) and mitigates the phosphorylation of Akt and I $\kappa$ B $\alpha$  in an *in vitro* model of THP-1 monocytes stimulated with LPS (Schink et al., 2018). Chen and colleagues had evaluated the effects of cinnamic aldehyde on I $\kappa$ B protein phosphorylation in NF- $\kappa$ B signaling pathway in the LPS-induced human osteoarthritis chondrocytes cells. The authors reported that different concentrations of cinnamic aldehyde (10, 20, and 50  $\mu$ M) for 24 h significantly downregulated the IL-1 $\beta$ , IL-6, TNF- $\alpha$  expression (Chen, Ruan et al., 2020). Overall, cinnamon chemical compositions can display anti-inflammatory properties *via* reducing TNF- $\alpha$ , IL-1 $\beta$ , IL-1, IL-2, IL-6, IL-8, COX-2, and PGE2 levels, and decreasing the I $\kappa$ B $\alpha$  phosphorylation and NO production through NF- $\kappa$ B activity disruption.

### *Covid-19 and cinnamon*

In recent years, the world has experienced outbreaks of coronavirus with a high mortality rate causing viral pneumonia (Ramazani et al., 2021). COVID-19 is caused by the SARS-CoV-2 family of coronaviruses with singular stranded RNA. Its shape is round or elliptic enveloped by special spike proteins required for attachment, and its diameter is about 60-140 nm (Weiss and Navas-Martin, 2005; Aspland et al., 2021). It has been reported that COVID-19 infection releases a considerable amount of inflammatory cytokines such as IL-1 $\beta$ , IL-2, IL-6, IL-8, TNF- $\alpha$ , leading to long-term neurological disturbances, kidney, and myocardial disorders, as well as chronic fatigue (Berlin et al., 2020; Heneka et al., 2020; Mitrani et al., 2020; Pelaia et al., 2020). In several cases, SARS-CoV-2 has been shown to cause a reduction in NO expression which leads to diffuse pneumonia and organs failure (Taneja, 2020). Few studies have been published on the immune system performance against COVID-19.

Cinnamon has shown inhibitory effects on covid-19 infection by interrupting TLR2 and TLR4, reducing pro-inflammatory cytokines, and preventing NF- $\kappa$ B/activator protein 1 (AP-1) signaling (Yakhchali et al., 2021). It has been demonstrated that cinnamon plant extract, which contains hydroxyl group, can deactivate the active components of the virus, specifically proteins and RNA, through an esterification process involving amino (-NH<sub>2</sub>) and carboxyl (-COOH) groups (Taneja, 2020). *in vitro* experiments have shown that cinnamon extract exhibits dose-dependent anti-viral activity against wild-type SARS-CoV-2, likely by disrupting viral endocytic pathways and viral replication (Zhuang et al., 2009). Altogether, these studies suggest potential roles for cinnamon against SARS-CoV-2 infection.

### *Other Uses*

Based on the numerous studies, Cinnamon exhibited various beneficial pharmacological and therapeutic properties for centuries (Ekor, 2014). This plant, which belongs to the Lauraceae family, has about 250 species spread across Australia, India, and China (Jayaprakasha et al., 2003). *Cinnamomum verum* essential Oil was used to treat neurological diseases, sinusitis, and colds (Jayaprakasha et al., 2003). Cinnamon bark is commonly used as spices and in tea to prevent the common cold

and cardiovascular diseases (Ranasinghe et al., 2013). Traditionally, it has been applied to relieve muscular pain, typhoid fever, and gastrointestinal diseases (Jazet Dongmo et al., 2007). Several studies have reported the use of cinnamon bark and its oil for treating toothaches and dental issues (Aneja et al., 2009; Gupta et al., 2011). Moreover, cinnamon has been mentioned in various sources as a remedy for blood circulation disorders, inflammatory disturbances, and digestive problems (Tanaka et al., 1989; Yu et al., 2007; Csikós et al., 2020).

#### *Strengths and limitations*

Numerous studies have evaluated the pharmacological and therapeutic effects of cinnamon, providing new insights into its molecular mechanisms and biological activities, such as antioxidant, anti-inflammatory, anti-diabetic, and antibacterial properties.

The potential therapeutic impacts of cinnamon on human health have been reported in a few articles. However, the effects of cinnamon on coronavirus infection and inflammation is not clear yet. Further research is needed to elucidate the biological molecular mechanisms and pharmacological activities of cinnamon comprehensively.

## Conclusion

In recent decades, extensive research has explored the phototherapeutic properties of cinnamon and its compounds, uncovering explicit and implicit anti-inflammatory and antioxidant effects. This study reviews cinnamon's molecular mechanisms in relation to coronavirus infections, highlighting its advantageous therapeutic potential for advancing clinical trials.

Cinnamon exerts antioxidant effects by reducing ROS through the induction of CAT, GSH-Px, HO-1 and NQO1 activities, while decreasing MDA and TAC levels. It also suppresses TNF- $\alpha$  and NF- $\kappa$ B gene expression, and enhances SOD as well as CAT mRNA levels. Additionally, cinnamon's antioxidant effect results in decreased levels of caspase-3 and caspase-9, while enhancing Erk1/2, Akt1, and JNK levels.

Furthermore, cinnamon's complex chemical composition demonstrates anti-inflammatory properties by reducing pro-inflammatory cytokines TNF- $\alpha$ , IL-1 $\beta$ , IL-1, IL-2, IL-6, IL-8, COX-2, and PGE2, and by suppressing I $\kappa$ B $\alpha$  phosphorylation and nitric oxide (NO) production through disruption of NF- $\kappa$ B activity. Cinnamon's abil-

ity to inactivate the NF- $\kappa$ B pathway via interruption of TLR4 and TLR2 underscores its pharmacological potential for therapeutic research against viral infections.

Experimental studies have indicated effective doses of cinnamon ranging from 110-500 mg/kg in animals and 10-100 mM in cell-based tests. However, while clinical studies have shown some efficacy, larger-scale investigations are needed to confirm these effects.

## Conflict of interest

The authors declare that there are no conflicts of interest.

## Acknowledgement

The authors wish to thank the Yazd University, Yazd, Iran.

## References

- Anderson R A. Chromium and polyphenols from cinnamon improve insulin sensitivity: plenary lecture. Proceedings of the Nutrition Society 2008; 67: 48-53. <https://doi.org/10.1017/S0029665108006010>
- Aneja K R, Joshi R, Sharma C. Antimicrobial activity of Dalchini (*Cinnamomum zeylanicum* bark) extracts on some dental caries pathogens. Journal of Pharmacy Research 2009; 2: 1387-1390.
- Aspland A M, Douagi I, Filby A, Jellison E R, Martinez L, Shinko D, et al. Biosafety during a pandemic: shared resource laboratories rise to the challenge. Cytometry Part A 2021; 99: 68-80. <https://doi.org/10.1002/cyto.a.24280>
- Atsamo A D, Lontsie Songmene A, Metchi Donfack M F, Ngouateu O B, Nguielefack TB, Dimo T. Aqueous Extract from *Cinnamomum zeylanicum* (Lauraceae) Stem Bark Ameliorates Gentamicin-Induced Nephrotoxicity in Rats by Modulating Oxidative Stress and Inflammatory Markers. Evidence-based Complementary and Alternative Medicine 2021; 2021: 1-12. <https://doi.org/10.1155/2021/5543889>
- Azab K S, Mostafa A-H A, Ali E M, Abdel-Aziz M A. Cinnamon extract ameliorates ionizing radiation-induced cellular injury in rats. Ecotoxicology and Environmental Safety 2011; 74: 2324-2329. <https://doi.org/10.1016/j.ecoenv.2011.06.016>
- Bendavit G, Aboukassim T, Hilmi K, Shah S, Batist G. Nrf2 transcription factor can directly regulate mTOR: linking cytoprotective gene expression to a major metabolic regulator that generates redox activity. Journal of Biological Chemistry 2016; 291: 25476254-25476288. <https://doi.org/10.1074/jbc.M115.014000>

- org/10.1074/jbc.M116.760249
- Berlin D A, Gulick R M, Martinez F J. Severe covid-19. The New England Journal of Medicine 2020; 383: 2451-2460. <https://doi.org/10.1056/NEJMcp2009575>
- Borzoei A, Rafrat M, Niromanesh S, Farzadi L, Narimani F, Doostan F. Effects of cinnamon supplementation on antioxidant status and serum lipids in women with polycystic ovary syndrome. Journal of Traditional and Complementary Medicine 2018; 8: 128-133. <https://doi.org/10.1016/j.jtcme.2017.04.008>
- Chan K W, Khong N M H, Iqbal S, Ch'Ng SE, Younas U, Babji A S. Cinnamon bark deodorised aqueous extract as potential natural antioxidant in meat emulsion system: a comparative study with synthetic and natural food antioxidants. Journal of Food Science and Technology 2014; 51: 3269-3276. <https://doi.org/10.1007/s13197-012-0818-5>
- Chen P, Ruan A, Zhou J, Huang L, Zhang X, Ma Y, et al. Cinnamic aldehyde inhibits lipopolysaccharide-induced chondrocyte inflammation and reduces cartilage degeneration by blocking the nuclear factor-kappa B signaling pathway. Frontiers in Pharmacology 2020;11: 1-9. <https://doi.org/10.3389/fphar.2020.00949>
- Chen J, Tang C, Zhou Y, Zhang R, Ye S, Zhao Z, et al. Anti-inflammatory property of the essential oil from *cinnamomum camphora* (Linn.) presl leaves and the evaluation of its underlying mechanism by using metabolomics analysis. Molecules 2020; 25:1-13. <https://doi.org/10.3390/molecules25204796>
- Csikós E, Cseko K, Ashraf AR, Kemény Á, Kereskai L, Kocsis B, et al. Effects of *Thymus vulgaris* L., *Cinnamomum verum* J.Presl and *Cymbopogon nardus* (L.) rendle essential oils in the endotoxin-induced acute airway inflammation mouse model. Molecules 2020; 25: 3553-3566. <https://doi.org/10.3390/molecules25153553>
- Das J, Ghosh J, Manna P, Sil PC. Taurine protects acetaminophen-induced oxidative damage in mice kidney through APAP urinary excretion and CYP2E1 inactivation. Toxicology 2010; 269: 24-34. <https://doi.org/10.1016/j.tox.2010.01.003>
- Dassanayake M, Larsen K. A revised handbook to the Flora of Ceylon. Nordic Journal of Botany. 1996; 16: 660. <https://doi.org/10.1111/j.1756-1051.1996.tb00284.x>
- Davari M, Hashemi R, Mirmiran P, Hedayati M, Sahranavard S, Bahreini S, et al. Effects of cinnamon supplementation on expression of systemic inflammation factors, NF-kB and Sirtuin-1 (SIRT1) in type 2 diabetes: A randomized, double blind, and controlled clinical trial. Nutrition Journal 2020; 19: 1-8. <https://doi.org/10.1186/s12937-019-0518-3>
- Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Frontiers in Pharmacology 2014; 4: 177-186. <https://doi.org/10.3389/fphar.2013.00177>
- Elshafie M M, Nawar I A, Algamal M A, Ahmad S M. Evaluation of the biological effects for adding cinnamon volatile oil and TBHQ as antioxidant on rats' lipid profiles. Asian Journal of Plant Sciences 2012; 11: 100-108. <https://doi.org/10.3923/ajps.2012.100.108>
- El-ezz A, Maher A, Sallam N, El-Brairy A, Kenawy S. Trans-cinnamaldehyde modulates hippocampal Nrf2 factor and inhibits amyloid beta aggregation in LPS-induced neuroinflammation mouse model. Neurochemical Research 2018; 43: 2333-2342. <https://doi.org/10.1007/s11064-018-2656-y>
- Ervina M, Nawu Y, Esar S. Comparison of in vitro antioxidant activity of infusion, extract and fractions of Indonesian Cinnamon (*Cinnamomum burmannii*) bark. International Food Research Journal 2016; 23: 1346-1350.
- Flohé L, Brigelius-Flohé R, Saliou C, Traber MG, Packer L. Redox regulation of NF-kappa B activation. Free Radical Biology and Medicine 1997; 22: 1115-1126. [https://doi.org/10.1016/S0891-5849\(96\)00501-1](https://doi.org/10.1016/S0891-5849(96)00501-1)
- Ghosh S, Karin M. Missing pieces in the NF-kB puzzle. Cell 2002; 109: 81-96. [https://doi.org/10.1016/S0092-8674\(02\)00703-1](https://doi.org/10.1016/S0092-8674(02)00703-1)
- Gilgun-sherki Y, Melamed E, Offen D. Oxidative stress induced-neurodegenerative diseases: the need for antioxidants that penetrate the blood brain barrier. Neuropharmacology 2001; 40: 959-975. [https://doi.org/10.1016/S0028-3908\(01\)00019-3](https://doi.org/10.1016/S0028-3908(01)00019-3)
- Giuliani C, Bucci I, Napolitano G. The role of the transcription factor Nuclear Factor-kappa B in thyroid autoimmunity and cancer. Frontiers in Endocrinology 2018; 9: 471-478. <https://doi.org/10.3389/fendo.2018.00471>
- Gunawardena D, Karunaweera N, Lee S, Van Der Kooy F, Harman DG, Raju R, et al. Anti-inflammatory activity of cinnamon (*C. zeylanicum* and *C. cassia*) extracts - Identification of E-cinnamaldehyde and o-methoxy cinnamaldehyde as the most potent bioactive compounds. Food & Function 2015; 6: 910-919. <https://doi.org/10.1039/C4FO00680A>
- Gupta C, Garg A P, Prakash D, Goyal S, Gupta S. Comparative study of cinnamon oil and clove oil on some oral microbiota. Acta Biomedica 2011; 82: 197-199.
- Haidari F, Mohammadshahi M, Abiri B, Zarei M, Fathi M.

- Cinnamon extract supplementation improves inflammation and oxidative stress induced by acrylamide: An experimental animal study. *Avicenna Journal of Phytomedicine* 2020; 10: 243-252.
- Han X, Parker T L. Antiinflammatory activity of cinnamon (*Cinnamomum zeylanicum*) bark essential oil in a human skin disease model. *Phytotherapy Research* 2017; 31: 1034-1038. <https://doi.org/10.1002/ptr.5822>
- Hao X, Sun W, Ke C, Wang F, Xue Y, Luo Z, et al. Anti-inflammatory activities of leaf oil from *Cinnamomum subavenium* in vitro and in vivo. *BioMed Research International* 2019; 2019. <https://doi.org/10.1155/2019/1823149>
- Heneka M T, Golenbock D, Latz E, Morgan D, Brown R. Immediate and long-term consequences of COVID-19 infections for the development of neurological disease. *Alzheimer's Research & Therapy* 2020; 12: 1-3. <https://doi.org/10.1186/s13195-020-00640-3>
- Hong J-W, Yang G-E, Kim YB, Eom S H, Lew J-H, Kang H. Anti-inflammatory activity of cinnamon water extract in vivo and in vitro LPS-induced models. *BMC Complementary Medicine and Therapies* 2012; 12: 1-8. <https://doi.org/10.1186/1472-6882-12-237>
- Hsueh W A, Law R E. Cardiovascular risk continuum: implications of insulin resistance and diabetes. *American Journal of Medicine* 1998; 105: 4-14. [https://doi.org/10.1016/S0002-9343\(98\)00205-8](https://doi.org/10.1016/S0002-9343(98)00205-8)
- Huang B, Yuan H D, Kim D Y, Quan H Y, Chung S H. Cinnamaldehyde prevents adipocyte differentiation and adipogenesis via regulation of peroxisome proliferator-activated receptor- $\gamma$  (PPAR $\gamma$ ) and AMP-activated protein kinase (AMPK) pathways. *Journal of Agricultural and Food Chemistry* 2011; 59: 3666-3673. <https://doi.org/10.1021/jf104814t>
- Hussain Z, Khan J A, Arshad A, Asif P, Rashid H, Arshad M I. Protective effects of *Cinnamomum zeylanicum* L. (Darchini) in acetaminophen-induced oxidative stress, hepatotoxicity and nephrotoxicity in mouse model. *Biomedicine & Pharmacotherapy* 2019; 109: 2285-2292. <https://doi.org/10.1016/j.biopha.2018.11.123>
- Hussain S, Ashafaq M, Alshahrani S, Siddiqui R, Ahmed R A, Khuwaja G, et al. Cinnamon oil against acetaminophen-induced acute liver toxicity by attenuating inflammation, oxidative stress and apoptosis. *Toxicology Reports* 2020; 7: 1296-1304. <https://doi.org/10.1016/j.toxrep.2020.09.008>
- Jayaprakasha G K, Jagan Mohan Rao L, Sakariah K K. Volatile constituents from *Cinnamomum zeylanicum* fruit stalks and their antioxidant activities. *Journal of Agricultural and Food Chemistry* 2003; 51: 4344-4348. <https://doi.org/10.1021/jf034169i>
- Jayaprakasha G K, Ohnishi-Kameyama M, Ono H, Yoshida M, Rao L J. Phenolic constituents in the fruits of *Cinnamomum zeylanicum* and their antioxidant activity. *Journal of Agricultural and Food Chemistry* 2006; 54: 1672-1679. <https://doi.org/10.1021/jf052736r>
- Jazet Dongmo PM, Tatsadjieu LN, Tchoumboungang F, Sameza ML, Dongnio BN, Amvam Zollo PH, et al. Chemical composition, antiradical and antifungal activities of essential oil of the leaves of *cinnamomum zeylanicum* blume from Cameroon. *Natural Product Communications* 2007; 2: 1287-1290. <https://doi.org/10.1177/1934578X0700021219>
- Jindarat S, Muangnoi C, Changsiriporn D, Platong A, Thanamontra B, Chiewchanwit D, Vongvanvatana V, Rongrungsri N, Krittasilp K, Kaewkong N, Kantawan S. Efficacy and safety of cinnamon stomachic mixture for patients with functional dyspepsia. *Siriraj Medical Journal* 2006; 58: 1103-1106.
- Kallel I, Hadrich B, Gargouri B, Chaabane A, Lassoued S, Gdoura R, et al. Optimization of cinnamon (*Cinnamomum zeylanicum* Blume) essential oil extraction: evaluation of antioxidant and antiproliferative effects. *Evidence-Based Complementary and Alternative Medicine* 2019; 2019. <https://doi.org/10.1155/2019/6498347>
- Kany S, Vollrath J T, Relja B. Cytokines in inflammatory disease. *International Journal of Molecular Sciences* 2019; 20: 6008-39. <https://doi.org/10.3390/ijms20236008>
- Kim M S, Kim J Y. Cinnamon subcritical water extract attenuates intestinal inflammation and enhances intestinal tight junction in a Caco-2 and RAW264. 7 co-culture model. *Food C Function* 2019; 10: 4350-4360. <https://doi.org/10.1039/C9FO00302A>
- Lawrence T. The nuclear factor NF- $\kappa$ B pathway in inflammation. *Cold Spring Harbor Perspectives in Biology* 2009; 1: a001651. <https://doi.org/10.1101/cshperspect.a001651>
- Lee S K, Hong C-H, Huh S-K, Kim S-S, Oh O-J, Min H-Y, et al. Suppressive effect of natural sesquiterpenoids on inducible cyclooxygenase (COX-2) and nitric oxide synthase (iNOS) activity in mouse macrophage cells. *Journal of Environmental Pathology, Toxicology and Oncology* 2002; 21: 141-148. <https://doi.org/10.1615/JEnvironPatholToxicolOncol.v21.i2.70>
- Lee H-S, Kim B-S, Kim M-K. Suppression effect of *Cinnamomum cassia* bark-derived component on nitric oxide synthase. *Journal of Agricultural and Food Chemistry* 2002; 50: 7700-7703. <https://doi.org/10.1021/jf020751f>

- Lee H J, Hyun E-A, Yoon W J, Kim B H, Rhee M H, Kang H K, et al. In vitro anti-inflammatory and anti-oxidative effects of *Cinnamomum camphora* extracts. *Journal of Ethnopharmacology* 2006; 103: 208-216. <https://doi.org/10.1016/j.jep.2005.08.009>
- Li W, Zhi W, Zhao J, Li W, Zang L, Liu F, et al. Cinnamaldehyde attenuates atherosclerosis: Via targeting the I $\kappa$ B/NF- $\kappa$ B signaling pathway in high fat diet-induced ApoE<sup>-/-</sup> mice. *Food & Function* 2019; 10: 4001-4009. <https://doi.org/10.1039/C9FO00396G>
- Lin C-T, Chen C-J, Lin T-Y, Tung J C, Wang S-Y. Anti-inflammatory activity of fruit essential oil from *Cinnamomum insularimontanum* Hayata. *Bioresource Technology* 2008; 99: 8783-8787. <https://doi.org/10.1016/j.biortech.2008.04.041>
- Mahmoodnia L, Aghadavod E, Rafieian-Kopaei M. Ameliorative impact of cinnamon against high blood pressure; an updated review. *Journal of Renal Injury Prevention* 2017; 6: 171-176. <https://doi.org/10.15171/jrip.2017.33>
- Mathew S, Abraham T E. In vitro antioxidant activity and scavenging effects of *Cinnamomum verum* leaf extract assayed by different methodologies. *Food and Chemical Toxicology* 2006; 44: 198-206. <https://doi.org/10.1016/j.fct.2005.06.013>
- Miguel M G. Antioxidant and anti-inflammatory activities of essential oils: a short review. *Molecules* 2010; 15: 9252-9287. <https://doi.org/10.3390/molecules15129252>
- Mitrani R D, Dabas N, Goldberger J J. COVID-19 cardiac injury: Implications for long-term surveillance and outcomes in survivors. *Heart Rhythm* 2020; 17: 1984-1990. <https://doi.org/10.1016/j.hrthm.2020.06.026>
- Noori S, Azmat M, Mahboob T. Study on antioxidant effects of cinnamon and garlic extract in liver, kidney and heart tissue of rat. *Bioscience Research* 2012; 9: 17-22.
- Pelaia C, Tinello C, Vatrella A, De Sarro G, Pelaia G. Lung under attack by COVID-19-induced cytokine storm: pathogenic mechanisms and therapeutic implications. *Therapeutic Advances in Respiratory Disease* 2020; 14: 1-9. <https://doi.org/10.1177/1753466620933508>
- Penckofer S, Schwartz D, Florczak K. Oxidative stress and cardiovascular disease in type 2 diabetes: the role of antioxidants and pro-oxidants. *Journal of Cardiovascular Nursing* 2002; 16: 68-85. <https://doi.org/10.1097/00005082-200201000-00007>
- Peng X, Cheng K W, Ma J, Chen B, Ho C T, Lo C, et al. Cinnamon bark proanthocyanidins as reactive carbonyl scavengers to prevent the formation of advanced glycation endproducts. *Journal of Agricultural and Food Chemistry* 2008; 56: 1907-1911. <https://doi.org/10.1021/jf073065v>
- Petrocelli G, Farabegoli F, Valerii M C, Giovannini C, Sardo A, Spisni E. Molecules present in plant essential oils for prevention and treatment of colorectal cancer (Crc). *Molecules* 2021; 26: 1-12. <https://doi.org/10.3390/molecules26040885>
- Ping H, Zhang G, Ren G. Antidiabetic effects of cinnamon oil in diabetic KK-Ay mice. *Food and Chemical Toxicology* 2010; 48: 2344-2349. <https://doi.org/10.1016/j.fct.2010.05.069>
- Plata-Rueda A, Campos J M, da Silva Rolim G, Martínez L C, Dos Santos M H, Fernandes FL, et al. Terpenoid constituents of cinnamon and clove essential oils cause toxic effects and behavior repellency response on granary weevil, *Sitophilus granarius*. *Ecotoxicology and Environmental Safety*. 2018; 156: 263-270. <https://doi.org/10.1016/j.ecoenv.2018.03.033>
- Prakash B, Singh P, Yadav S, Singh S C, Dubey N K. Safety profile assessment and efficacy of chemically characterized *Cinnamomum glaucescens* essential oil against storage fungi, insect, aflatoxin secretion and as antioxidant. *Food and Chemical Toxicology* 2013; 53: 160-167. <https://doi.org/10.1016/j.fct.2012.11.044>
- Ramazani E, YazdFazeli M, Emami S A, Mohtashami L, Javadi B, Asili J, et al. Protective effects of *Cinnamomum verum*, *Cinnamomum cassia* and cinnamaldehyde against 6-OHDA-induced apoptosis in PC12 cells. *Molecular Biology Reports* 2020; 47: 2437-2445. <https://doi.org/10.1007/s11033-020-05284-y>
- Ramazani E, Emami S A, Tayarani-Najaran N, Sahebkar A, Tayarani-Najaran Z. Antiviral plants in view of avicenna's the canon of medicine and modern medicine against common cold. *Natural Products and Human Diseases: Pharmacology, Molecular Targets, and Therapeutic Benefits*. 2021: 99-121. [https://doi.org/10.1007/978-3-030-73234-9\\_7](https://doi.org/10.1007/978-3-030-73234-9_7)
- Ranasinghe P, Pigera S, Premakumara G, Galappaththy P, Constantine GR, Katulanda P. Medicinal properties of 'true' cinnamon (*Cinnamomum zeylanicum*): a systematic review. *BMC Complementary Medicine and Therapies* 2013; 13: 1-10. <https://doi.org/10.1186/1472-6882-13-275>
- Roussel A-M, Hininger I, Benaraba R, Ziegenfuss T N, Anderson R A. Antioxidant effects of a cinnamon extract in people with impaired fasting glucose that are overweight or obese. *Journal of the American College of Nutrition* 2009; 28: 16-21. <https://doi.org/10.1080/07315724.2009.10719756>
- Sahib A S. Anti-diabetic and antioxidant effect of cinnamon

- in poorly controlled type-2 diabetic Iraqi patients: A randomized, placebo-controlled clinical trial. *Journal of Intercultural Ethnopharmacology* 2016; 5: 108-113. <https://doi.org/10.5455/jice.20160217044511>
- Salamatian M, Mohammadi V, Abtahi Froushani S M. Ameliorative effects of aqueous cinnamon extract on ulcerative colitis in rats. *Physiology and Pharmacology* 2019; 23: 140-149.
- Schink A, Naumoska K, Kitanovski Z, Kampf C J, Fröhlich-Nowoisky J, Thines E, et al. Anti-inflammatory effects of cinnamon extract and identification of active compounds influencing the TLR2 and TLR4 signaling pathways. *Food & Function* 2018; 9: 5950-5964. <https://doi.org/10.1039/C8FO01286E>
- Seyed Ahmadi S G, Farahpour M R, Hamishehkar H. Topical application of Cinnamon verum essential oil accelerates infected wound healing process by increasing tissue antioxidant capacity and keratin biosynthesis. *Kaohsiung Journal of Medical Sciences* 2019; 35: 686-694. <https://doi.org/10.1002/kjm2.12120>
- Shalaby M A, Saifan H Y. Some pharmacological effects of cinnamon and ginger herbs in obese diabetic rats. *Journal of Intercultural Ethnopharmacology* 2014; 3: 144-149. <https://doi.org/10.5455/jice.20140818050741>
- Sharma P, Jha A B, Dubey R S, Pessarakli M. Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *Journal of Botany* 2012; 2012: 217037. <https://doi.org/10.1155/2012/217037>
- Somade O T, Ajayi B O, Adeyi O E, Aina B O, David B O, Sodiya I D. Activation of NF- $\kappa$ B mediates up-regulation of cerebellar and hypothalamic pro-inflammatory chemokines (RANTES and MCP-1) and cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-6) in acute edible camphor administration. *Scientific African* 2019; 5: e00114. <https://doi.org/10.1016/j.sciaf.2019.e00114>
- Taghizadeh M, Hamedifard Z, Jafarnejad S. Effect of garlic supplementation on serum C-reactive protein level: A systematic review and meta-analysis of randomized controlled trials. *Phytotherapy Research* 2019; 33: 243-252. <https://doi.org/10.1002/ptr.6225>
- Tanaka S, Yoon YH, Fukui H, Tabata M, Akira T, Okano K, et al. Antiulcerogenic compounds isolated from Chinese cinnamon *Planta Medica* 1989; 55: 245-248. <https://doi.org/10.1055/s-2006-961994>
- Taneja MK. Modified Bhramari Pranayama in Covid 19 Infection. *Indian Journal of Otolaryngology* 2020; 72: 395-397. <https://doi.org/10.1007/s12070-020-01883-0>
- Tulunay M, Aypak C, Yikilkan H, Gorpelioglu S. Herbal medicine use among patients with chronic diseases. *Journal of Intercultural Ethnopharmacology* 2015; 4: 217-220. <https://doi.org/10.5455/jice.20150623090040>
- Tung Y-T, Chua M-T, Wang S-Y, Chang S-T. Anti-inflammation activities of essential oil and its constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) twigs. *Bioresource Technology* 2008; 99: 3908-3913. <https://doi.org/10.1016/j.biortech.2007.07.050>
- Tuzcu Z, Orhan C, Sahin N, Juturu V, Sahin K. Cinnamon polyphenol extract inhibits hyperlipidemia and inflammation by modulation of transcription factors in high-fat diet-fed rats. *Oxidative Medicine and Cellular Longevity* 2017; 2017. <https://doi.org/10.1155/2017/1583098>
- Udayaprakash N K, Ranjithkumar M, Deepa S, Sripriya N, Al-Arfaj AA, Bhuvaneswari S. Antioxidant, free radical scavenging and GC-MS composition of *Cinnamomum iners* Reinw. ex Blume. *Industrial Crops and Products* 2015; 69: 175-179. <https://doi.org/10.1016/j.indcrop.2015.02.018>
- Vallianou N, Tsang C, Taghizadeh M, Davoodvandi A, Jafarnejad S. Effect of cinnamon (*Cinnamomum Zeylanicum*) supplementation on serum C-reactive protein concentrations: A meta-analysis and systematic review. *Complementary Therapies in Medicine* 2019; 42: 271-278. <https://doi.org/10.1016/j.ctim.2018.12.005>
- Vetal S, Bodhankar S L, Mohan V, Thakurdesai P A. Anti-inflammatory and anti-arthritis activity of type-A procyanidine polyphenols from bark of *Cinnamomum zeylanicum* in rats. *Food Science and Human Wellness* 2013; 2: 59-67. <https://doi.org/10.1016/j.fshw.2013.03.003>
- Wang S-Y, Chen P-F, Chang S-T. Antifungal activities of essential oils and their constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) leaves against wood decay fungi. *Bioresource Technology* 2005; 96: 813-818. <https://doi.org/10.1016/j.biortech.2004.07.010>
- Wang F, Pu C, Zhou P, Wang P, Liang D, Wang Q, et al. Cinnamaldehyde prevents endothelial dysfunction induced by high glucose by activating Nrf2. *Cellular Physiology and Biochemistry* 2015; 36: 315-324. <https://doi.org/10.1159/000374074>
- Weiss S R, Navas-Martin S. Coronavirus pathogenesis and the emerging pathogen severe acute respiratory syndrome coronavirus. *Microbiology and Molecular Biology Reviews* 2005; 69: 635-664. <https://doi.org/10.1128/MMBR.69.4.635-664.2005>
- Wondrak G T, Villeneuve N F, Lamore S D, Bause A S, Jiang T, Zhang D D. The cinnamon-derived dietary factor cinnamic

- aldehyde activates the Nrf2-dependent antioxidant response in human epithelial colon cells. *Molecules* 2010; 15: 3338-3355. <https://doi.org/10.3390/molecules15053338>
- Wong Y C, Ahmad-Mudzaqir M Y, Wan-Nurdiyana W A. Extraction of essential oil from cinnamon (*Cinnamomum zeylanicum*). *Oriental Journal of Chemistry* 2014; 30: 37-47. <https://doi.org/10.13005/ojc/300105>
- Woolbright B L, Jaeschke H. Mechanisms of inflammatory liver injury and drug-induced hepatotoxicity. *Current Pharmacology Reports* 2018; 4: 346-357. <https://doi.org/10.1007/s40495-018-0147-0>
- Wu C-T, Deng J-S, Huang W-C, Shieh P-C, Chung M-I, Huang G-J. Salvianolic acid C against acetaminophen-induced acute liver injury by attenuating inflammation, oxidative stress, and apoptosis through inhibition of the Keap1/Nrf2/HO-1 signaling. *Oxidative Medicine and Cellular Longevity* 2019; 2019. <https://doi.org/10.1155/2019/9056845>
- Yakhchali M, Taghipour Z, Mirabzadeh Ardakani M, Alizadeh Vaghasloo M, Vazirian M, Sadrai S. Cinnamon and its possible impact on COVID-19: The viewpoint of traditional and conventional medicine. *Biomedicine & Pharmacotherapy* 2021; 143: 112221-112231. <https://doi.org/10.1016/j.biopha.2021.112221>
- Youn H S, Lee J K, Choi Y J, Saitoh S I, Miyake K, Hwang D H, et al. Cinnamaldehyde suppresses toll-like receptor 4 activation mediated through the inhibition of receptor oligomerization. *Biomedicine & Pharmacotherapy* 2008; 75: 494-502. <https://doi.org/10.1016/j.bcp.2007.08.033>
- Yu H-S, Lee S-Y, Jang C-G. Involvement of 5-HT1A and GABAA receptors in the anxiolytic-like effects of *Cinnamomum cassia* in mice. *Pharmacology Biochemistry and Behavior* 2007; 87: 164-170. <https://doi.org/10.1016/j.pbb.2007.04.013>
- Yu T, Lee S, Yang W S, Jang H-J, Lee Y J, Kim T W, et al. The ability of an ethanol extract of *Cinnamomum cassia* to inhibit Src and spleen tyrosine kinase activity contributes to its anti-inflammatory action. *Journal of Ethnopharmacology* 2012; 139: 566-573. <https://doi.org/10.1016/j.jep.2011.11.051>
- Zhuang M, Jiang H, Suzuki Y, Li X, Xiao P, Tanaka T, et al. Procyanidins and butanol extract of *Cinnamomi Cortex* inhibit SARS-CoV infection. *Antiviral Research* 2009; 82: 73-81. <https://doi.org/10.1016/j.antiviral.2009.02.001>
- Zu Y, Yu H, Liang L, Fu Y, Efferth T, Liu X, et al. Activities of ten essential oils towards *Propionibacterium acnes* and PC-3, A-549 and MCF-7 cancer cells. *Molecules* 2010; 15: 3200-3210. <https://doi.org/10.3390/molecules15053200>